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Pablo Lemercier, Andrea Vergallo, Simone Lista, Henrik Zetterberg, Kaj Blennow, et al.. Association of plasma Aβ40/Aβ42 ratio and brain Aβ accumulation: testing a whole-brain PLS-VIP approach in individuals at risk of Alzheimer's disease. Neurobiology of Aging, 2021, 107, pp.57 - 69. 10.1016/j.neurobiolaging.2021.07.005 . hal-03391168

HAL Id: hal-03391168

https://hal.sorbonne-universite.fr/hal-03391168

Submitted on 21 Oct 2021

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Neurobiology of Aging 107 (2021) 57–69

Contents lists available at ScienceDirect

Neurobiology of Aging

journal homepage: www.elsevier.com/locate/neuaging.org

Association of plasma Aβ40/Aβ42 ratio and brain Aβ accumulation: testing a whole-brain PLS-VIP approach in individuals at risk of Alzheimer's disease

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ARTICLE INFO

Article history:
Received 17 March 2021
Revised 25 June 2021
Accepted 7 July 2021
Available online 14 July 2021

Keywords:
Subjective memory complaints
Preclinical Alzheimer's disease
Plasma amyloid β
Partial least square
whole-brain

ABSTRACT

Molecular and brain regional/network-wise pathophysiological changes at preclinical stages of Alzheimer's disease (AD) have primarily been found through knowledge-based studies conducted in late-stage mild cognitive impairment/dementia populations. However, such an approach may compromise the objective of identifying the earliest spatial-temporal pathophysiological processes. We investigated 261 individuals with subjective memory complaints, a condition at increased risk of AD, to test a whole-brain, non-a-priori method based on partial least squares in unraveling the association between plasma Aβ42/Aβ40 ratio and an extensive set of brain regions characterized through molecular imaging of Aβ accumulation and cortical metabolism. Significant associations were mapped onto large-scale networks, identified through an atlas and by knowledge, to elaborate on the reliability of the results. Plasma Aβ42/40 ratio was associated with Aβ-PET uptake (but not FDG-PET) in regions generally investigated in preclinical AD such as those belonging to the default mode network, but also in regions/networks normally not accounted - including the central executive and salience networks - which likely have a selective vulnerability to incipient Aβ accumulation.

The present whole-brain approach is promising to investigate early pathophysiological changes of AD to fully capture the complexity of the disease, which is essential to develop timely screening, detection, diagnostic, and therapeutic interventions.

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1. Introduction

Brain accumulation of amyloid-β (Aβ) aggregation species is one of the earliest pathophysiological alterations in the preclinical phase of the Alzheimer's disease (AD) clinical-biological continuum (Bateman et al., 2012; Jack et al., 2018; Villemagne et al.,

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https://doi.org/10.1016/j.neurobiolaging.2021.07.005

2013). From a therapeutic perspective, the early detection of incipient Aβ accumulation and downstream co-occurring alterations, such as spreading of tau pathology, synaptic failure, and neurodegeneration, holds the potential to effectively intervene using targeted disease-modifying therapies that can slow down AD disease progression during preclinical or prodromal stages (Aisen et al., 2017; Hampel et al., 2019a). By contrast, individuals at risk for AD or any other neurodegenerative disease may have different clinical-biological trajectories with some developing brain resilience, at the molecular and network level, while other individuals may develop multi-scale system failure and cognitive decline (Arenaza-Urquijo and Vemuri, 2018; Elman et al., 2014; Perez-Nievas et al., 2013). Therefore, reliable multi-modal exploratory and integrative approaches are needed for investigating network dysfunction in AD trajectories but also resilience in individuals displaying incipient molecular signatures of AD or any other neurodegenerative disease.

Positron emission tomography (PET) studies have shown that the regional Aβ accumulation in preclinical AD or individuals at genetic/clinical risk for AD is associated with different predictors, including bodily fluid biomarkers (McKhann et al., 2011). Coupling these findings with functional magnetic resonance imaging (MRI) studies and experimental models of AD revealed that a selective vulnerability of distinct brain regions to AD incipient pathophysiology exists. However, Aβ PET-based studies in preclinical AD typically employ a-priori, knowledge-based selected regions of interest (ROIs) such as the hippocampus, the anterior and posterior cingulate, or the precuneus (Fan et al., 2018; Fandos et al., 2017; Nakamura et al., 2018; Pérez-Grijalba et al., 2019; Vergallo et al., 2019). A limitation of this approach is represented by the fact that most of these regions have been identified in studies conducted in populations with more advanced stages of AD-like mild cognitive impairment (MCI) and dementia. Using approaches that constrain the number of hypotheses tested may hinder the comprehensive understanding of early AD pathophysiological dynamics. A model capable of accounting for a broad set of aging and AD-vulnerable regions is urgently needed to tackle incipient pathophysiological alterations timely.

Novel approaches that allow investigating more brain regions and different hubs of large-scale networks without a-priori constraints are needed in preclinical AD studies to develop timely screening, diagnostic, and therapeutic strategies. An ideal knowledge-free method should limit type 1 error inflation - induced by an increased number of hypotheses to test - and should deal with multicollinearity. This is of significant relevance since neuroimaging studies revealed significant pairwise correlations of AD hallmarks, like amyloid-β and also tau proteinopathies, between different regional values (Lockhart et al., 2017; Veronese et al., 2019).

In the present study, we investigated a non-a-priori design (i.e., hypothesis-free) based on Partial Least Square (PLS) analysis, a statistical method that allows an exploratory whole-brain approach. PLS estimates principal components, maximizing the covariance within and between 2 tables of the same set of observations to find shared information (Abdi and Williams, 2013). PLS is a component-based tool with several advantages compared to univariate methods, including higher suitability for modeling of datasets with considerable collinearity among variables. We employed the Variable Importance in Projection (VIP) criteria applied to PLS (PLS-VIP) to select the most relevant hypothesis. Indeed, simulation studies showed that PLS-VIP outperforms other variable selection methods and is less sensitive to noise and collinearity (Chong and Jun, 2005; Palermo et al., 2009).

We tested the PLS-VIP approach by investigating the association of plasma Aβ42/Aβ40 ratio, a validated biomarker for screening of AD pathophysiology (Jack et al., 2018; Nakamura et al., 2018; Palmqvist et al., 2019; Vergallo et al., 2019), with whole-brain Aβ-PET regional indexes. We used multi-modal biomarkers charting the Aβ pathway about which several studies have corroborated a significant covariance between different modality measures (i.e., blood concentrations and molecular imaging indexes). Such a confidence in the relationship between the 2 set of variables is pivotal for the scope of the present article, which consists in testing the reliability of a PLS-based methodological approach.

Previous studies in preclinical and prodromal AD showed good to optimal accuracy of plasma Aβ42/Aβ40 ratio in predicting Aβ-PET status (positive versus negative) (Nakamura et al., 2018; Palmqvist et al., 2019). Moreover, a correlation between the former and global/regional Aβ-PET standard uptake value ratios (SUVRs) has been reported across different AD cohorts (Fandos et al., 2017; Nakamura et al., 2018; Vergallo et al., 2019).

To test the conceptual validity of our model, we also tested plasma Aβ42/Aβ40 ratio association with ^{18}F-fluorodeoxyglucose (FDG) regional radiotracer binding, a marker of neuronal metabolism, assuming no relation would stand out as the existing literature suggests. We investigated this non-a-priori approach in a cohort of cognitively intact individuals facing subjective memory complaints (SMC), a clinical condition characterized by normal performance at a multi-domain neuropsychometric assessment despite a self-perceived memory impairment (Buckley et al., 2016; Teipel et al., 2020; Timmers et al., 2019; van Harten et al., 2018). Several multi-centric and multi-modal biomarkers studies indicate that SMC, coupled with positivity to Aβ biomarkers, is associated with increased risk of developing MCI or dementia within the clinical spectrum of AD (Buckley et al., 2016; Teipel et al., 2020; Timmers et al., 2019; van Harten et al., 2013). While this association is consistent across studies, partially controversial is the link between SMC and common risk factors for dementia – including vascular pathology and or tau pathophysiology (Clancy et al., 2021; Dubois et al., 2018; Van Etten et al., 2020).

We carried out our research proposal in the INSIGHT-preAD study cohort, a well-defined, large-scale, observational, monocentric, university-based longitudinal cohort of individuals with SMC and no significant medical comorbidities (see below for more details).

2. Materials and methods

2.1. Study participants

The study sample consisted of 318 participants with subjective memory complain (SMC), who were enrolled in the standardized, large-scale, observational, monocentric, French academic university-based “INveStIGation of AlzHeimer’s PredicTors in Subjective Memory Complainers” (INSIGHT-preAD) study (Dubois et al., 2018) – that is part of the Alzheimer Precision Medicine Initiative (APMI) and its established Cohort Program (APMI-CP) (Hampel et al., 2019b) . Participants were enrolled at the Institute of Memory and Alzheimer’s disease (Institut de la Mémoire et de la Maladie d’Alzheimer, IM2A) at the Pitié-Salpêtrière University Hospital in Paris, France. The main objective of the INSIGHT-preAD study is to explore the earliest preclinical stages of AD through intermediate to later stages until progression to conversion to first cognitive symptoms, using comprehensive clinical parameters and biomarkers associated with cognitive decline. In brief, the INSIGHT-preAD study includes 318 cognitively and physically healthy white (Caucasian) individuals, recruited from the community in the wider Paris area, France, aged 70 to 85, with SMC.

SMC was defined as a positive response (“YES”) to both of the following questions: “Are you complaining about your memory?”

and "Is it a regular complaint that has lasted for more than 6 months?" All participants were required to have an intact cognitive function – defined as a Mini-Mental State Examination score (MMSE) $\geq$ 27, a Clinical Dementia Rating scale (CDR) of 0, and a Free and Cued Selective Rating Test (FCSRT) total recall score $\geq$ 41.

Aβ-PET investigation was performed at the baseline visit, as a mandatory inclusion criterion. Thus, all subjects enrolled into the study have SMC and are stratified as either positive or negative for cerebral Aβ deposition. At the point of the study inclusion, several data were collected, namely demographic and clinical data, and Apolipoprotein E (APOE) genotype (see Supplementary materials for more details).

Exclusion criteria were a history of neurological or psychiatric diseases, including depressive disorders. Medical conditions, potential causing cognitive decline of non-AD biological nature, were ruled out at baseline (see Dubois et al., 2018 for more details). The study was conducted following the tenets of the Declaration of Helsinki of 1975 and approved by the local Institutional Review Board at the participating center. All participants or their representatives gave written informed consent to use their clinical data for research purposes.

2.2. Blood sample and plasma immunoassay

Ten (10) mL of venous blood was collected in 1 BD Vacutainer lithium heparin tube, which was used for all subsequent immunochemical analyses. Blood samples were taken in the morning, after a 12-hour fast, handled in a standardized way, and centrifuged for 15 minutes at 2000 G-force at 4°C. Per sample, plasma fraction was collected, homogenized, aliquoted into multiple 0.5 mL cryovial-sterilized tubes, and finally stored at 80°C within 2 hours from collection.

Analyses of plasma Aβ_{42} and Aβ_{40} concentrations were performed at the Clinical Neurochemistry Laboratory, Sahlgrenska University Hospital, Sweden. In particular, a volume of 0.5 mL of plasma for each subject was required for performing the analyses using the platforms mentioned above. Plasma Aβ1-42 and Aβ1-40 were analyzed using the Single-molecule array (Simoa) immunoassay (Quanterix, Billerica, Lexington, MA, USA). Regarding Aβ_{42}, repeatability was 4.1% and intermediate precision 7.0% for an internal QC plasma sample with a concentration of 10.5 pg/mL. Regarding Aβ_{40}, repeatability was 4.0% and intermediate precision 6.4% for an internal QC plasma sample with a concentration of 203 pg/mL.

2.3. PET scan acquisition and processing

Aβ and ^{18}F-FDG-PET investigations were performed at the baseline visit (M0) – as part of the inclusion criteria – and at 2-year follow-up (M24). Both scans were acquired in separate sessions, with a 48 hours interval.

Brain Aβ-PET scans were acquired 50 minutes after injection of 370 MBq (10 mCi) of ^{18}F-Florbetapir, which has high affinity for amyloid plaques. Brain ^{18}F-FDG scans were obtained 30 minutes after injection of 2 MBq/kg of 2-deoxy-2-(^{18}F)fluoro-D-glucose (^{18}F-FDG). All acquisitions were performed in a single session on a Philips Gemini GXL scanner and consisted of 3 $\times$ 5 minutes frames with a voxel size of 2 $\times$ 2 $\times$ 2 mm^3. Images were then reconstructed using an iterative LOR-RAMLA algorithm (10 iterations), with a "smooth" post-reconstruction filter. All corrections (attenuation, scatter and random coincidence) were integrated into the reconstruction. Lastly, frames were realigned, averaged, and quality-checked by the CATI team (*Centre d'Acquisition et Traitement des Images;* http://cati-neuroimaging.com). CATI is a French neuroimaging platform (available at http://cati-neuroimaging.com). PET images were then analyzed with an in-house pipeline developed by the CATI, including partial volume effect correction, as previously described (Dubois et al., 2018; Habert et al., 2018).

Finally, standard uptake value ratios (SUVR) were calculated using as reference regions a composite volume of interest (ROI) including pons and the whole cerebellum for Florbetapir images, whereas the pons for FDG images. SUVR were obtained in 84 cortical and neocortical ROIs extrapolated through Automated Anatomical Labeling (AAL) atlas. The global Aβ-PET SUVR was calculated as the average of the 84 cortical ROIs similarly as previously described in Farrell et al., 2018; Landau et al., 2010; Whittington et al., 2018.

3. Statistical analysis

3.1. Preprocessing strategy

A robust approach for variable selection, widely used in data science (Hastie et al., 2009; Xu and Goodacre, 2018), consists of dividing the dataset into a training set, used for selecting the variables, and a testing set, used to evaluate the selection. This latter provides an unbiased assessment of the variable selection procedure and, thus, enhances the confidence in the findings. To this end, we randomly split the dataset into a training and a testing dataset with a ratio of 1:1 stratifying by age, sex, *APOE* ε4 allele, education level, and total intracranial volume (TIV).

We reported descriptive statistics for the strata variables in the split datasets using the mean and standard deviation (SD) for quantitative variables and the frequency counts and percentages for categorical variables. We used Student's t-tests or chi-squared tests to evaluate the sampling homogeneity at a significance level of $p < 0.05$.

In the sequel, the training dataset was used for the PLS-VIP selection of the most relevant association between the regional PET signal and the plasma Aβ42/Aβ40 ratio, thus reducing the number of hypotheses. Then, the test data set was used to further validate the remaining assumptions.

3.2. Correlation between ROIs

First, we conducted correlation analyses on the training dataset to explore how imaging measurements were related across the different brain sub-regions according to the two PET techniques. To this end, for each pair of the 84 Aβ-PET (as well as FDG-PET) ROIs, we calculate the Pearson's correlations of the SUVR. Histograms and boxplots were build up to represent distributions of the pairwise correlation coefficients obtained with Aβ-PET and FDG-PET data.

3.3. Data modeling and feature selection

Associations between plasma Aβ42/Aβ40 ratio and either regional Aβ-PET SUVR or FDG-PET were determined through PLS models (Wold et al., 1993), on the training dataset (one model was fitted for each PET technique). Based on the NIPALS (Non-Linear iterative Partial Least Square) algorithm, PLS is a dimension reduction method, which is well-suited for managing high levels of correlations like, in the present study, between regional PET measures. In PLS, the X-matrix of imaging data can be reduced to a subset of orthogonal latent variables (or components), where each latent variable is constructed as a weighted sum of the X-variables, maximizing the covariance with the variable Y of Aβ42/Aβ40 ratios.

For feature selection, the method combining PLS and the variable importance in projection (VIP) scores (also called PLS-VIP method) have been introduced to detect the most influential predictors in explaining the variation of the Y-variable (Chong and Jun, 2005). Based on the PLS components, a VIP score

can be calculated for each sub-region to quantify its contribution to the variance explained by the component (Mehmood et al., 2012; Wold et al., 1993). Predictors with large VIP, larger than 1, are commonly the most relevant for explaining Y (Chong and Jun, 2005).

To achieve a stable selection, we used a bootstrap procedure to compute the VIP scores. Average VIP scores were thus calculated for each sub-region based on 5000 models obtained from bootstrap versions of the original dataset. Finally, the averaged VIP scores, sorted by descending values, were used to select and rank the most associated regions. For each PET technique, the PLS-VIP method was performed using the R package mixOmics (Chong and Jun, 2005; Rohart et al., 2017) specifying the "canonical mode" with 1 PLS component for the PLS model. To avoid confounding effects of age, sex, *APOE* ε4 carriers, and education level the training dataset were pre-processed prior to modeling with the ComBat method (Johnson et al., 2007) implemented in the R package sva available at https://bioconductor.org/packages.

McNemar chi-squared test was used to check for lateralized dominance between hemispheres for the selected regions.

3.4. Validation

The selected regions were further assessed for association with the plasma biomarkers using data of the testing set with no preprocessing. This was done through linear regression models (one model for each selected sub-region used as the dependent variable) to test the predicting effect of plasma Aβ42/Aβ40 ratio adjusted for the covariates age, sex, and *APOE* ε*4* genotype.

For each model, the Aβ42/Aβ40 ratio coefficient was reported with standard error (SE), and p-value. The Cohen's f^2 value was also calculated to indicate the effect size of Aβ42/Aβ40. Finally, all *p*-values obtained for Aβ42/Aβ40 were corrected using False Discovery Rate (FDR) for multiple testing, and its association with a sub-region was confirmed when the adjusted *p*-value was smaller than 0.05.

3.5. Internal reproducibility

As no independent cohort was available, we assessed the reproducibility of our method on data collected at months 12 (M12) and M24. We expected to find the same regions as previously identified. Besides, since some associations may vanish over time, it inquires sustainability of the identified associations during the subjects' follow-up period.

Thus, we replicated the same procedure described above for selecting and validating the regions at the longitudinal level with data at M12 and M24 with the same split samples used at M0. We tested Aβ accumulation and metabolism over a 1-year follow-up using plasma Aβ42/Aβ40 ratio measure at M12 and PET SUVRs at M24. Similarly, we tested Aβ accumulation and neuronal metabolism over a 2-year follow-up using plasma Aβ42/Aβ40 ratio measured at baseline and PET measured at M24.

3.6. Confounding analysis

As regional Aβ-PET SUVR and plasma Aβ42/Aβ40 can be influenced by the total amount of Aβ accumulation in patients, we conducted a confounding analysis to test whether the association between regional Aβ-PET SUVR and plasma Aβ42/Aβ40 can be explained by the brain overall amount of Aβ accumulation. This question was addressed at a cross-sectional level by using structural equation modeling (SEM) on the testing dataset. SEM is a convenient approach to model at the same time the relationships between regional Aβ and plasma Aβ42/Aβ40, between global Aβ and plasma Aβ42/Aβ40 and between regional and global Aβ.

All models (one model by sub-region), including the covariates age, sex and *APOE* ε*4* genotype, were built with the R package lavaan (Rosseel, 2012). We calculated p-values on all plasma Aβ42/Aβ40, global and regional Aβ-PET SUVR based on models fitted with 5000 bootstrap replicates of the original testing set. Finally, we corrected the *p*-values for controlling the FDR, and all associations remaining without the confounding effect of the total Aβ accumulation were established for all regions with an adjusted p-value < 0.05.

3.7. Interpretation of the PLS results relevance through knowledge-based functional networks

We performed a matching between the ROIs selected by the PLS and a knowledge-driven selection of large-scale networks, typically impaired in AD, to assist in the interpretation of the PLS-based results and gain more insights on whether the approach we employed may represent viable methodological solution for future clinical research studies in the field of aging and AD.

The definition of functional network components was made by grouping regions based on resting-state network atlas (Shirer et al., 2012), available at https://findlab.stanford.edu/functional_ROIs.html). We selected networks that have hypothesized functions in AD-related pathology: auditory network (AN), default mode network (DMN), central executive network (CEN), salience network (SaN), sensorimotor network (SeN), and visual network (VN). We matched the cortical regions of the AAL atlas and the resting-state network atlas based on our knowledge and visual inspection (see Supplementary Table S1 for more details).

Cohen's kappa (κ) with 95% confidence interval (95% CI) was used to assess agreement between selected ROIs and each brain large-scale functional network. Cohen's kappa is commonly used for quantifying agreement between two sets of binary classification tasks, considering that the agreement may occur by chance (Warrens, 2015).

All statistical analyses were performed with R software, version 3.6.0 (R Development Core Team, 2019) and plots were generated with the ggplot2 package (Wickham, 2009). Feature selection using PLS-VIP and confounding analyses were performed respectively with the R packages mixOmics (Rohart et al., 2017), available at https://bioconductor.org/packages, and lavaan (Rosseel, 2012), available at http://cran.r-project.org/web/packages. A script is supplied to allow the reader to apply our method (see Supplementary material).

4. Results

4.1. Datasets description

Analyses were performed on the 261 participants from the INSIGHT-preAD cohort who completed structural MRI, Aβ-PET, and FDG-PET acquisition at study inclusion.

Demographic characteristics, *APOE* ε4 genotype subgroups, and TIV are summarized for the training and the test datasets in Table 1. We did not find any significant difference between the two datasets regarding sex, *APOE* ε4 allele, age, education level, or TIV (p-values > 0.39). The distributions of the stratifying variables in the training and test sets are provided in the supplementary material (*Figure S1*). Because of the good accordance of data, we considered both datasets were homogeneous for the subject sampling.

Table 1
Description of training and test databases at baseline

		Training dataset N=134	Test dataset N=127	*p*-value
Age, mean ± SD		75.83 ± 3.23	75.81 ± 3.44	0.967
Sex, n (%)	Female	83 (61.9%)	77 (60.6%)	0.828
	Male	51 (38.1%)	50 (39.4%)	
APOE ε4 allele, n (%)	Carriers	30 (22.4%)	23 (18.1%)	0.391
	Non-carriers	104 (77.6%)	104 (81.9%)	
Education level, mean ± SD		6.29 ± 2.03	6.40 ± 1.90	0.651
Total intracranial volume, mean ± SD		1375 ± 136	1369 ± 127	0.705

Student and Chi-squared tests are used to calculate p-values
Abbreviations: APOE = apolipoprotein E, N = number of participants, % = percent of participants, and SD = Standard deviation

4.2. Regional Aβ accumulation

4.2.1. Cross-sectional study

Aβ-PET SUVR showed significant pairwise correlations with cortical regions (Fig. 1A). Indeed, the regional pairwise correlation ranged from 0.58 to 0.99 with a median of 0.87.

Out of the 84 cortical structures investigated with bootstrapped PLS-VIP, the averaged VIP scores highlighted a subset of 34 regions with a score above 1 (Table 2). Most of these regions were located in the frontal cortex, the cingulate cortex, the bilateral precuneus, and the right insula. We observed that the selected regions are mainly located in the left hemisphere (see Supplementary material for more details). From the testing dataset, we observed a significant negative association between Aβ42/Aβ40 ratio and all the regions previously selected (Table 2). These associations remained valid after corrections for multiple testing. The values of Cohen's f^2 ranging from 0.04 to 0.15 indicated a small to medium effect size of plasma Aβ42/Aβ40.

Interestingly, the confounding analysis highlighted the significant association of seven regions with the plasma Aβ42/Aβ40 ratio, which has been characterized after controlling for the confounding effect of the global Aβ-PET SUVR. These regions concerned the bilateral medial cingulate, the left posterior cingulate, the right inferior frontal orbital, the right frontal superior medial, the left superior parietal, and the left precuneus (*Table S1*).

4.2.2. Longitudinal study

The 1-year follow-up was assessed using Aβ42/Aβ40 ratio at M12 and Aβ-PET imaging at M24. Bootstrapped PLS-VIP analysis selected 25 of the 84 regions (Table 3). These regions were mainly located in the frontal cortex and the cingulate cortex, including the bilateral insula. Likewise, the 2-year follow-up investigation on Aβ42/Aβ40 ratio at M0 and Aβ-PET imaging at M24 selected 19 of the 84 regions (Table 4) mainly located in the frontal cortex and the cingulate cortex including right insula. Results showed a left hemisphere dominance for the 1-year follow-up analysis but not the 2-year follow-up (see Supplementary material for more details).

On the testing dataset, we observed a significant negative association between Aβ42/Aβ40 ratio and amyloid-PET in all the regions selected over a 1-year follow-up (Table 3) and a 2-year follow-up (Table 4). Besides, all of these results survived after FDR correction. Cohen's f^2 values revealed a small to medium effect size of plasma Aβ42/Aβ40 over a 1-year follow-up and a small effect size of plasma Aβ42/Aβ40 over a 2-year follow-up.

4.3. Regional cortical metabolism

4.3.1. Cross-sectional study

Measures of cortical FDG-PET showed signifcant pairwise correlations with cortical regions (Fig. 1B). The minimum regional pairwise correlation was 0.50, the maximum was 0.98, and the median 0.82.

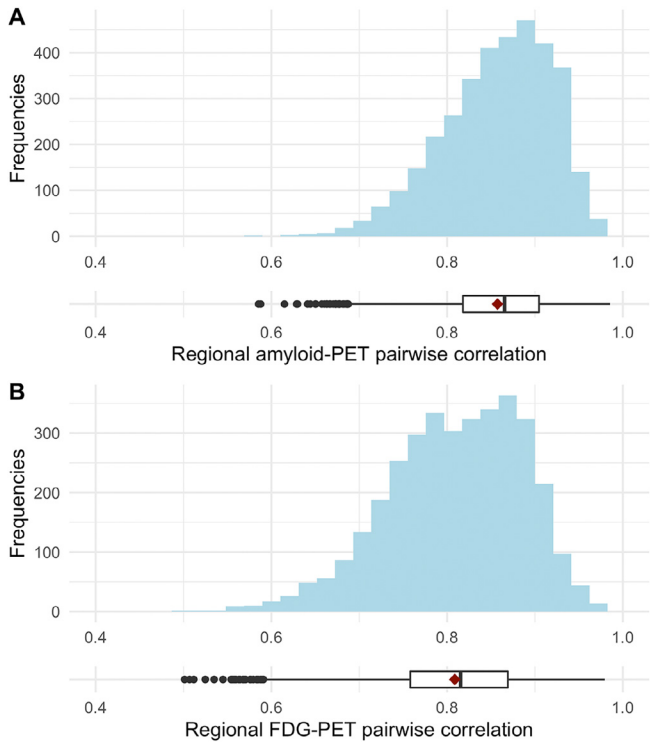


Fig. 1. Regional pairwise correlation.

Bootstrapped PLS-VIP selected 16 regions over 84 (Table 5). These regions were mainly located in the temporal cortex. Investigation on the testing dataset showed that Aβ42/Aβ40 ratio was not significantly associated with FDG-PET measure in any of these regions (Table 5).

4.3.2. Longitudinal study

Bootstrapped PLS-VIP investigation on FDG-PET imaging at M24 identified 21 regions associated with baseline Aβ42/Aβ40 ratio and 16 regions with M12 Aβ42/Aβ40 ratio. Investigation on the testing dataset showed that Aβ42/Aβ40 ratio was not significantly associated with regional FDG-PET measure over a 1-year (*Table S3*) or a 2-year follow-up (*Table S4*).

4.4. Selected regions and functional networks

Here we investigated whether selected Aβ-PET regions at different follow-up visits were involved in specific resting state networks (see Supplementary Table S1 for more details). At baseline, the Aβ-PET regional subset predominantly overlapped the

Table 2
Association between Aβ42/Aβ40 ratio and amyloid-PET at baseline

Region	VIP score	Regression coefficient (SE)	*p*-value	Adjusted p-value	Cohen's f^2
Angular left	1.04	-3.46 (1.25)	$6.35e^{-3}$	$6.99e^{-3}$	0.10
Angular right	1.21	-4.40 (1.24)	$5.80e^{-4}$	$1.46e^{-3}$	0.12
Calcarine left	1.04	-2.84 (0.98)	$4.44e^{-3}$	$5.59e^{-3}$	0.04
Cingulate Ant left	1.64	-4.32 (1.17)	$3.47e^{-4}$	$1.46e^{-3}$	0.11
Cingulate Ant right	1.29	-5.03 (1.20)	$5.10e^{-5}$	$9.20e^{-4}$	0.12
Cingulate Mid left	1.39	-3.89 (1.19)	$1.41e^{-3}$	$2.09e^{-3}$	0.10
Cingulate Mid right	1.41	-4.13 (1.14)	$4.43e^{-4}$	$1.46e^{-3}$	0.12
Cingulate Post left	1.27	-4.56 (1.29)	$6.02e^{-4}$	$1.46e^{-3}$	0.11
Cuneus left	1.10	-2.90 (1.01)	$4.87e^{-3}$	$5.92e^{-3}$	0.04
Frontal Inf Oper left	1.18	-3.29 (1.01)	$1.52e^{-3}$	$2.15e^{-3}$	0.10
Frontal Inf Orb 2 left	1.21	-3.85 (1.14)	$9.83e^{-4}$	$1.76e^{-3}$	0.06
Frontal Inf Orb 2 right	1.29	-3.71 (1.11)	$1.05e^{-3}$	$1.76e^{-3}$	0.13
Frontal Inf Tri left	1.16	-3.71 (1.11)	$1.09e^{-3}$	$1.76e^{-3}$	0.09
Frontal Med Orb left	1.57	-4.91 (1.33)	$3.27e^{-4}$	$1.46e^{-3}$	0.14
Frontal Med Orb right	1.26	-5.35 (1.28)	$5.40e^{-5}$	$9.20e^{-4}$	0.15
Frontal Mid 2 left	1.24	-4.28 (1.21)	$5.62e^{-4}$	$1.46e^{-3}$	0.11
Frontal Mid 2 right	1.35	-4.54 (1.21)	$2.85e^{-4}$	$1.46e^{-3}$	0.13
Frontal Sup 2 left	1.19	-4.30 (1.20)	$5.05e^{-4}$	$1.46e^{-3}$	0.10
Frontal Sup 2 right	1.20	-4.69 (1.18)	$1.26e^{-4}$	$1.27e^{-3}$	0.12
Frontal Sup Medial left	1.33	-4.20 (1.23)	$8.68e^{-4}$	$1.74e^{-3}$	0.12
Frontal Sup Medial right	1.21	-5.07 (1.29)	$1.49e^{-4}$	$1.27e^{-3}$	0.12
Insula right	1.06	-3.54 (0.97)	$3.92e^{-4}$	$1.46e^{-3}$	0.10
OFClat left	1.08	-4.26 (1.40)	$2.96e^{-3}$	$3.87e^{-3}$	0.07
OFCpost right	1.10	-3.04 (1.15)	$8.99e^{-3}$	$9.56e^{-3}$	0.12
Olfactory left	1.34	-2.46 (1.01)	$1.63e^{-2}$	$1.68e^{-2}$	0.11
Parietal Inf left	1.03	-3.34 (1.20)	$6.37e^{-3}$	$6.99e^{-3}$	0.12
Parietal Sup left	1.05	-3.34 (1.38)	$1.71e^{-2}$	$1.71e^{-2}$	0.09
Precuneus left	1.35	-4.00 (1.42)	$5.80e^{-3}$	$6.80e^{-3}$	0.08
Precuneus right	1.37	-4.25 (1.29)	$1.32e^{-3}$	$2.04e^{-3}$	0.10
Rectus left	1.43	-4.32 (1.26)	$8.00e^{-4}$	$1.71e^{-3}$	0.12
Rectus right	1.26	-4.49 (1.19)	$2.40e^{-4}$	$1.46e^{-3}$	0.13
SupraMarginal right	1.09	-3.78 (1.10)	$8.04e^{-4}$	$1.71e^{-3}$	0.10
Temporal Mid left	1.09	-3.13 (1.03)	$2.82e^{-3}$	$3.83e^{-3}$	0.10
Temporal Sup left	1.12	-2.98 (0.89)	$1.03e^{-3}$	$1.76e^{-3}$	0.09

VIP scores derive from the feature selection with PLS-VIP, while regression coefficient, *p*-value, adjusted p-value, and Cohen's f^2 derived from the validation with linear models.
All models are adjusted for age, sex, and *APOE ε4* genotype. p-values refer to the test of regression coefficient. Adjusted p-values are calculated using false discovery rate correction.
Abbreviations: SE = Standard error and VIP = Variable Importance in Projection

Table 3
Association between Aβ42/Aβ40 ratio at M12 and amyloid-PET at M24

Region	VIP score	Regression coefficient (SE)	*p*-value	Adjusted *p*-value	Cohen's f^2
Cingulate Ant left	1.39	-5.12 (1.34)	$2.46e^{-4}$	$1.88e^{-3}$	0.17
Cingulate Ant right	1.38	-5.25 (1.33)	$1.47e^{-4}$	$1.88e^{-3}$	0.18
Frontal Inf Oper left	1.22	-3.21 (1.29)	$1.43e^{-2}$	$1.49e^{-2}$	0.10
Frontal Inf Oper right	1.13	-3.80 (1.25)	$3.09e^{-3}$	$4.19e^{-3}$	0.14
Frontal Inf Orb 2 left	1.02	-3.66 (1.32)	$6.75e^{-3}$	$7.67e^{-3}$	0.12
Frontal Inf Tri left	1.43	-3.69 (1.39)	$9.21e^{-3}$	$1.00e^{-2}$	0.10
Frontal Inf Tri right	1.40	-4.22 (1.37)	$2.69e^{-3}$	$3.95e^{-3}$	0.15
Frontal Med Orb left	1.32	-5.68 (1.51)	$3.00e^{-4}$	$1.88e^{-3}$	0.18
Frontal Med Orb right	1.05	-5.62 (1.49)	$2.89e^{-4}$	$1.88e^{-3}$	0.19
Frontal Mid 2 left	1.22	-4.20 (1.47)	$5.07e^{-3}$	$6.04e^{-3}$	0.12
Frontal Mid 2 right	1.29	-5.16 (1.49)	$7.85e^{-4}$	$2.45e^{-3}$	0.18
Frontal Sup 2 left	1.28	-4.47 (1.44)	$2.45e^{-3}$	$3.87e^{-3}$	0.13
Frontal Sup 2 right	1.23	-5.12 (1.43)	$5.27e^{-4}$	$2.20e^{-3}$	0.18
Frontal Sup Medial left	1.20	-4.71 (1.48)	$1.93e^{-3}$	$3.70e^{-3}$	0.13
Frontal Sup Medial right	1.25	-5.30 (1.50)	$6.61e^{-4}$	$2.36e^{-3}$	0.17
Insula left	1.06	-3.69 (1.17)	$2.08e^{-3}$	$3.71e^{-3}$	0.13
Insula right	1.11	-3.74 (1.20)	$2.48e^{-3}$	$3.87e^{-3}$	0.13
OFCmed left	1.34	-4.72 (1.44)	$1.41e^{-3}$	$3.20e^{-3}$	0.15
OFCpost left	1.03	-3.75 (1.28)	$4.38e^{-3}$	$5.48e^{-3}$	0.14
Rectus left	1.10	-5.30 (1.46)	$4.40e^{-4}$	$2.20e^{-3}$	0.18
Rectus right	1.26	-4.47 (1.38)	$1.65e^{-3}$	$3.44e^{-3}$	0.18
Temporal Mid left	1.10	-4.31 (1.29)	$1.21e^{-3}$	$3.03e^{-3}$	0.15
Temporal Pole Sup left	1.37	-2.95 (0.97)	$3.19e^{-3}$	$4.19e^{-3}$	0.18
Temporal Pole Sup right	1.08	-2.59 (1.07)	$1.70e^{-2}$	$1.70e^{-2}$	0.17
Temporal Sup left	1.07	-3.86 (1.15)	$1.15e^{-3}$	$3.03e^{-3}$	0.15

VIP scores derive from the feature selection with PLS-VIP, while regression coefficient, p-value, adjusted p-value, and Cohen's f^2 derived from the validation with linear models.
All models are adjusted for age, sex, and *APOE ε4* genotype. p-values refer to the test of regression coefficient. Adjusted p-values are calculated using false discovery rate correction.
Abbreviations: SE = Standard error and VIP = Variable Importance in Projection

Table 4
Association between baseline Ab42/Ab40 ratio and amyloid-PET at M24

Region	VIP score	Regression coefficient (SE)	*p*-value	Adjusted *p*-value	Cohen's f^2
Angular right	1.03	-2.82 (0.79)	0.001	0.004	0.08
Cingulate Ant left	1.20	-2.28 (0.72)	0.002	0.004	0.04
Cingulate Ant right	1.40	-2.24 (0.71)	0.002	0.004	0.05
Cingulate Mid right	1.01	-2.29 (0.68)	0.001	0.004	0.06
Frontal Inf Oper left	1.03	-1.96 (0.68)	0.005	0.005	0.04
Frontal Inf Oper right	1.16	-2.16 (0.65)	0.001	0.004	0.05
Frontal Inf Tri left	1.34	-2.02 (0.73)	0.007	0.007	0.04
Frontal Inf Tri right	1.34	-2.21 (0.72)	0.003	0.004	0.07
Frontal Mid 2 left	1.22	-2.19 (0.76)	0.005	0.005	0.04
Frontal Mid 2 right	1.43	-2.46 (0.76)	0.001	0.004	0.06
Frontal Sup 2 left	1.20	-2.26 (0.74)	0.003	0.004	0.04
Frontal Sup 2 right	1.34	-2.42 (0.72)	0.001	0.004	0.06
Frontal Sup Medial left	1.15	-2.55 (0.75)	0.001	0.004	0.03
Frontal Sup Medial right	1.10	-2.56 (0.76)	0.001	0.004	0.05
Insula right	1.08	-1.90 (0.62)	0.003	0.004	0.04
Occipital Inf left	1.03	-2.23 (0.79)	0.006	0.006	0.06
OFCpost left	1.05	-2.06 (0.68)	0.003	0.004	0.05
Rectus right	1.01	-2.22 (0.72)	0.003	0.004	0.07
Temporal Pole Sup left	1.39	-1.60 (0.52)	0.003	0.004	0.07

VIP scores derive from the feature selection with PLS-VIP, while regression coefficient, p-value, adjusted p-value, and Cohen's f^2 derived from the validation with linear models.
All models are adjusted for age, sex, and *APOE ε4* genotype. p-values refer to the test of regression coefficient. Adjusted p-values are calculated using false discovery rate correction.
Abbreviations: SE = Standard error and VIP = Variable Importance in Projection

Table 5
Association between Aβ42/Aβ40 ratio and FDG-PET at baseline

Region	VIP score	Regression coefficient (SE)	*p*-value	Adjusted *p*-value	Cohen's f^2
Cingulate Post left	1.97	2.75 (2.62)	0.296	0.804	0.05
Cingulate Post right	2.27	1.52 (2.44)	0.533	0.804	0.07
Cuneus left	1.19	3.57 (2.54)	0.162	0.804	< 0.01
Frontal Med Orb left	1.42	1.10 (2.44)	0.653	0.804	0.01
Heschl right	1.59	-0.54 (2.70)	0.843	0.865	0.01
Hippocampus left	1.29	0.67 (1.03)	0.520	0.804	0.04
Hippocampus right	1.86	0.67 (0.99)	0.503	0.804	0.02
Lingual right	1.62	2.45 (2.17)	0.261	0.804	0.01
Occipital Inf right	1.78	2.89 (2.78)	0.301	0.804	0.01
Occipital Sup left	1.61	1.75 (2.83)	0.538	0.804	0.01
Olfactory right	1.70	0.39 (1.42)	0.783	0.865	0.02
Paracentral Lobule left	1.64	1.63 (2.30)	0.479	0.804	< 0.01
ParaHippocampal left	1.69	0.78 (1.01)	0.444	0.804	0.03
ParaHippocampal right	1.32	0.55 (1.08)	0.609	0.804	0.02
Temporal Pole Mid left	1.48	0.78 (1.43)	0.586	0.804	0.01
Temporal Pole Sup left	1.59	0.26 (1.55)	0.865	0.865	< 0.01

VIP scores derive from the feature selection with PLS-VIP, while regression coefficient, p-value, adjusted p-value, and Cohen's f^2 derived from the validation with linear models.
All models are adjusted for age, sex, and *APOE ε4* genotype. *p*-values refer to the test of regression coefficient. Adjusted *p*-values are calculated using false discovery rate correction.
Abbreviations: SE = Standard error and VIP = Variable Importance in Projection

CEN (κ = 0.46; 95% CI = [0.27–0.65]) and the DMN (κ = 0.30; 95% CI = [0.09–0.51]) (Fig. 2A). The amyloid-PET regional subset selected at 1-year follow-up, predominantly overlapped the CEN (κ = 0.37; 95% CI = [0.16–0.59]) and the SaN (κ = 0.26; 95% CI = [0.05–0.46]) (Fig. 2B). Likewise, the two-year follow-up amyloid-PET regional subset predominantly overlapped the CEN (κ = 0.27; 95% CI = [0.04–0.49]) and the SaN (κ = 0.27; 95% CI = [0.03–0.5]) (Fig. 2C).

5. Discussion

This study probes the potential clinical value of a whole-brain, non-a-priori approach based on PLS-VIP by investigating the association between plasma levels of Aβ42/Aβ40 ratio and an extensive set of regions characterized for Aβ accumulation and cortical metabolism.

We used multi-modal biomarkers charting the Aβ pathway, which is a valuable tool to explore our approach since several studies have corroborated a significant covariance between different modality measures (i.e., blood concentrations and molecular imaging indexes).

5.1. The PLS-VIP feature selection

PLS-VIP method allows comprehensive exploration of brain single ROIs and hubs of large-scale networks that are likely to be involved in preclinical/early prodromal stages of AD. Most of the conventional study designs using knowledge-based a priori selection of ROIs may fail to capture several particularly vulnerable regions biologically connected to incipient AD pathophysiological events, including early stages of proteinopathies.

To probe our approach, we used data generated in a population of cognitively healthy individuals with SMC, a clinical condition as-

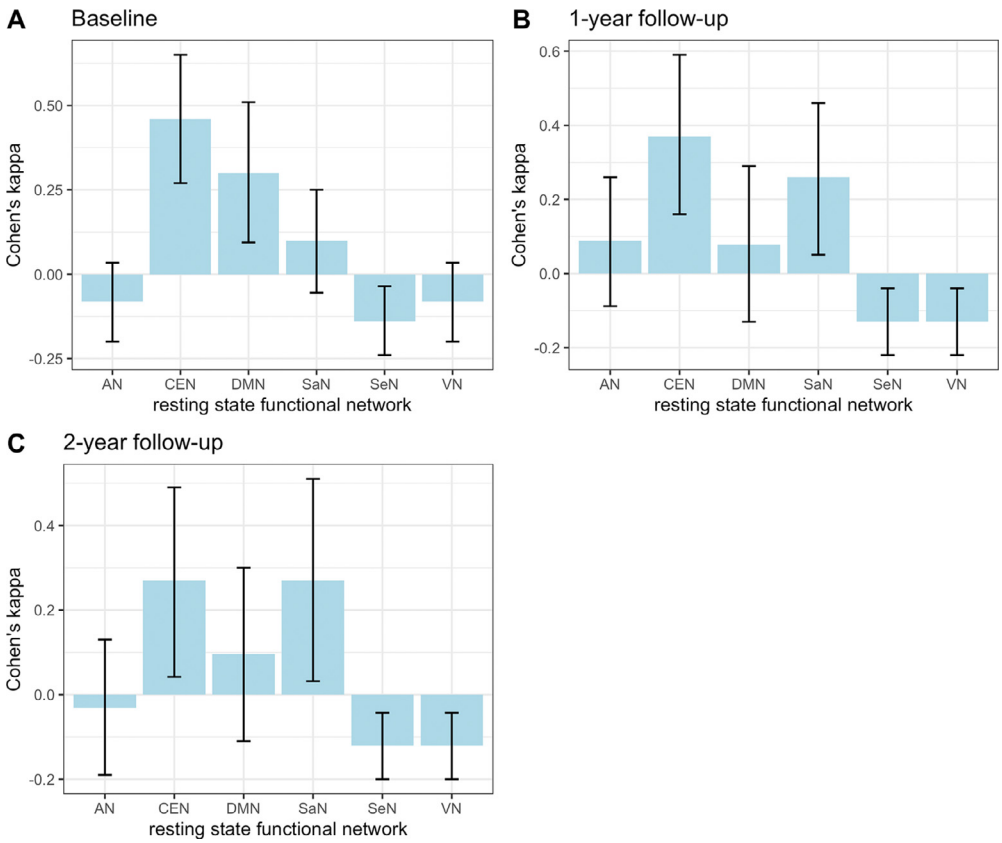


Fig. 2. Cohen's Kappa between resting state network and amyloid-PET selected regions Bar represent 95% confidence intervalAbbreviations: AN = auditory network, CEN = central executive network, DMN = default mode network, SaN = salience network, SeN = sensorimotor network, and VN = visual network.

sociated with increased risk of AD (Buckley et al., 2016; van Harten et al., 2018). At baseline, we found that plasma Aβ42/Aβ40 ratio was negatively associated with Aβ-PET indexes in the frontal cortex, the cingulate cortex, the precuneus, and the insula. The solutions delivered by PLS-VIP were consistent when the analysis was reproduced on the same cohort at different timepoints, indicating a robust and stable approach.

In the present study we confirmed the previously reported association between fluid biological signatures of the Aβ pathway and Aβ-PET signal in regions typically picked by a-priory study designs, such as the hippocampus, the anterior and posterior cingulate, or the precuneus (Fandos et al., 2017; Nakamura et al., 2018; Pérez-Grijalba et al., 2019). Moreover, we also identify additional regions vulnerable to AD pathophysiology, including the insula, the angular gyrus, and frontal cortex, as pointed out by previous ROI-based studies (Fan et al., 2018; Vergallo et al., 2019).

5.2. A preliminary selected ROI – knowledge-based network matching

Finally, we conducted a stepwise knowledge-driven process to check whether the identified Aβ-PET ROIs, including those who remained after controlling for global SUVR, matched with specific brain large-scale networks. We observed that the majority of regions whose Aβ accumulation rates correlate with plasma Aβ42/Aβ40 ratio belonged to distinct macroscale networks either typically investigated in AD (DMN) or usually not taken into account for association studies (salience network (SaN), central executive network (CEN)), although their decline over age has been extensively reported (Agosta et al., 2012; Badhwar et al., 2017; Chiesa et al., 2020; Hampel et al., 2019a; Zhao et al., 2019).

There is consolidated evidence about the DMN functional connectivity decline and its association with AD pathophysiological hallmarks during the disease's early stages (Badhwar et al., 2017; Bero et al., 2011; Palmqvist et al., 2017; Teipel et al., 2016).

Functional MRI in-human studies indicate that there is spatial-temporal overlap between DMN activity decay and deposition Aβ and tau (Hampel et al., 2019a; Li et al., 2019; Mormino et al., 2011) and that a decreased functional connectivity in the DMN is associated with neurodegeneration (Chhatwal et al., 2018; Palmqvist et al., 2017), cortical shrinking (Hampton et al., 2020) and worse cognitive trajectories in individuals displaying elevated Aβ burden (Buckley et al., 2017).

Spatial covariance between Aβ accumulation and connectivity and metabolism in the CEN (decreased) has been reported in aging and AD individuals (Andrews-Hanna et al., 2007; Grothe and Teipel, 2016; Palmqvist et al., 2017). A decreased functional connectivity has been reported within the SaN in aged individuals and patients suffering from early AD in which networks breakdown take place at different temporal coordinates (Brier et al., 2012; He et al., 2014; Zhou et al., 2010). Therefore, our results are in line with previous evidence indicating that distinct brain regions may have a higher intrinsic vulnerability to AD early pathophysiological alterations (Bero et al., 2011; Chiesa et al., 2019; Crossley et al., 2014; Hampel et al., 2019a).

5.3. Potential utilization of PLS-VIP approch in resilience studies

We also found a discrepancy between time points, with DMN and SaN being associated at baseline but not over time. A clinical explanation for these findings lies in the general and study-wise characteristic of our cohort.

Although SMC is a condition itself associated with increased risk for developing AD cognitive decline, the biology underlining the SMC clinical label, for instance, presence or not of brain Aβ accumulation, is heterogeneous and longitudinal trajectories of SMC individuals may be different with some having or developing brain resilience, at the molecular and network level, and some other going toward multi-scale system failure and cognitive decline (Dubois et al., 2018; Hohman et al., 2016; Negash et al., 2013).

In the present study, only 4 individuals - out of 261 - developed objective cognitive decline (MCI or dementia) within the 3-year follow-up (all of them have positive Aβ-PET). In line with previous INSIGHT-preAD publications, we argue that such rates of cognitive decline, lower than expected in a population at risk for AD [i.e., SMC plus signs of Aβ accumulation] is partially explained by the average high education level and cognitive reserve of the INSIGHT-preAD participants, a cohort in which potential compensatory mechanisms have already been reported (Babiloni et al., 2020; Dubois et al., 2018; Gaubert et al., 2019). This cohort is mainly represented by people who still engage in small post-retirement works, continuous social and intellectual activity (Babiloni et al., 2020; Cacciamani et al., 2020; Dubois et al., 2018).

In summary, our DMN-related divergent results and believe that this evidence-based conceptual construct can also explain the apparent DMN-related inconsistent association over time-points in our study. We have based our hypothesis on the presence of cognitive reserve-related resiliency dynamics in our cohort dataset by capitalizing on extensive evidence and data-driven conceptual frameworks previously published (Babiloni et al., 2020; Dubois et al., 2018; Gaubert et al., 2019; Hohman et al., 2016; Negash et al., 2013).

In this conceptual framework lies the potential clinical value of our proposed study design. In fact, the PLS-based whole-brain approach could be used not only for investigating network dysfunction in AD trajectories but also resilience in individuals displaying incipient molecular signatures of AD or any other neurodegenerative disease.

Such speculation connects to our intention of carrying out a non-a-priori approach, like PLS-VIP, to facilitate the investigation of a broader set of brain functional networks and their association with AD pathophysiology, thus, enabling the identification of other network-network connection patterns involved in disease progression or compensation.

5.4. No association between glucose metabolism and Aβ accumulation according to the PLS analysis

Our FDG-PET based results suggest that plasma Aβ42/Aβ40 ratio is not a suitable marker of neuronal metabolism that has been reported as a potential surrogate marker of neuronal loss (Dubois et al., 2014). To our knowledge, only 1 study assessed the association between Aβ42/Aβ40 ratio and FDG-PET in MCI/dementia pooled cohort (Pérez-Grijalba et al., 2019). By contrast, no results have been reported on cognitively healthy normal individuals at clinical and or biological risk for AD. Therefore, an interpretation of our data in light of the existing literature is hard to perform. However, it is conceivable to argue that the lack of FDG-PET signal with plasma Aβ42/Aβ40 ratio indicates that the latter is not a suitable marker for either neurodegeneration or disease progression.

5.5. The PLS-VIP method main advantages for AD clinical research

We believe that our workplan is easily replicable and shows good intrinsic operability. Of note, we used a set of variables with known covariance to test the performance of the modeling approach. The true potential of our approach is for exploratory purposes besides validation studies. PLS can be considered as a generalization of multiple regression since, for identical problem formulations, they both produce similar answers (Cramer, 1993). Unlike multiple regression, PLS can handle extensive datasets that may contain more predictors than subjects (Cramer, 1993; Krishnan et al., 2011).

In addition, PLS is a very versatile method that provides relevant tools for different settings of clinical investigation. For instance, in AD clinical research, PLS was used for diagnostic classification tasks (Aguilar et al., 2014), omics studies (Lorenzi et al., 2018; Vardarajan et al., 2020; Xicota et al., 2019) and early longitudinal cognitive decline (Langbaum et al., 2020). The diversity of applications shows that PLS is a suitable tool for both hypothesis testing and explanatory model building. Likewise, PLS-VIP was used in AD studies to select variables predicting brain volume changes (Thambisetty et al., 2011) or associated with AD biomarkers (Baldacci et al., 2020). PLS-VIP approach is not sensitive to multicollinearity among predictors, which is a remarkable advantage compared to conventional feature selection method (Chong and Jun, 2005; Cramer, 1993; Palermo et al., 2009).

The Least Absolute Shrinkage and Selection Operator (LASSO) regression is another method previously used in AD clinical research for feature selection able to handle correlated variables (Kohannim, 2012; Li et al., 2018; Yang et al., 2015) (Dayon et al., 2018). However, LASSO presents significant inconsistency among the variable selection (Zou and Hastie, 2005) and constraints all correlated variables but one to have null coefficients i.e., the method selects only one feature and drops the others (Desboulets, 2018; Hastie et al., 2009). Elastic net regularization is a good alternative to LASSO that provides better performance and stability in variable selection (Zou and Hastie, 2005). However, with the elastic net, the coefficients' weight is distributed among the correlated variables, so when we interpret the model, the strength of the association is underrated. The most relevant advantage of PLS compred to other methods is to keep all relevant associations, providing a complete overview of the process of interest, which is appropriate for explicative models and critical for interpreting the early pathophysiological dynamics process.

As normally done in machine learning approaches, we split the dataset to increase the confidence in the significant findings. By contrast, such a step may reduce statistic power, thus hindering the detection of some subtle albeit clinically meaningfull associations.

5.6. Study limitations

We acknowledge that different atlases, including cytoarchitectonic and probabilistic maps, have been published lately and some of them are expected to perform better, especially for those regions with no clear macro-anatomical landmarks. This methodological limitation explains why we did not include subcortical regions in the present study.

The present study does not include a long-term neuropsychological clinical follow-up of participants, which represents a limitation. Moreover, although education is widely considered a component of the cognitive reserve we thought not to overload our PLS-based approach and region models with too many covariates, considering that the dataset is not large enough.

To expand on the clinical potential of our approach, we set out to extend the multi-modal biomarker and neuropsychological follow-up of the individuals involved in the present study. In addition, we aim at expanding the dataset through a multicentric study where we would use PLS to explore trajectories of decline versus resilience, also keeping education levels into model.

This future study would include an independent validation cohort, the lack of which represents caveat itself. To compensate this issue, we carried out several additional analytical steps (training/test datasets, bootstrap, correction for multiple tests, and internal reproduction at different timepoints) to increase confidence in the findings. We also acknowledge the lack of a test cohort of MCI and dementia individuals with AD pathophysiology as a major caveat of our study since it may have supported data interpretation from a biological standpoint and to draw some clinical conclusions. In this sense, we set out to perform a large-scale longitudinal multicenter study including at least one discovery and one independent validation cohort spanning the entire AD continuum, and including SMC and MCI either converters or stable. We believe that this follow-up study is essential for the standardization and harmonization of our proposed analytical protocol, and at the same time, to investigate whether our whole-brain approach may generate reliable predictive measures to employ in clinical trials and evolving healthcare practice.

6. Conclusions

Our study shows that plasma Aβ42/Aβ40 ratio is negatively associated with several regional Aβ-PET indexes at a cross-sectional and longitudinal level in SMC individuals. Leveraging the PLS-VIP computational power and using a non-a-priori strategy (i.e., hypothesis-free), we find several associations between plasma Aβ42/Aβ40 ratio and brain regions belonging to multiple large-scale functional networks, including networks normally not investigated in a-priori study designs for preclinical AD.

Of note, we do not suggest to set aside a priori hypothesis investigations but instead promote an integrated approach based on the notion that a-priori and non-a-priori investigations can be complementary and provide partially different information for clinical research. We propose that non-a-priori investigations should be employed in biomarker-guided exploratory studies conducted in preclinical populations to facilitate the understanding of early pathophysiological trajectories of neurodegenerative diseases and to indentify vulnerable regions that may deserve specific attention for preventive strategies.

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Disclosure

HH is an employee of Eisai Inc. eclares no competing financial interests related to the present article and his contribution to this article reflects entirely and only his own academic expertise on the matter. HH serves as Senior Associate Editor for the Journal Alzheimer's & Dementia and does not receive any fees or honoraria since May 2019; before May 2019 he had received lecture fees from Servier, Biogen and Roche, research grants from Pfizer, Avid, and MSD Avenir (paid to the institution), travel funding from Eisai, Functional Neuromodulation, Axovant, Eli Lilly and company, Takeda and Zinfandel, GE-Healthcare and Oryzon Genomics, consultancy fees from Qynapse, Jung Diagnostics, Cytox Ltd., Axovant, Anavex, Takeda and Zinfandel, GE Healthcare, Oryzon Genomics, and Functional Neuromodulation, and participated in scientific advisory boards of Functional Neuromodulation, Axovant, Eisai, Eli

Lilly and company, Cytox Ltd., GE Healthcare, Takeda and Zinfandel, Oryzon Genomics and Roche Diagnostics. He is inventor of the following patents as a scientific expert and has received no royalties:

- *In Vitro* Multiparameter Determination Method for The Diagnosis and Early Diagnosis of Neurodegenerative Disorders Patent Number: 8916388
- *In Vitro* Procedure for Diagnosis and Early Diagnosis of Neurodegenerative Diseases Patent Number: 8298784
- Neurodegenerative Markers for Psychiatric Conditions Publication Number: 20120196300
- *In Vitro* Multiparameter Determination Method for The Diagnosis and Early Diagnosis of Neurodegenerative Disorders Publication Number: 20100062463
- *In Vitro* Method for The Diagnosis and Early Diagnosis of Neurodegenerative Disorders Publication Number: 20100035286
- *In Vitro* Procedure for Diagnosis and Early Diagnosis of Neurodegenerative Diseases Publication Number: 20090263822
- *In Vitro* Method for The Diagnosis of Neurodegenerative Diseases Patent Number: 7547553
- CSF Diagnostic in Vitro Method for Diagnosis of Dementias and Neuroinflammatory Diseases Publication Number: 20080206797
- *In Vitro* Method for The Diagnosis of Neurodegenerative Diseases Publication Number: 20080199966
- Neurodegenerative Markers for Psychiatric Conditions Publication Number: 20080131921
- Method for diagnosis of dementias and neuroinflammatory diseases based on an increased level of procalcitonin in cerebrospinal fluid: Publication number: United StatesPatent 10921330

AV declares no competing financial interests related to the present article and his contribution to this article reflects entirely and only his own academic expertise on the matter. This work was initiated during his previous position at Sorbonne University, Paris, France. AV was an employee of Eisai Inc. AV does not receive any fees or honoraria since November 2019. Before November 2019 he had he received lecture honoraria from Roche, MagQu LLC, and Servier.

SL received lecture honoraria from Roche and Servier.

PL , MCP , MOH , FXL , BD declare no conflict of interest.

KB has served as a consultant, at advisory boards, or at data monitoring committees for Abcam, Axon, Biogen, JOMDD/Shimadzu. Julius Clinical, Lilly, MagQu, Novartis, Roche Diagnostics, and Siemens Healthineers, and is a co-founder of Brain Biomarker Solutions in Gothenburg AB (BBS), which is a part of the GU Ventures Incubator Program, outside the work presented in this paper.

HZ has served at scientific advisory boards for Eisai, Denali, Roche Diagnostics, Wave, Samumed, Siemens Healthineers, Pinteon Therapeutics, Nervgen, AZTherapies and CogRx, has given lectures in symposia sponsored by Cellectricon, Fujirebio, Alzecure and Biogen, and is a co-founder of Brain Biomarker Solutions in Gothenburg AB (BBS), which is a part of the GU Ventures Incubator Program (outside submitted work).

Acknowledgments

The research and this manuscript was part of the translational research program "*PHOENIX*", awarded to *HH*, and administered by the Sorbonne University Foundation and sponsored by la *Fondation pour la Recherche sur Alzheimer*. The study was promoted by INSERM in collaboration with ICM, IHU-A-ICM and Pfizer and has received support within the *"Investissement d'Avenir"* (ANR-10-AIHU-06) French program. The study was promoted in collaboration with the "CHU de Bordeaux" (coordination CIC EC7), the promoter of Memento cohort, funded by the Foundation Plan-Alzheimer. The study was further supported by AVID/Lilly.CATI is a French neuroimaging platform funded by the French Plan Alzheimer (available at http://cati-neuroimaging.com). *HH* is an employee of Eisai Inc. This work has been performed during his previous position at Sorbonne University, Paris, France. At Sorbonne University he was supported by the AXA Research Fund, the "*Fondation partenariale Sorbonne Université*" and the "*Fondation pour la Recherche sur Alzheimer*", Paris, France.

KB is supported by the Swedish Research Council (#2017-00915), the Alzheimer Drug Discovery Foundation (ADDF), USA (#RDAPB-201809-2016615), the Swedish Alzheimer Foundation (#AF-742881), Hjärnfonden, Sweden (#FO2017-0243), the Swedish state under the agreement between the Swedish government and the County Councils, the ALF-agreement (#ALFGBG-715986), the European Union Joint Program for Neurodegenerative Disorders (JPND2019-466-236), and the National Institute of Health (NIH), USA, (grant #1R01AG068398-01).

HZ is a Wallenberg Scholar supported by grants from the Swedish Research Council (#2018-02532), the European Research Council (#681712), Swedish State Support for Clinical Research (#ALFGBG-720931), the Alzheimer Drug Discovery Foundation (ADDF), USA (#201809-2016862), the AD Strategic Fund and the Alzheimer's Association (#ADSF-21-831376-C, #ADSF-21-831381-C and #ADSF-21-831377-C), the Olav Thon Foundation, the Erling-Persson Family Foundation, Stiftelsen för Gamla Tjänarinnor, Hjärnfonden, Sweden (#FO2019-0228), the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement No 860197 (MIRIADE), and the UK Dementia Research Institute at UCL.

Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.neurobiolaging.2021.07.005.

References

Abdi, H., Williams, L.J., 2013. Partial least squares methods: partial least squares correlation and partial least square regression. 549–579. doi:10.1007/978-1-62703-059-5_23

Agosta, F., Pievani, M., Geroldi, C., Copetti, M., Frisoni, G.B., Filippi, M., 2012. Resting state fMRI in Alzheimer's disease: beyond the default mode network. Neurobiol. Aging 33, 1564–1578. doi:10.1016/j.neurobiolaging.2011.06.007.

Aguilar, C., Muehlboeck, J.S., Mecocci, P., Vellas, B., Tsolaki, M., Kloszewska, I., Soininen, H., Lovestone, S., Wahlund, L.O., Simmons, A., Westman, E., 2014. Application of a MRI based index to longitudinal atrophy change in Alzheimer disease, mild cognitive impairment and healthy older individuals in the AddNeuroMed cohort. Front. Aging Neurosci. 6, 1–12. doi:10.3389/fnagi.2014.00145.

Aisen, P.S., Cummings, J., Jack, C.R., Morris, J.C., Sperling, R., Frölich, L., Jones, R.W., Dowsett, S.A., Matthews, B.R., Raskin, J., Scheltens, P., Dubois, B., 2017. On the path to 2025: understanding the Alzheimer's disease continuum. Alzheimers. Res. Ther. 9, 60. doi:10.1186/s13195-017-0283-5.

Andrews-Hanna, J.R., Snyder, A.Z., Vincent, J.L., Lustig, C., Head, D., Raichle, M.E., Buckner, R.L., 2007. Disruption of large-scale brain systems in advanced aging. Neuron 56, 924–935. doi:10.1016/j.neuron.2007.10.038.

Arenaza-Urquijo, E.M., Vemuri, P., 2018. Resistance vs resilience to Alzheimer disease. Neurology 90, 695–703. doi:10.1212/WNL.0000000000005303.

Babiloni, C., Lopez, S., Del Percio, C., Noce, G., Pascarelli, M.T., Lizio, R., Teipel, S.J., González-Escamilla, G., Bakardjian, H., George, N., Cavedo, E., Lista, S., Chiesa, P.A., Vergallo, A., Lemercier, P., Spinelli, G., Grothe, M.J., Potier, M.-C., Stocchi, F., Ferri, R., Habert, M.-O., Fraga, F.J., Dubois, B., Hampel, H., 2020. Resting-state posterior alpha rhythms are abnormal in subjective memory complaint seniors with preclinical alzheimer's neuropathology and high education level: the insight-pread study. Neurobiol. Aging. doi:10.1016/j.neurobiolaging.2020.01.012.

Badhwar, A.P., Tam, A., Dansereau, C., Orban, P., Hoffstaedter, F., Bellec, P., 2017. Resting-state network dysfunction in Alzheimer's disease: A systematic review

and meta-analysis. Alzheimer's dement. Diagnosis, Assess. Dis. Monit. 8, 73–85. doi:10.1016/j.dadm.2017.03.007.

Baldacci, F., Lista, S., Manca, M.L., Chiesa, P.A., Cavedo, E., Lemercier, P., Zetterberg, H., Blennow, K., Habert, M.-O., Potier, M.C., Dubois, B., Vergallo, A., Hampel, H., 2020. Age and sex impact plasma NFL and t-Tau trajectories in individuals with subjective memory complaints: a 3-year follow-up study. Alzheimers. Res. Ther. 12, 147. doi:10.1186/s13195-020-00704-4.

Bateman, R.J., Xiong, C., Benzinger, T.L.S., Fagan, A.M., Goate, A., Fox, N.C., Marcus, D.S., Cairns, N.J., Xie, X., Blazey, T.M., Holtzman, D.M., Santacruz, A., Buckles, V., Oliver, A., Moulder, K., Aisen, P.S., Ghetti, B., Klunk, W.E., McDade, E., Martins, R.N., Masters, C.L., Mayeux, R., Ringman, J.M., Rossor, M.N., Schofield, P.R., Sperling, R.A., Salloway, S., Morris, J.C., 2012. Clinical and biomarker changes in dominantly inherited alzheimer's disease. N. Engl. J. Med. 367, 795–804. doi:10.1056/NEJMoa1202753.

Bero, A.W., Yan, P., Roh, J.H., Cirrito, J.R., Stewart, F.R., Raichle, M.E., Lee, J.-M., Holtzman, D.M., 2011. Neuronal activity regulates the regional vulnerability to amyloid-β deposition. Nat. Neurosci. 14, 750–756. doi:10.1038/nn.2801.

Brier, M.R., Thomas, J.B., Snyder, A.Z., Benzinger, T.L., Zhang, D., Raichle, M.E., Holtzman, D.M., Morris, J.C., Ances Dr., B.M., 2012. Loss of intranetwork and internetwork resting state functional connections with Alzheimer's disease progression. J. Neurosci. 32, 8890–8899. doi:10.1523/JNEUROSCI.5698-11.2012.

Buckley, R.F., Maruff, P., Ames, D., Bourgeat, P., Martins, R.N., Masters, C.L., Rainey-Smith, S., Lautenschlager, N., Rowe, C.C., Savage, G., Villemagne, V.L., Ellis, K.A., 2016. Subjective memory decline predicts greater rates of clinical progression in preclinical Alzheimer's disease. Alzheimer's Dement 12, 796–804. doi:10.1016/j.jalz.2015.12.013.

Buckley, R.F., Schultz, A.P., Hedden, T., Papp, K.V., Hanseeuw, B.J., Marshall, G., Sepulcre, J., Smith, E.E., Rentz, D.M., Johnson, K.A., Sperling, R.A., Chhatwal, J.P., 2017. Functional network integrity presages cognitive decline in preclinical Alzheimer disease. Neurology 89, 29–37. doi:10.1212/WNL.0000000000004059.

Cacciamani, F., Sambati, L., Houot, M., Habert, M.-O., Dubois, B., Epelbaum, S., 2020. Awareness of cognitive decline trajectories in asymptomatic individuals at risk for AD. Alzheimers. Res. Ther. 12, 129. doi:10.1186/s13195-020-00700-8.

Chhatwal, J.P., Schultz, A.P., Johnson, K.A., Hedden, T., Jaimes, S., Benzinger, T.L.S., Jack, C., Ances, B.M., Ringman, J.M., Marcus, D.S., Ghetti, B., Farlow, M.R., Danek, A., Levin, J., Yakushev, I., Laske, C., Koeppe, R.A., Galasko, D.R., Xiong, C., Masters, C.L., Schofield, P.R., Kinnunen, K.M., Salloway, S., Martins, R.N., McDade, E., Cairns, N.J., Buckles, V.D., Morris, J.C., Bateman, R., Sperling, R.A., 2018. Preferential degradation of cognitive networks differentiates Alzheimer's disease from ageing. Brain 141, 1486–1500. doi:10.1093/brain/awy053.

Chiesa, P.A., Cavedo, E., Vergallo, A., Lista, S., Potier, M.-C., Habert, M.-O., Dubois, B., Thiebaut de Schotten, M., Hampel, H.Alzheimer Precision Medicine Initiative (APMI), 2019. Differential default mode network trajectories in asymptomatic individuals at risk for Alzheimer's disease. Alzheimers. Dement 15, 940–950. doi:10.1016/j.jalz.2019.03.006.

Chiesa, P.A., Houot, M., Vergallo, A., Cavedo, E., Lista, S., Potier, M.C., Zetterberg, H., Blennow, K., Vanmechelen, E., De Vos, A., Dubois, B., Hampel, H., 2020. Association of brain network dynamics with plasma biomarkers in subjective memory complainers. Neurobiol. Aging 88, 83–90. doi:10.1016/j.neurobiolaging.2019.12.017.

Chong, I.G., Jun, C.H., 2005. Performance of some variable selection methods when multicollinearity is present. Chemom. Intell. Lab. Syst. 78, 103–112. doi:10.1016/j.chemolab.2004.12.011.

Clancy, U., Gilmartin, D., Jochems, A.C.C., Knox, L., Doubal, F.N., Wardlaw, J.M., 2021. Neuropsychiatric symptoms associated with cerebral small vessel disease: a systematic review and meta-analysis. the lancet psychiatry 8, 225–236. doi:10.1016/S2215-0366(20)30431-4

Cramer, R.D., 1993. Partial least squares (PLS): Its strengths and limitations. Perspect. Drug Discov. Des. 1, 269–278. doi:10.1007/BF02174528.

Crossley, N.A., Mechelli, A., Scott, J., Carletti, F., Fox, P.T., Mcguire, P., Bullmore, E.T., 2014. The hubs of the human connectome are generally implicated in the anatomy of brain disorders. Brain 137, 2382–2395. doi:10.1093/brain/awu132.

Dayon, L., Núñez Galindo, A., Wojcik, J., Cominetti, O., Corthésy, J., Oikonomidi, A., Henry, H., Kussmann, M., Migliavacca, E., Severin, I., Bowman, G.L., Popp, J., 2018. Alzheimer disease pathology and the cerebrospinal fluid proteome. Alzheimers. Res. Ther. 10, 66. doi:10.1186/s13195-018-0397-4.

Desboulets, L., 2018. A review on variable selection in regression analysis. Econometrics 6, 45. doi:10.3390/econometrics6040045, https://doi.org/.

Dubois, B., Epelbaum, S., Nyasse, F., Bakardjian, H., Gagliardi, G., Uspenskaya, O., Houot, M., Lista, S., Cacciamani, F., Potier, M.C., Bertrand, A., Lamari, F., Benali, H., Mangin, J.F., Colliot, O., Genthon, R., Habert, M.O., Hampel, H., Audrain, C., Auffret, A., Baldacci, F., Benakki, I., Bertin, H., Boukadida, L., Cavedo, E., Chiesa, P., Dauphinot, L., Santos, Dos, A., Dubois, M., Durrleman, S., Fontaine, G., Genin, A., Glasman, P., Jungalee, N., Kas, A., Kilani, M., La Corte, V., Lehericy, S., Letondor, C., Levy, M., Lowrey, M., Ly, J., Makiese, O., Metzinger, C., Michon, A., Mochel, F., Poisson, C., Ratovohery, S., Revillon, M., Rojkova, K., Roy, P., Santos-Andrade, K., Schindler, R., Seux, L., Simon, V., Sole, M., Tandetnik, C., Teichmann, M., Thiebaut de Shotten, M., Younsi, N., 2018. Cognitive and neuroimaging features and brain β-amyloidosis in individuals at risk of Alzheimer's disease (INSIGHT-preAD): a longitudinal observational study. Lancet Neurol 17, 335–346. doi:10.1016/S1474-4422(18)30029-2.

Dubois, B., Feldman, H.H., Jacova, C., Hampel, H., Molinuevo, J.L., Blennow, K., DeKosky, S.T., Gauthier, S., Selkoe, D., Bateman, R., Cappa, S., Crutch, S., Engelborghs, S., Frisoni, G.B., Fox, N.C., Galasko, D., Habert, M.-O., Jicha, G.A., Nordberg, A., Pasquier, F., Rabinovici, G., Robert, P., Rowe, C., Salloway, S., Sarazin, M., Epelbaum, S., de Souza, L.C., Vellas, B., Visser, P.J., Schneider, L., Stern, Y., Scheltens, P., Cummings, J.L., 2014. Advancing research diagnostic criteria for Alzheimer's disease: the IWG-2 criteria. Lancet Neurol 13, 614–629. doi:10.1016/S1474-4422(14)70090-0.

Elman, J.A., Oh, H., Madison, C.M., Baker, S.L., Vogel, J.W., Marks, S.M., Crowley, S., O'Neil, J.P., Jagust, W.J., 2014. Neural compensation in older people with brain amyloid-β deposition. Nat. Neurosci. 17, 1316–1318. doi:10.1038/nn.3806.

Fan, L.Y., Tzen, K.Y., Chen, Y.F., Chen, T.F., Lai, Y.M., Yen, R.F., Huang, Y.Y., Shiue, C.Y., Yang, S.Y., Chiu, M.J., 2018. The relation between brain amyloid deposition, cortical atrophy, and plasma biomarkers in amnesic mild cognitive impairment and Alzheimer's. Disease. Front. Aging Neurosci. 10, 1–10. doi:10.3389/fnagi.2018.00175.

Fandos, N., Pérez-Grijalba, V., Pesini, P., Olmos, S., Bossa, M., Villemagne, V.L., Doecke, J., Fowler, C., Masters, C.L., Sarasa, M., 2017. Plasma amyloid β 42/40 ratios as biomarkers for amyloid β cerebral deposition in cognitively normal individuals. Alzheimer's Dement. Diagnosis, Assess. Dis. Monit. 8, 179–187. doi:10.1016/j.dadm.2017.07.004.

Farrell, M.E., Chen, X., Rundle, M.M., Chan, M.Y., Wig, G.S., Park, D.C., 2018. Regional amyloid accumulation and cognitive decline in initially amyloid-negative adults. Neurology 91, e1809–e1821. doi:10.1212/WNL.0000000000006469.

Gaubert, S., Raimondo, F., Houot, M., Corsi, M.-C., Naccache, L., Diego Sitt, J., Hermann, B., Oudiette, D., Gagliardi, G., Habert, M.-O., Dubois, B., De Vico Fallani, F., Bakardjian, H., Epelbaum, S., 2019. EEG evidence of compensatory mechanisms in preclinical Alzheimer's disease. Brain 142, 2096–2112. doi:10.1093/brain/awz150.

Grothe, M.J., Teipel, S.J., 2016. Spatial patterns of atrophy, hypometabolism, and amyloid deposition in Alzheimer's disease correspond to dissociable functional brain networks. Hum. Brain Mapp. 37, 35–53. doi:10.1002/hbm.23018.

Habert, Marie Odile, Bertin, Hugo, Labit, M., Diallo, M., Marie, S., Martineau, K., Kas, Aurélie, Causse-Lemercier, Valérie, Bakardjian, Hovagim, Epelbaum, Stéphane, Chételat, G., Houot, M., Hampel, Harald, Dubois, Bruno, Mangin, J.F., Audrain, C., Bakardjian, H., Benali, H., Bertin, H., Boukadida, L., Cacciamani, F., Causse-Lemercier, V., Cavedo, E., Chiesa, P., Colliot, O., Santos, Dos, A., Dubois, B., Durrleman, S., Epelbaum, S., Gagliardi, G., Genthon, R., Habert, M., O., Hampel, H., Jungalee, N., Kas, A., Lehericy, S., Lamari, F., Letondor, C., Levy, M., Lista, S., Mochel, F., Nyasse, F., Poisson, C., Potier, M.C., Revillon, M., Rojkova, K., Roy, P., Santos-Andrade, K., Santos, A., Simon, V., Sole, M., Tandetnik, C., Thiebaud De Schotten, M., 2018. Evaluation of amyloid status in a cohort of elderly individuals with memory complaints: validation of the method of quantification and determination of positivity thresholds. Ann. Nucl. Med. 32, 75–86. doi:10.1007/s12149-017-1221-0.

Hampel, H., Lista, S., Neri, C., Vergallo, A., 2019a. Time for the systems-level integration of aging: Resilience enhancing strategies to prevent Alzheimer's disease. Prog. Neurobiol. 181, 101662. doi:10.1016/j.pneurobio.2019.101662.

Hampel, H., Vergallo, A., Perry, G., Lista, S., 2019b. The alzheimer precision medicine initiative. J. Alzheimer's Dis. 68, 1–24. doi:10.3233/JAD-181121.

Hampton, O.L., Buckley, R.F., Manning, L.K., Scott, M.R., Properzi, M.J., Peña-Gómez, C., Jacobs, H.I.L., Chhatwal, J.P., Johnson, K.A., Sperling, R.A., Schultz, A.P., 2020. Resting-state functional connectivity and amyloid burden influence longitudinal cortical thinning in the default mode network in preclinical Alzheimer's disease. NeuroImage Clin 28, 102407. doi:10.1016/j.nicl.2020.102407.

Hastie, T., Tibshirani, R., Friedman, J., 2009. The Elements of Statistical Learning. Springer, New York, New York, NY Springer Series in Statistics. Springer-https://doi.org/ doi:10.1007/978-0-387-84858-7.

He, X., Qin, W., Liu, Y., Zhang, X., Duan, Y., Song, J., Li, K., Jiang, T., Yu, C., 2014. Abnormal salience network in normal aging and in amnestic mild cognitive impairment and Alzheimer's disease. Hum. Brain Mapp 35, 3446–3464. doi:10.1002/hbm.22414.

Hohman, T.J., McLaren, D.G., Mormino, E.C., Gifford, K.A., Libon, D.J., Jefferson, A.L., 2016. Asymptomatic Alzheimer disease: defining resilience. Neurology 87, 2443–2450. doi:10.1212/WNL.0000000000003397.

Jack, C.R., Bennett, D.A., Blennow, K., Carrillo, M.C., Dunn, B., Haeberlein, S.B., Holtzman, D.M., Jagust, W., Jessen, F., Karlawish, J., Liu, E., Molinuevo, J.L., Montine, T., Phelps, C., Rankin, K.P., Rowe, C.C., Scheltens, P., Siemers, E., Snyder, H.M., Sperling, R., Elliott, C., Masliah, E., Ryan, L., Silverberg, N., 2018. NIA-AA Research framework: toward a biological definition of alzheimer's disease. alzheimer's dement. 14, 535–562. doi:10.1016/j.jalz.2018.02.018

Johnson, W.E., Li, C., Rabinovic, A., 2007. Adjusting batch effects in microarray expression data using empirical Bayes methods. Biostatistics 8, 118–127. doi:10.1093/biostatistics/kxj037.

Kohannim, O., 2012. Discovery and replication of gene influences on brain structure using LASSO regression. Front. Neurosci. 6. doi:10.3389/fnins.2012.00115.

Krishnan, A., Williams, L.J., McIntosh, A.R., Abdi, H., 2011. Partial Least Squares (PLS) methods for neuroimaging: A tutorial and review. Neuroimage 56, 455–475. doi:10.1016/j.neuroimage.2010.07.034.

Landau, S.M., Harvey, D., Madison, C.M., Reiman, E.M., Foster, N.L., Aisen, P.S., Petersen, R.C., Shaw, L.M., Trojanowski, J.Q., Jack, C.R., Weiner, M.W., Jagust, W.J., 2010. Comparing predictors of conversion and decline in mild cognitive impairment. Neurology 75, 230–238. doi:10.1212/WNL.0b013e3181e8e8b8.

Langbaum, J.B., Ellison, N.N., Caputo, A., Thomas, R.G., Langlois, C., Riviere, M.-E., Graf, A., Lopez Lopez, C., Reiman, E.M., Tariot, P.N., Hendrix, S.B., 2020. The Alzheimer's prevention initiative composite cognitive test: a practical measure for tracking cognitive decline in preclinical alzheimer's disease. Alzheimers. Res. Ther. 12, 66. doi:10.1186/s13195-020-00633-2.

Li, X., Wang, H., Long, J., Pan, G., He, T., Anichtchik, O., Belshaw, R., Albani, D., Edison, P., Green, E.K., Scott, J., 2018. Systematic analysis and biomarker study for alzheimer's. Disease. Sci. Rep. 8, 17394. doi:10.1038/s41598-018-35789-3.

Li, Y., Yao, Z., Yu, Y., Zou, Y., Fu, Y., Hu, B., 2019. Brain network alterations in individuals with and without mild cognitive impairment: parallel independent component analysis of AV1451 and AV45 positron emission tomography. BMC Psychiatry 19, 165. doi:10.1186/s12888-019-2149-9.

Lockhart, S.N., Schöll, M., Baker, S.L., Ayakta, N., Swinnerton, K.N., Bell, R.K., Mellinger, T.J., Shah, V.D., O'Neil, J.P., Janabi, M., Jagust, W.J., 2017. Amyloid and tau PET demonstrate region-specific associations in normal older people. Neuroimage 150, 191–199. doi:10.1016/j.neuroimage.2017.02.051.

Lorenzi, M., Altmann, A., Gutman, B., Wray, S., Arber, C., Hibar, D.P., Jahanshad, N., Schott, J.M., Alexander, D.C., Thompson, P.M., Ourselin, S., 2018. Susceptibility of brain atrophy to TRIB3 in Alzheimer's disease, evidence from functional prioritization in imaging genetics. Proc. Natl. Acad. Sci. 115, 3162–3167. doi:10.1073/pnas.1706100115.

McKhann, G.M., Knopman, D.S., Chertkow, H., Hyman, B.T., Jack, C.R., Kawas, C.H., Klunk, W.E., Koroshetz, W.J., Manly, J.J., Mayeux, R., Mohs, R.C., Morris, J.C., Rossor, M.N., Scheltens, P., Carrillo, M.C., Thies, B., Weintraub, S., Phelps, C.H., 2011. The diagnosis of dementia due to alzheimer's disease: recommendations from the national institute on aging-alzheimer's association workgroups on diagnostic guidelines for alzheimer's disease. alzheimer's dement. 7, 263–269. doi:10.1016/j.jalz.2011.03.005

Mehmood, T., Liland, K.H., Snipen, L., Sæbø, S., 2012. A review of variable selection methods in partial least squares regression. Chemom. Intell. Lab. Syst. 118, 62–69. doi:10.1016/j.chemolab.2012.07.010.

Mormino, E.C., Smiljic, A., Hayenga, A.O., Onami, H., Greicius M.D., S., G.D., Rabinovici, M., Janabi, S.L., Baker, I., V.Yen, C.M., Madison, B.L., Miller, W.J., Jagust, 2011. Relationships between beta-amyloid and functional connectivity in different components of the default mode network in aging. Cereb. Cortex 21, 2399–2407. doi:10.1093/cercor/bhr025.

Nakamura, A., Kaneko, N., Villemagne, V.L., Kato, T., Doecke, J., Doré, V., Fowler, C., Li, Q.-X., Martins, R., Rowe, C., Tomita, T., Matsuzaki, K., Ishii, K.K., Ishii, K.K., Arahata, Y., Iwamoto, S., Ito, K., Tanaka, K., Masters, C.L., Yanagisawa, K., Rowe, C., Li, Q.-X., Fowler, C., Kato, T., Nakamura, A., Kaneko, N., Doecke, J., Yanagisawa, K., Arahata, Y., Ishii, K.K., Tanaka, K., Villemagne, V.L., Iwamoto, S., Ito, K., Matsuzaki, K., Doré, V., Tomita, T., Martins, R., Ishii, K.K., 2018. High performance plasma amyloid-β biomarkers for Alzheimer's disease. Nature 554, 249–254. doi:10.1038/nature25456.

Negash, S., Xie, S., Davatzikos, C., Clark, C.M., Trojanowski, J.Q., Shaw, L.M., Wolk, D.A., Arnold, S.E., 2013. Cognitive and functional resilience despite molecular evidence of Alzheimer's disease pathology. Alzheimers. Dement. 9, e89–e95. doi:10.1016/j.jalz.2012.01.009.

Palermo, G., Piraino, P., Zucht, H.-D., 2009. Performance of PLS regression coefficients in selecting variables for each response of a multivariate PLS for omics-type data. Adv. Appl. Bioinform. Chem. 2, 57–70. doi:10.2147/aabc.s3619.

Palmqvist, S., Janelidze, S., Stomrud, E., Zetterberg, H., Karl, J., Zink, K., Bittner, T., Mattsson, N., Eichenlaub, U., Blennow, K., Hansson, O., 2019. Performance of fully automated plasma assays as screening tests for alzheimer disease–related β-amyloid status. JAMA Neurol 76, 1060. doi:10.1001/jamaneurol.2019.1632.

Palmqvist, S., Schöll, M., Strandberg, O., Mattsson, N., Stomrud, E., Zetterberg, H., Blennow, K., Landau, S., Jagust, W., Hansson, O., 2017. Earliest accumulation of β-amyloid occurs within the default-mode network and concurrently affects brain connectivity. Nat. Commun. 8, 1214. doi:10.1038/s41467-017-01150-x.

Pérez-Grijalba, V., Romero, J., Pesini, P., Sarasa, L., Monleón, I., San-José, I., Arbizu, J., Martínez-Lage, P., Munuera, J., Ruiz, A., Tárraga, L., Boada, M., Sarasa, M., 2019. Plasma Aβ42/40 ratio detects early stages of alzheimer's disease and correlates with csf and neuroimaging biomarkers in the AB255 study. J. Prev. Alzheimer's Dis. 6, 34–41. doi:10.14283/jpad.2018.41.

Perez-Nievas, B.G., Stein, T.D., Tai, H.-C., Dols-Icardo, O., Scotton, T.C., Barroeta-Espar, I., Fernandez-Carballo, L., de Munain, E.L., Perez, J., Marquie, M., Serrano-Pozo, A., Frosch, M.P., Lowe, V., Parisi, J.E., Petersen, R.C., Ikonomovic, M.D., López, O.L., Klunk, W., Hyman, B.T., Gómez-Isla, T., 2013. Dissecting phenotypic traits linked to human resilience to Alzheimer's pathology. Brain 136, 2510–2526. doi:10.1093/brain/awt171.

Rohart, F., Gautier, B., Singh, A., Lê Cao, K.-A., 2017. mixOmics: An R package for 'omics feature selection and multiple data integration. PLOS Comput. Biol. 13, e1005752. doi:10.1371/journal.pcbi.1005752.

Rosseel, Y., 2012. lavaan : An R package for structural equation modeling. J. Stat. Softw. 48. doi:10.18637/jss.v048.i02.

Shirer, W.R., Ryali, S., Rykhlevskaia, E., Menon, V., Greicius, M.D., 2012. Decoding subject-driven cognitive states with whole-brain connectivity patterns. Cereb. Cortex 22, 158–165. doi:10.1093/cercor/bhr099.

Teipel, S., Grothe, M.J., Zhou, J., Sepulcre, J., Dyrba, M., Sorg, C., Babiloni, C., 2016. Measuring cortical connectivity in Alzheimer's disease as a brain neural network pathology: Toward clinical applications. J. Int. Neuropsychol. Soc. 22, 138–163. doi:10.1017/S1355617715000995.

Teipel, S.J., Dyrba, M., Chiesa, P.A., Sakr, F., Jelistratova, I., Lista, S., Vergallo, A., Lemercier, P., Cavedo, E., Habert, M.O., Dubois, B., Hampel, H., Grothe, M.J., 2020. In vivo staging of regional amyloid deposition predicts functional conversion in the preclinical and prodromal phases of Alzheimer's disease. Neurobiol. Aging 93, 98–108. doi:10.1016/j.neurobiolaging.2020.03.011.

Thambisetty, M., Simmons, A., Hye, A., Campbell, J., Westman, E., Zhang, Y., Wahlund, L.O., Kinsey, A., Causevic, M., Killick, R., Kloszewska, I., Mecocci, P., Soininen, H., Tsolaki, M., Vellas, B., Spenger, C., Lovestone, S., 2011. Plasma biomarkers of brain atrophy in Alzheimer's disease. PLoS One 6, 1–7. doi:10.1371/journal.pone.0028527.

Timmers, T., Ossenkoppele, R., Verfaillie, S.C.J., van der Weijden, C.W.J., Slot, R.E.R., Wesselman, L.M.P., Windhorst, A.D., Wolters, E.E., Yaqub, M., Prins, N.D., Lammertsma, A.A., Scheltens, P., van der Flier, W.M., van Berckel, B.N.M., 2019. Amyloid PET and cognitive decline in cognitively normal individuals: the SCIENCe project. Neurobiol. Aging 79, 50–58. doi:10.1016/j.neurobiolaging.2019.02.020.

Van Etten, E.J., Bharadwaj, P.K., Nguyen, L.A., Hishaw, G.A., Trouard, T.P., Alexander, G.E., 2020. Right hippocampal volume mediation of subjective memory complaints differs by hypertension status in healthy aging. Neurobiol. Aging 94, 271–280. doi:10.1016/j.neurobiolaging.2020.06.012.

van Harten, A.C., Mielke, M.M., Swenson-Dravis, D.M., Hagen, C.E., Edwards, K.K., Roberts, R.O., Geda, Y.E., Knopman, D.S., Petersen, R.C., 2018. Subjective cognitive decline and risk of MCI: the mayo clinic study of aging. Neurology 91, e300–e312. doi:10.1212/WNL.0000000000005863.

van Harten, A.C., Visser, P.J., Pijnenburg, Y.A.L., Teunissen, C.E., Blankenstein, M.A., Scheltens, P., van der Flier, W.M., 2013. Cerebrospinal fluid Aβ42 is the best predictor of clinical progression in patients with subjective complaints. Alzheimer's Dement. 9, 481–487. doi:10.1016/j.jalz.2012.08.004

Vardarajan, B., Kalia, V., Manly, J., Brickman, A., Reyes-Dumeyer, D., Lantigua, R., Ionita-Laza, I., Jones, D.P., Miller, G.W., Mayeux, R., 2020. Differences in plasma metabolites related to Alzheimer's disease, APOE ε4 status, and ethnicity. Alzheimer's Dement. Transl. Res. Clin. Interv. 6. doi:10.1002/trc2.12025.

Vergallo, A., Mégret, L., Lista, S., Cavedo, E., Zetterberg, H., Blennow, K., Vanmechelen, E., De Vos, A., Habert, M.-O., Potier, M.-C., Dubois, B., Neri, C., Hampel, H.INSIGHT-preAD study group, Alzheimer Precision Medicine Initiative (APMI), 2019. Plasma amyloid β 40/42 ratio predicts cerebral amyloidosis in cognitively normal individuals at risk for Alzheimer's disease. Alzheimers. Dement. 15, 764–775. doi:10.1016/j.jalz.2019.03.009.

Veronese, M., Moro, L., Arcolin, M., Dipasquale, O., Rizzo, G., Expert, P., Khan, W., Fisher, P.M., Svarer, C., Bertoldo, A., Howes, O., Turkheimer, F.E., 2019. Covariance statistics and network analysis of brain PET imaging studies. Sci. Rep. 9, 1–15. doi:10.1038/s41598-019-39005-8.

Villemagne, V.L., Burnham, S., Bourgeat, P., Brown, B., Ellis, K.A., Salvado, O., Szoeke, C., Macaulay, S.L., Martins, R., Maruff, P., Ames, D., Rowe, C.C., Masters, C.L., 2013. Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. Lancet Neurol 12, 357–367. doi:10.1016/S1474-4422(13)70044-9.

Warrens, M.J., 2015. Five ways to look at cohen's kappa. J. Psychol. Psychother. 05. doi:10.4172/2161-0487.1000197.

Whittington, A., Sharp, D.J., Gunn, R.N., 2018. Spatiotemporal distribution of β-amyloid in alzheimer disease is the result of heterogeneous regional carrying capacities. J. Nucl. Med. 59, 822–827. doi:10.2967/jnumed.117.194720.

Wickham, H., 2009. ggplot2. Springer, New York, New York, NY doi:10.1007/978-0-387-98141-3.

Wold, S., Johansson, E., Cocchi, M., 1993. PLS - Partial least squares projections to latent structures. In: Kubinyi, H. (Ed.), 3D QSAR in Drug Design: Theory Methods and Applications. Escom, pp. 523–550.

Xicota, L., Ichou, F., Lejeune, F.-X., Colsch, B., Tenenhaus, A., Leroy, I., Fontaine, G., Lhomme, M., Bertin, H., Habert, M.-O., Epelbaum, S., Dubois, B., Mochel, F., Potier, M.-C., 2019. Multi-omics signature of brain amyloid deposition in asymptomatic individuals at-risk for Alzheimer's disease: The INSIGHT-preAD study. EBioMedicine 47, 518–528. doi:10.1016/j.ebiom.2019.08.051.

Xu, Y., Goodacre, R., 2018. On splitting training and validation set: a comparative study of cross-validation, bootstrap and systematic sampling for estimating the generalization performance of supervised learning. J. Anal. Test. 2, 249–262. doi:10.1007/s41664-018-0068-2.

Yang, T., Wang, J., Sun, Q., Hibar, D.P., Jahanshad, N., Liu, L., Wang, Y., Zhan, L., Thompson, P.M., Ye, J., 2015. Detecting genetic risk factors for Alzheimer's disease in whole genome sequence data via Lasso screening, in: 2015 IEEE 12th International Symposium on Biomedical Imaging (ISBI). IEEE 985–989. doi:10.1109/ISBI.2015.7164036.

Zhao, Q., Sang, X., Metmer, H., Swati, Z., nawab, N.K., Lu, J., 2019. Functional segregation of executive control network and frontoparietal network in Alzheimer's disease. Cortex 120, 36–48. doi:10.1016/j.cortex.2019.04.026.

Zhou, J., Greicius, M.D., Gennatas, E.D., Growdon, M.E., Jang, J.Y., Rabinovici, G.D., Kramer, J.H., Weiner, M., Miller, B.L., Seeley, W.W., 2010. Divergent network connectivity changes in behavioural variant frontotemporal dementia and Alzheimer's disease. Brain 133, 1352–1367. doi:10.1093/brain/awq075.

Zou, H., Hastie, T., 2005. Regularization and variable selection via the elastic net. J. R. Stat. Soc. Ser. B Stat. Methodol. 67, 301–320. doi:10.1111/j.1467-9868.2005.00503.x.