

Gut microbiota and vitamin status in persons with obesity: a key interplay

Lise Voland, Tiphaine Le Roy, Jean Debédat, Karine Clément

To cite this version:

Lise Voland, Tiphaine Le Roy, Jean Debédat, Karine Clément. Gut microbiota and vitamin status in persons with obesity: a key interplay. Obesity Reviews, 2021, 10.1111/obr.13377. hal-03440317

HAL Id: hal-03440317 <https://hal.sorbonne-universite.fr/hal-03440317v1>

Submitted on 22 Nov 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

- **Abstract**
-

 There are numerous factors involved in obesity progression and maintenance including systemic low-grade inflammation, adipose tissue dysfunction or gut microbiota dysbiosis. Recently, a growing interest has arisen for vitamins' role in obesity and related disorders, both 34 at the host and gut bacterial level. Indeed, vitamins are provided mostly by food but some, from the B and K groups in particular, can be synthesized by the gut bacterial ecosystem and absorbed in the colon. Knowing that vitamin deficiency can alter many important cellular functions and lead to serious health issues, it is important to carefully monitor the vitamin status of patients with obesity and potentially already existing comorbidities as well as dysbiotic gut microbiota and thus potentially altered bacterial metabolism of vitamins. In this review, we examined both murine and human studies, to assessthe prevalence of sub-optimal levels of several vitamins in obesity and metabolic alterations. This review also examines the 42 relationship between vitamins and the gut microbiota in terms of vitamin production and the modulation of the gut bacterial ecosystem in conditions of vitamin shortage or supplementation. Furthermore, some strategies to improve vitamin status of patients with severe obesity are proposed within this review.

Key words: biotin, gut microbiota, micronutrients, obesity, vitamins

-
-
-

 Obesity represents a global health issue with an ever-increasing prevalence over the 56 . Iast 50 years^{1,2}. It is expected that over 40% of the world's population will suffer from obesity 57 by 2030³. Obesity is associated with several complications such as type 2 diabetes, 58 cardiovascular diseases and cancers⁴. Triggered by multiple environmental factors, the development and progression of obesity are characterized by organ structural and biological alterations and systemic low-grade inflammation. Recently, the gut microbiota (GM) has emerged as a contributive "organ" in the development and maintenance of obesity.

 The GM is a complex and abundant ecosystem composed of trillions of microorganisms including bacteria, viruses, archaea, fungi, and phages. The microorganisms present within the gut harbor a myriad of functions, and in particular, the GM is implicated in the fermentation of non-digestible food substrates. This microbial metabolism leads to the production of an important variety of metabolites, which in return are known to have a broad range of bioactivities, including immune modulation and intestinal barrier homeostasis. During obesity, an imbalance in the GM composition, functionality, as well as a decreased bacterial richness 69 and diversity, collectively referred to as GM dysbiosis, have been reported by many studies⁵.

 Among the numerous GM-derived metabolites (as short chain fatty acids, secondary bile acid molecules, tryptophan derivates to cite only a few), vitamins have been relatively 72 neglected in this context. Yet, gut bacteria are known to produce vitamins from the B and K 73 groups⁶. Vitamins are essential micronutrients, obtained from food and, to some extent, from gut bacterial synthesis. Vitamins serve as coenzymes or cofactors for numerous processes 75 involved in host energy metabolism⁷ (Table 1). For example, B vitamins play a key role in maintaining mitochondrial energy metabolism, which is altered by inadequate levels of any of

77 those vitamins⁸ (Table 1). Vitamin deficiency, due to an imbalanced diet, increased host physiological requirements or altered drug-micronutrient interactions, affects host health in varying ways and can lead to severe clinical consequences, ranging from fatigue to 80 dermatological, digestive, cardiovascular or neurological and depressive disorders⁹ (Table 1). 81 Dietary vitamin absorption occurs mainly in the small intestine for the dietary vitamins but it 82 is now acknowledged that the distal gut may also contribute to vitamin absorption and 83 especially bacterially-produced vitamins¹⁰. Thus, the alteration of the intestinal environment and absorptive capacities potentially alters vitamins host vitamin status.

 Despite increasing evidence of deep alterations of gut physiology as well as microbiota composition and functionality observed during obesity and metabolic disorders, the consequences on vitamins biosynthesis and availability have scarcely been studied. Herein, we review vitamin status alterations observed in obesity and metabolic disorders and focus on the production potential of vitamins by the GM as well as its modulation by both vitamin supplementation and deficiency, mostly focusing on B and D vitamins. We also discuss some future strategies for vitamin status management in severe obesity.

Evidence of vitamin deficiency in obesity and related metabolic disorders in humans

 The optimal vitamin reference levels for the general population are reported by national or international expert committees such as the European Food Safety Authority 97 (EFSA) or the US Centers for Disease and Prevention through the National Health and Nutrition Examination Survey (NHANES). Lately, an increasing interest for vitamin status in subjects with obesity has arisen as many studies have revealed that obesity and related

100 metabolic complications are associated with deficiencies or at least sub-optimal blood 101 concentrations of many vitamins, which has been well-reviewed recently^{12,13}.

102 Briefly, a series of studies exploring different populations with metabolic alterations 103 revealed suboptimal blood level of B vitamins, including vitamin B1 (thiamine), B9 (folate), B6 104 (pyridoxine), B12 (cobalamin). A decrease of vitamin B9 has been observed in pregnant 105 women with overweight in comparison to women with a BMI ranging between 18 and 25 106 $\,$ kg/m^{2 14}. Similarly, another study on patients with severe obesity reported an important 107 decrease of vitamin B6 serum concentrations as compared to patients without obesity¹⁵. 108 Moreover, up to 75% patients with type 2 diabetes presented low blood concentration of 109 vitamin B1¹⁶. Vitamin B12 levels have also been linked with obesity with a significant negative 110 correlation reported between body mass index (BMI) and vitamin B12 blood levels in a cohort 111 of almost 1000 patients with obesity¹⁷. Moreover, the authors of this study also found a trend 112 toward decreased vitamin B12 blood concentrations in subjects with insulin resistance or 113 metabolic syndrome in comparison to participants without metabolic alterations¹⁷. In another 114 cohort of participants with overweight or obesity, 13% of participants were found to be 115 deficient in vitamin B12¹⁸ while in the general US population, the prevalence of vitamin B12 116 was found to be around 5% and increases with age¹⁹. Interestingly, the serum levels of vitamin 117 B12 are increased after a weight loss induced by a Low Energy Diet (LED)¹⁸. Another B vitamin, 118 vitamin B7 (biotin) has been shown to be decreased in subjects with moderate obesity²⁰ and 119 t type 2 diabetes²¹. Yet, this vitamin has been neglected in the context of obesity and metabolic 120 disorders; as well as other B vitamins such as vitamins B2 (riboflavin), B3 (niacin).

121 Vitamin deficiency in metabolic disorders extends to vitamin D as well. In Geiker et al., 122 insufficient levels of vitamin D were observed in 72% of the participants before the LED-123 induced weight loss, and again, the serum concentration of vitamin D was increased after the

124 nutritional intervention¹⁸. Unlike B vitamins, the links between vitamin D status and obesity 125 and its complications have been investigated in a larger number of studies and were reported 126 as early as 1985²². It has been commonly accepted that vitamin D deficiency can be estimated 127 via the serum concentration of the vitamin D metabolite, 25-hydroxycholecalciferol 128 [25(OH)D], the active form of vitamin D, which integrates both the dietary intake and the 129 endogenous production. For example, analyses in a large population of older individuals 130 revealed a significant inverse association between BMI and more specifically adiposity 131 (especially in the 3rd and 4th quartile when fat percentage was divided in quartiles) and serum 132 25(OH)D concentrations²³. Moreover, several groups also showed inverse correlations 133 between 25(OH)D serum concentrations and markers of metabolic alterations such as 134 triglycerides²⁴ or fasting glucose and the HOMA-IR index, a surrogate marker of insulin 135 resistance²⁵. An observational transversal study including 73 patients with morbid obesity 136 confirmed the higher prevalence of vitamin D deficiency in comparison to the population 137 without obesity²⁴. Here, they found that 50.7% of the participants had vitamin D deficiency. 138 They also reported an increased prevalence of vitamin D deficiency associated with the 139 occurrence of metabolic syndrome in this population. This correlation was confirmed in a large 140 study of over 2 500 Chinese men and women, in which, lower 25(OH)D serum concentration 141 was associated with increased insulin resistance among the participants with overweight or 142 obesity 26 . Finally, a recent meta-analysis of 43 studies confirmed this inverse association 143 between serum concentration of vitamin D and risk of metabolic syndrome in the general 144 adult population²⁷. In animals, vitamin D deficiency associates with worsened obesity 145 condition, glucose intolerance and insulin resistance induced by a high-fat diet whereas no 146 such effect for animals experimenting vitamin D deficiency under standard diet were 147 observed²⁸. Thus, it seems that inadequate levels of vitamin D in the context of obesity could worsen the individual metabolic health. Low levels of vitamin D are not only linked with different health conditions such as loss of bone density but also diabetes, high blood pressure, or multiple sclerosis. It also appears that vitamin D deficiency can impede bariatric surgery- induced metabolic improvements. Interestingly, in a mouse model of vertical sleeve gastrectomy, deficient levels of vitamin D were linked with decreased weight loss and glucose 153 homeostasis improvements, as well as intestinal inflammation and permeability 29 . In addition to those metabolic observations regarding vitamin D, many studies also reported strong links of this vitamin with adipose tissue in terms of vitamin D storage but also modulation of adipocyte differentiation or even energy metabolism and inflammation by this vitamin. These 157 aspects have been largely and well-reviewed recently.

Alteration of vitamin status in the context of bariatric surgery

 Vitamin deficiency in obesity has also been investigated in subjects with severe obesity undergoing bariatric surgery. For example, studies showed that the prevalence of vitamin B1 162 deficiency in patients with severe obesity before bariatric surgery is between 15.5% and 29% ³². In the general population, vitamin B1 deficiency is associated with inadequate intake and is more prevalent in underdeveloped countries whereas in industrialized countries, 165 vitamin B1 deficiency is rare and mostly related to alcoholism³³. Vitamin B9 has also been 166 found to be decreased in patients with severe obesity before the surgery at a rate of 63.2% ³⁴, 167 a much higher percentage in comparison to the general population prevalence of 2% reported 168 by a recent study in the US^{35} . Importantly, these studies focused on bariatric surgery candidates who follow weight-loss efforts for several months before the procedures, as 170 recommended by international guidelines³⁶. As such, these patients may have reduced vitamin intake due to changes in their diet. Flancbaum and colleagues also revealed a high

 proportion (68%) of patients presenting with a vitamin D deficiency before the surgery, which 173 was confirmed in two other cohorts with deficiency rates of 74.35% and 76% 38 . In comparison, a study of vitamin D blood concentrations in elderly people across 11 European countries reported a prevalence of vitamin D deficiency in 36% of men and 42 % of women in 176 the general population³⁹. This percentage was even lower in the US population with a 177 prevalence of vitamin D deficiency of 4.0% and 17.4% for inadequacy⁴⁰. To note, vitamin D blood concentration is dependent on the season as well as geographical location, which could explain such discrepancies between prevalence in different populations and studies. The two latter studies investigating vitamin D blood concentrations also reported vitamin B12 181 deficiency in those patients, reaching $12.3\%^{37}$ and $16\%^{38}$ respectively.

 Moreover, most vitamins are absorbed in the small intestine, a segment bypassed after malabsorptive procedures (e.g., Roux-en-Y Gastric Bypass or biliopancreatic derivation). This purposefully-induced malabsorption further deteriorates vitamin status after the surgical procedure, which may not normalize despite systematic vitamin supplementation. During surgical follow-up, monitoring patients' vitamin status should be reinforced, which is highlighted by studies of micronutrient deficiencies after Roux-en-Y gastric bypass as well as 188 sleeve gastrectomy^{41,42}. In fact, despite medical recommendation of micronutrients and especially multivitamins supplementation after the surgery, evidence of decreased compliance has been reported after 10 years of follow-up increasing the risk of vitamin 191 deficiency⁴³. It is also possible that the optimization of the pre-operative vitamin status could also contribute to better outcomes of the surgery. It has been suggested that vitamin D could, 193 for example, contribute to improved weight loss following bariatric surgery⁴⁴. A recent review addressed this concern in terms of vitamin status and the physiological, metabolic and also 195 bacterial changes associated with bariatric surgery . More attention should be given to

 vitamin status of patients before and after the bariatric surgery as suggested by recent 197 guidelines published by the British Obesity and Metabolic Surgery Society⁴⁶. The aforementioned studies suggest that acting on the pre-intervention vitamin deficiency might help potentialize metabolic improvement, although more clinical interventions ought to be performed to confirm this hypothesis.

Gut microbiota; a source of vitamin production

 The composition of the GM has now been meticulously studied over the last decade thanks to major advances in sequencing technologies. Nevertheless, a recent shift towards understanding the functionality of the ecosystem allows for more complete comprehension 206 of how GM modulations are involved in health and diseases. Gut bacteria produce a very large 207 number of metabolites having an impact on the host metabolism. The gut bacteria can 208 produce vitamin K and B vitamins⁴⁸; however the interest for the study of the relationship between GM and vitamin metabolism is only recent and increasing.

 The number of available bacterial genomes increased significantly with the advances in sequencing technologies and metagenomic processing methods, which have allowed for better characterization of GM species and their metabolic capabilities. In 2015, Magnúsdóttir et al., used the PubSEED platform, which provides genome annotations across thousands of 215 genomes, to analyze the ability of human GM organisms to synthesize B vitamins ⁴⁹. Thus, they were able to determine that the biosynthesis of B vitamins was common among the species found in the human GM. They also predicted B vitamin synthesis abilities that matched published experimental data. It appears that the synthesis of vitamin B2 and vitamin B3 were most commonly found among all the genomes assessed. Altogether, this study reinforces the

220 idea that the GM, as an ecosystem, is able to produce vitamins and possibly participates to 221 vitamin homeostasis of the host. Yet, this paper revealed that the production of vitamins by 222 the GM is not sufficient to cover host daily requirements. Moreover, the vitamins synthesized in the gut mostly benefit the neighboring non-producing bacteria through cross-feeding between organisms. To this end, bacterial genome analyses have illustrated that bacterial species can be vitamin prototrophs (capable of *de novo* synthesis) or auxotrophs (dependent 226 on the uptake and salvage of vitamins) highlighting the importance of cross-feeding between 227 $\,$ GM species⁵⁰.

 Further evidence of GM mutualism for vitamins comes from co-culture *in vitro* experiments showing that *Anaerobutyricum halii* is able to synthesize and provide 230 Akkermansia muciniphila with B12 vitamin, which is used for propionate production⁵¹. Such cross-feeding mechanisms have also been observed *in vivo,* where non-producing bacteria survive during limitation of vitamin intake suggesting the existence of vitamin sharing 233 between producing and non-producing bacteria⁵². Bacterial mutualism thus contributes to maintaining a relative GM stability thanks to the sharing of micronutrient resources in case of shortage or level modulation due to vitamin intake variability⁵² . Noteworthy *Anaerobutyricum halii* and *Akkermansia muciniphila* are both commensal bacteria associated with improved 237 metabolism^{53,54}.

 Together with experimental work, the access to genomic annotation allowed for the identification of specific vitamin producing bacterial strains. It is now known for example, that species from the genera *Lactobacillus* and *Bifidobacterium* can produce vitamin B9. Indeed, both *in vitro* and *in vivo* experiments demonstrated that Bifidobacterial strains are able to 242 increase the vitamin B9 concentration either in the medium or of the host ^{55,56}. This ability of vitamin B9 production by *Lactobacillus* and *Bifidobacterium* was assessed via the analysis of

 genome sequences. The presence of genes and enzymes involved in different steps of the vitamin B9 production pathway allowed to infer theoretical capacity of the studied strains to 246 produce this vitamin at least from a precursor molecule⁵⁷. Nevertheless, it is important to note 247 that the presence of genes involved in the production of a vitamin does not account for its effective production by the bacteria. Thus, experimental validation (*in vitro* and *in vivo)* remains crucial to identify potential vitamin-producing strains that could then be developed as probiotics.

 To explore the contribution of GM's vitamin synthesis, in various diseases including metabolic diseases, *in* vivo experiments are needed. Germ-Free (GF) rodents are an initial model used to determine the general contribution of the GM to metabolic pathways or host physiology. In the absence of GM in GF mice, a systemic deficiency in different vitamins was 255 reported^{58,59}, suggesting again that those vitamins can be produced by the GM and that microbial vitamin production contributes, at least partially, to vitamin homeostasis of the host. Among those vitamins, earlier studies showed that GF rats developed spontaneously a 258 vitamin K deficiency leading to increased mortality of the animals⁵⁸. The side effects of vitamin K deficiency were corrected by vitamin K1 supplementation and most interestingly with the gut colonization of single strains of bacteria (*Escherichia coli* or *Sarcina-like micrococcus*). Similar results have been found for B vitamins as rats fed a diet deprived of B vitamins stopped 262 growing after 1 to 3 weeks and died prematurely ⁵⁹. Yet, in this early study, one rat behaved differently and recovered from the deficiency. When its feces were added to the diet of another group of rats fed the same diet, the symptoms of vitamin B deficiency disappeared in those animals, suggesting that B vitamin-producing bacteria could be present in the feces these rats were given.

 Other evidence of B vitamin production by intestinal microorganisms have been reported. The total output of vitamin B7 measured in feces and urine has been shown to be 269 six times greater than the intake in humans, suggesting a production by the GM 60,61 while this needs further exploration. Interestingly, vitamin B7 deficiency induced alopecia only in GF 271 animals and not in conventionally raised animals suggesting that the GM contributes to this 272 vitamin metabolism⁶². A similar phenotype was observed in vitamin B7-deficient mice treated with an antibiotic, vancomycin, again suggesting a contribution of GM to vitamin B7 production. Moreover, those animals presented further reduction in bacterial diversity as well as major increase of *Lactobacillus* genus and in particular *Lactobacillus murinus* in comparison to antibiotic treated animals receiving sufficient levels of vitamin B7. Regarding other B 277 vitamins, GM depletion via antibiotic treatment resulted in a decrease of vitamin B6 levels in 278 colonic content in mice .

 These observations all identify the GM as a major contributor to the host's vitamin status. Nevertheless, the contribution of the GM's vitamin production to host vitamin circulating levels remains to be experimentally established, in healthy and disease conditions. Indeed, in the context of obesity, alteration in GM richness, diversity and functional anomalies 283 have repeatedly been reported $47,64-66$. Whether these alterations are linked to deficient bacterial vitamin metabolism and production need to be investigated in depth.

Consequence of vitamin deficiencies on GM composition

 As raised above, the production of vitamin K by the GM has been known for several decades; but the effect of its deficiency on the GM composition has been studied very recently. Thus, mice fed a vitamin K deficient diet displayed reduced abundance of

 Lactobacillus and enrichment in *Ruminococcus, Anaerostipes* genera as well as 291 Muribaculaceae family in comparison to animals supplemented with vitamin K ⁶⁷.

 A murine study investigating the effect of different micronutrient-deficient (vitamin A, vitamin B9, iron, zinc, or multiple deficiencies) diet on the GM revealed that the vitamin A 295 deficiency had the higher impact⁶⁸, although vitamin A is not produced by the GM. The analysis of the GM of mice fed a vitamin A deficient diet revealed an increase of *Bacteroides vulgatus* and an increase of *Bacteroides dorei.* Once the deficiency was corrected via a micronutrient sufficient supplementation of the diet, the initial levels of those bacteria were also restored. In a human study investigating the GM children with vitamin A deficiency, changes in the bacterial community composition were also reported⁶⁹ . Increased abundances of *Blautia* and *Faecalibacterium* genera and decreased abundances of *Bifidobacterium, Bacteroides* and *Shigella* were observed in these children. The modulations of *Bifidobacterium* and *Bacteroides* levels were even transmitted via fecal microbiota transfer of the children's feces into germ- free animals. At the phylum level, a decrease of *Bacteroidetes* was observed in mice fed a 305 vitamin A-deficient diet . This decrease is associated with an increase of the *Firmicutes/Bacteroidetes* ratio in those animals, which has been reported associated with 307 obesity and metabolic alterations⁷¹, despite discrepancies in recent studies^{72,73}. To be noted, in this study, vitamin A-deficient animals presented characteristics of pre-diabetes with hyperglycemia and delayed response to insulin injection, emphasizing the potential links with metabolic alterations.

 In the last decade, there has been an increasing number of studies investigating the links between vitamin D and the GM. One of them investigated the factors explaining the most

 the variance in alpha-diversity of the human GM, and found that the serum concentration of 315 1,25(OH)₂D was the most contributing variable, explaining 5% of the variance of the alpha- diversity⁷⁴. In terms of specific compositional changes associated with vitamin D deficiency, it seems that vitamin D deficiency (in models of knock-out mice for the vitamin D receptor (VDR), 318 not able to produce $1,25(OH)_2D$) appeared to be associated with a decrease of the abundance of the phylum *Firmicutes*⁷⁵ and the associated families *Ruminococcaceae, Lachnospiraceae, Lactobacillaceae* and *Streptococcaceae*. Interestingly, in another study on VDR KO mice, a significant decrease of the abundance of the genus *Lactobacillus* was observed in comparison 322 to wild-type animals 76 . In a human trial on older men, a positive association was also found between serum 1,25(OH)2D levels and the abundance of the genus *Ruminococcus* ⁷⁴ . On the contrary, vitamin D deficiency seems to be associated with an increase in *Bacteroidetes* and *Proteobacteria* phyla, and more precisely *Bacteroidaceae* and *Desulfovibrionaceae* families ⁷⁵. This increase was again confirmed in the study from Jin and colleagues, in which an increase 327 in the genus *Bacteroides* was noticed in VDR KO animals⁷⁶. In the murine study by Zhang *et al.*, investigating the effect of vitamin D deficiency on the metabolic improvements after vertical sleeve gastrectomy, they also analyzed the effects on the GM composition. The authors revealed that, despite known beneficial effects induced by the surgery itself, deficient levels of vitamin D were associated with limited improvements of the gut dysbiosis induced by the high-fat diet feeding. Indeed, the effects of the surgery, a decrease of *Firmicutes* and an increase of *Bacteroidetes* phyla abundance, were weaker in the group of vitamin-D-deficient 334 animals post-surgery in comparison to the group with normal levels of vitamin D^{29} .

 The effect of B vitamins deficiency on the GM has also been recently studied in the context of metabolic diseases. In a recent study, the effects of low B6 vitamin levels on host 337 phenotype and GM of weaning rats was explored⁷⁷. The animals receiving a standard chow

 diet with low levels of vitamin B6 displayed a decreased body weight and circulating triglycerides levels compared to control animals as well as changes in the GM composition. Indeed, an increase in *Lachnospiraceae* and a decrease of *Bacteroides* were observed in both male and female rats.

 In patients with obesity, an association was found between low niacin (vitamin B3) intake and reduced alpha-diversity as well as levels of Bacteroidetes. These findings were particularly significant in subjects with obesity and insulin-resistance but not with overt type 345 2 diabetes⁷⁸. The authors suggested that in patients with type 2 diabetes, medications such as 346 metformin known to modulate the $GM^{79,80}$ should be taken into account to explain those differences.

 These collective findings illustrate the need to investigate the interplay between GM, vitamin and host metabolism. Yet, few studies investigated the effects of vitamin deprivation on the GM composition in the context of obesity and metabolic disorders. Considering the links between obesity, GM dysbiosis and alteration of vitamin status as presented above, it is important to study these factors to decipher the mechanisms involved.

Effect of vitamin supplementation on GM

 Recent interest regarding the effect of vitamin supplementation on the composition of the GM has led to an increase of publications on this topic. A recent review extensively summarized the effects of vitamins and other dietary micronutrients such as polyphenols and 359 minerals on the composition of the GM^{81} . Although no clear patterns of GM modulation can be identified from this review due to a relatively low number of studies for some vitamins and

 a great variability in experimental designs, it seems that vitamin supplementation can induce significant changes in GM composition.

 In the context of obesity, the establishment of multiple large cohorts in the last decade has allowed the collection of important data sets of microbiome analysis, clinical and lifestyle information. Data from the American Gut Project were used to assess the impact of vitamin B and/or D supplementation on the GM in groups of people with normal weight (NW) or 367 overweight (OW) ⁸². Despite no significant impact on alpha-diversity indexes in all groups, the vitamin supplementation alone had almost no effect in NW group but had beneficial impact in OW. Noteworthy, vitamin B supplementation induced a significant decrease in *Tenericutes* at the phylum level and several groups at the family level (*Mogibacteriaceae, Porphyromonadaceae, Coriobacteriaceae* and *Comamonadaceae*). All these bacterial families have been associated with infections, inflammation and even obesity in the literature, suggesting that vitamin B supplementation in the context of obesity could help decrease the 374 abundance of pathogens in the GM⁸³⁻⁸⁶. Regarding vitamin D supplementation, effects on the GM composition were observed both in the NW and OW groups. In NW group, it induced a decrease of several families *Erysipelotrichaceae*, *Turicibacteraceae* and *Bifidobacteriaceae*. The same effect on this last beneficial family was observed among OW participants, as well as a decrease of pathogens (*Actinomycetaceae*) for this group. The effects of a combined vitamin B and D supplementation was also investigated and showed a decrease of *Lachnospiraceae* and *Enterobacteriaceae*, which have been shown to be associated with obesity and production 381 of endotoxins respectively $87,88$.

 Vitamin supplements, and in particular vitamin D, also had an important impact on the 383 functionality of the GM 82 . In NW groups, there was a decrease in genes implicated in amino-acids biosynthesis and an increase of vitamin biosynthesis (related to vitamin B1 and B5) in

 the GM of people supplemented with vitamin D. Those pathways were not impacted by vitamin D in the OW group. Here again, the association of the two vitamins had specific effect regarding the functional machinery of bacteria. The authors observed a decrease in the potential of amino acid consumption by GM and of the degradation of polysaccharides in the OW group. This paper supports the beneficial effects of vitamin D (alone or combined with vitamin B) on microbial communities in a context of overweight or obesity, although effects vary according to the severity of obesity.

 Another recent trial of vitamin D supplementation over 16 weeks in overweight and 393 patients with obesity showed modulation of the GM composition⁸⁹. Despite no effect on alpha-diversity, vitamin D supplementation was associated with an increase of *Lachnospira* genus and a decrease of the abundance of *Blautia* genus in comparison to the group receiving placebo. Moreover, the authors observed a significant correlation between serum levels of 25(OH)D and the abundance of *Coprococcus* genus. Yet, these studies remain quite small in size as only 17 people received vitamin D supplementation and 15 received the placebo in the 399 Naderpoor study⁸⁹. In the study from Jiang and colleagues, the groups of people with overweight supplemented with only vitamin D, or only vitamin B, or both vitamins comprised respectively 88, 16 and 214 individuals, and supplementation was based on dietary 402 supplement intakes self-reported by patients .

 In a murine study of weaning male C57BL/6J, the effect of vitamin A supplementation 404 in condition of standard chow diet (CD) or high fat high sugar diet (HFSD) was studied . Vitamin A was administered in the form of retinol, directly incorporated to the diet. After 9 weeks of follow-up, the supplementation with vitamin A (+VitA) limited HFSD-induced fat mass gain but was not able to reverse metabolic alterations induced by the diet. At the GM level, this study revealed that retinol supplementation prevented most of the compositional

 changes induced by HFSD. First, vitamin A supplementation protected against the decrease of alpha-diversity induced by HFSD, as HFSD+VitA animals had alpha-diversity values to the level of the CD group. In CD groups, a decrease of alpha-diversity was observed in the vitamin A 412 supplemented group, to the surprise of the authors. Second, HFSD-feeding led to a decrease of *Porphyromonadaceae* and *Mycoplasmataceae* and an increase of *Family XIII* in comparison to CD group. All these changes were prevented by administration of vitamin A, which in addition increased the abundance of *Lachnospiraceae*.

 These studies reinforce the links between vitamins and GM as oral vitamin supplementation is reported to induce modifications of the GM composition.

Future strategies of vitamin status management in obesity

 Knowing the alteration of vitamin status in some patients with obesity, the obesity- associated dysbiosis and relationships observed between GM and vitamin metabolism, several strategies can be considered to optimize vitamin levels subjects with obesity.

 Whereas obesity is often considered as a disease characterized by over-nutrition, it is also well recognized that there are issues of malnutrition, explaining at least partially inadequate vitamin intakes observed during obesity. Thus, a well-balanced and micronutrient- sufficient diet could help prevent deficiencies in some cases. In case of deficiency or sub- optimal levels, the oral vitamin supplementation also seems rational to rescue the host's vitamin status. However, most vitamins are absorbed in the small intestine and only small amounts of dietary intake reach the colonic bacterial ecosystem. Yet, as summarized in this review, many studies showed that vitamins can modulate the distal GM composition and have even been included in the definition of prebiotics by the International Scientific Association

 for Probiotics and Prebiotics (ISAPP) in 2016. Thus, delivery tools allowing for oral vitamin supplementation specifically targeting the colon might be useful. A recent control trial investigated such systems to administer several vitamins (vitamin C, B2, E, D and A) in 96 healthy volunteers and revealed modifications of the microbial composition and metabolic 437 activity such as an increased of bacterial diversity and fecal short-chain fatty acids⁹¹. At the species level, the authors observed an increase in *Bifidobacterium longum* and Anaerobutyricum hallii, both studied for their beneficial effects in several health conditions⁹¹. Similar studies in patients with severe obesity would be helpful to further investigate the effects of vitamin supplementation in this context, also taking into account the potential risk of over-dosing vitamin supplementation.

 Probiotic supplementation of strains known to produce vitamins can also be a strategy to rescue vitamin bacterial metabolism, altered in obesity. As reviewed by LeBlanc and colleagues, it has been demonstrated that specific bacterial strains are able to produce vitamins. For example, *Lactobacillus rhamnosus* GG can produce three B vitamins in culture 447 medium which are vitamin B1, B2 and B9⁹². Bifidobacterium adolescentis DSM 1835O⁹³ has been shown to increase vitamin B9 fecal levels in humans. For such a strategy, it is important to identify strains capable of complete *de novo* synthesis. As proposed by Engevik and colleagues, an alternative could be to rely on cross-feeding, mentioned above, and construct probiotic combinations of strains depending on one another for the production of 452 intermediates involved in the vitamin synthesis⁹⁴. It would be thus of interest to investigate how the host can benefit from these bacterial cross-talks. Yet, the long-term engraftment of probiotics remains a challenge for long-term therapeutic perspectives.

 Prebiotics can influence the growth and metabolism of specific gut bacteria, thus, their use to promote bacterial vitamin production could also be envisaged. A recent *in silico* study

 reported that inulin and short-chain fructo-oligosaccharides supplementation could increase 458 the production of an intermediate of the vitamin D biosynthesis pathway⁹⁵. This effect seems to be mediated by the secretion of short-chain fatty acids induced by the prebiotic 460 supplementation. The authors thus suggest that prebiotic supplementation could help decrease vitamin D deficiency linked to insufficiency of this intermediate in this study.

 Appropriate *in vivo* and clinicalstudies should be designed to confirm the potential use of these strategies in vitamin status management in severe obesity.

Conclusion

 Increasing evidence of sub-optimal or even deficient blood concentrations of a number 467 of vitamins in obesity and metabolic alterations have been consistently reported. There is therefore a clinical need to increase the awareness of medical care givers and patients regarding vitamin status, especially for long-term follow-up and in the context of nutritional 470 or surgical interventions. The reported evidence of modulations of both bacterial vitamin production and GM composition and functionality in condition of vitamin depletion and supplementation reinforce the links between vitamins and GM. In this direction, the GM dysbiosis observed in metabolic alterations and obesity could participate to the altered vitamin host status observed in several cohorts. The restoration of a functional ecosystem via a balanced diet, colon-targeted vitamin, probiotic or prebiotic supplementation of either vitamin-producing single strains or combination of strains relying on cross-feeding (Figure 1) 477 can thus contribute to an improved vitamin metabolism and status. The potential effects on metabolic health of such strategies should also be studied as well as the mechanisms involved in these interactions between vitamins, gut microbiota and metabolic health.

Acknowledgement

- The authors thank Leducq Foundation, the European commission (Join Program Initiative
- 483 (JPI)), the European Union's 7th Framework Program for research, technological development
- and demonstration (HEALTH-F4-2012-305312 METACARDIS), and the Fondation pour la
- Recherche Médicale (FDT202106012793) for supporting the research topic addressed in this
- 486 review. We thank Timothy Swartz for his thorough re-reading of this article.

Table 1 – Functions, recommended levels and deficiency consequences of B and D vitamins - Adapted from *Vitamin and mineral requirements in human nutrition* ⁹⁶

Figure 1 – **Intended strategies for host vitamin status improvement in obesity and metabolic alterations.** Improvement of vitamin metabolism can be envisioned via nutritional intervention with micronutrient sufficient and balanced diet, but also oral vitamin supplementation, for example via colon targeted system to deliver the vitamin directly to the gut microbial ecosystem and finally via probiotic administration of either single strains known to synthesize a specific vitamin or a bacterial community capable of cross-feeding to synthesize both vitamins and other metabolites such as short-chain fatty acids (SCFA) - *Created with BioRender.com*

References:

- 1. NCD Risk Factor Collaboration (NCD-RisC). Trends in adult body-mass index in 200 countries from 1975 to 2014: a pooled analysis of 1698 population-based measurement studies with 19·2 million participants. *Lancet Lond Engl*. 2016;387(10026):1377-1396. doi:10.1016/S0140- 6736(16)30054-X
- 2. NCD Risk Factor Collaboration (NCD-RisC). Worldwide trends in body-mass index, underweight, overweight, and obesity from 1975 to 2016: a pooled analysis of 2416 population-based measurement studies in 128·9 million children, adolescents, and adults. *Lancet Lond Engl*. 2017;390(10113):2627-2642. doi:10.1016/S0140-6736(17)32129-3
- 3. Kelly T, Yang W, Chen C-S, Reynolds K, He J. Global burden of obesity in 2005 and projections to 2030. *Int J Obes*. 2008;32(9):1431-1437. doi:10.1038/ijo.2008.102
- 4. Segula D. Complications of obesity in adults: A short review of the literature. *Malawi Med J*. 2014;26(1):20-24.
- 5. Crovesy L, Masterson D, Rosado EL. Profile of the gut microbiota of adults with obesity: a systematic review. *Eur J Clin Nutr*. 2020;74(9):1251-1262. doi:10.1038/s41430-020-0607-6
- 6. Valdes AM, Walter J, Segal E, Spector TD. Role of the gut microbiota in nutrition and health. *BMJ*. 2018;361:k2179. doi:10.1136/bmj.k2179
- 7. Huskisson E, Maggini S, Ruf M. The Role of Vitamins and Minerals in Energy Metabolism and Well-Being. *J Int Med Res*. 2007;35(3):277-289. doi:10.1177/147323000703500301
- 8. Depeint F, Bruce WR, Shangari N, Mehta R, O'Brien PJ. Mitochondrial function and toxicity: role of B vitamins on the one-carbon transfer pathways. *Chem Biol Interact*. 2006;163(1-2):113-132. doi:10.1016/j.cbi.2006.05.010
- 9. Porter K, Hoey L, Hughes CF, Ward M, McNulty H. Causes, Consequences and Public Health Implications of Low B-Vitamin Status in Ageing. *Nutrients*. 2016;8(11). doi:10.3390/nu8110725
- 10. Said HM. Intestinal absorption of water-soluble vitamins in health and disease. *Biochem J*. 2011;437(3):357-372. doi:10.1042/BJ20110326
- 11. European Food Safety Authority, ed. *Tolerable Upper Intake Levels for Vitamins and Minerals*. European Food Safety Authority; 2006.
- 12. Astrup A, Bügel S. Overfed but undernourished: recognizing nutritional inadequacies/deficiencies in patients with overweight or obesity. *Int J Obes*. 2019;43(2):219- 232. doi:10.1038/s41366-018-0143-9
- 13. Thomas-Valdés S, Tostes M das GV, Anunciação PC, da Silva BP, Sant'Ana HMP. Association between vitamin deficiency and metabolic disorders related to obesity. *Crit Rev Food Sci Nutr*. 2017;57(15):3332-3343. doi:10.1080/10408398.2015.1117413
- 14. Santacruz A, Collado MC, García-Valdés L, et al. Gut microbiota composition is associated with body weight, weight gain and biochemical parameters in pregnant women. *Br J Nutr*. 2010;104(1):83-92. doi:10.1017/S0007114510000176
- 15. Aasheim ET, Hofsø D, Hjelmesæth J, Birkeland KI, Bøhmer T. Vitamin status in morbidly obese patients: a cross-sectional study. *Am J Clin Nutr*. 2008;87(2):362-369. doi:10.1093/ajcn/87.2.362
- 16. Thornalley PJ, Babaei-Jadidi R, Al Ali H, et al. High prevalence of low plasma thiamine concentration in diabetes linked to a marker of vascular disease. *Diabetologia*. 2007;50(10):2164-2170. doi:10.1007/s00125-007-0771-4
- 17. Baltaci D, Kutlucan A, Turker Y, et al. Association of vitamin B12 with obesity, overweight, insulin resistance and metabolic syndrome, and body fat composition; primary care-based study. *Med Glas Off Publ Med Assoc Zenica-Doboj Cant Bosnia Herzeg*. 2013;10(2):203-210.
- 18. Geiker NRW, Veller M, Kjoelbaek L, et al. Effect of low energy diet for eight weeks to adults with overweight or obesity on folate, retinol, vitamin B12, D and E status and the degree of inflammation: a post hoc analysis of a randomized intervention trial. *Nutr Metab*. 2018;15. doi:10.1186/s12986-018-0263-1
- 19. Allen LH. How common is vitamin B-12 deficiency? *Am J Clin Nutr*. 2009;89(2):693S-696S. doi:10.3945/ajcn.2008.26947A
- 20. Järvinen E, Ismail K, Muniandy M, et al. Biotin-dependent functions in adiposity: a study of monozygotic twin pairs. *Int J Obes*. 2016;40(5):788-795. doi:10.1038/ijo.2015.237
- 21. Maebashi M, Makino Y, Furukawa Y, Ohinata K, Kimura S, Sato T. Therapeutic Evaluation of the Effect of Biotin on Hyperglycemia in Patients with Non-Insulin Dependent Diabetes Mellitus. *J Clin Biochem Nutr*. 1993;14:211-218. doi:10.3164/jcbn.14.211
- 22. Liel Y, Ulmer E, Shary J, Hollis BW, Bell NH. Low circulating vitamin D in obesity. *Calcif Tissue Int*. 1988;43(4):199-201. doi:10.1007/BF02555135
- 23. Oliai Araghi S, van Dijk SC, Ham AC, et al. BMI and body fat mass is inversely associated with vitamin D levels in older individuals. *J Nutr Health Aging*. 2015;19(10):980-985. doi:10.1007/s12603-015-0657-y
- 24. Botella-Carretero JI, Alvarez-Blasco F, Villafruela JJ, Balsa JA, Vázquez C, Escobar-Morreale HF. Vitamin D deficiency is associated with the metabolic syndrome in morbid obesity. *Clin Nutr Edinb Scotl*. 2007;26(5):573-580. doi:10.1016/j.clnu.2007.05.009
- 25. Devaraj S, Jialal G, Cook T, Siegel D, Jialal I. Low Vitamin D Levels in Northern American Adults with the Metabolic Syndrome. *Horm Metab Res*. 2011;43(1):72-74. doi:10.1055/s-0030- 1268485
- 26. Lu L, Yu Z, Pan A, et al. Plasma 25-Hydroxyvitamin D Concentration and Metabolic Syndrome Among Middle-Aged and Elderly Chinese Individuals. *Diabetes Care*. 2009;32(7):1278-1283. doi:10.2337/dc09-0209
- 27. Hajhashemy Z, Shahdadian F, Moslemi E, Mirenayat FS, Saneei P. Serum vitamin D levels in relation to metabolic syndrome: A systematic review and dose–response meta-analysis of epidemiologic studies. *Obes Rev*. 2021;22(7):e13223. doi:10.1111/obr.13223
- 28. Su D, Nie Y, Zhu A, et al. Vitamin D Signaling through Induction of Paneth Cell Defensins Maintains Gut Microbiota and Improves Metabolic Disorders and Hepatic Steatosis in Animal Models. *Front Physiol*. 2016;7:498. doi:10.3389/fphys.2016.00498
- 29. Zhang J, Feng M, Pan L, et al. Effects of vitamin D deficiency on the improvement of metabolic disorders in obese mice after vertical sleeve gastrectomy. *Sci Rep*. 2021;11(1):6036. doi:10.1038/s41598-021-85531-9
- 30. Park CY, Han SN. The Role of Vitamin D in Adipose Tissue Biology: Adipocyte Differentiation, Energy Metabolism, and Inflammation. *J Lipid Atheroscler*. 2021;10(2):130. doi:10.12997/jla.2021.10.2.130
- 31. Carrodeguas L, Kaidar-Person O, Szomstein S, Antozzi P, Rosenthal R. Preoperative thiamine deficiency in obese population undergoing laparoscopic bariatric surgery. *Surg Obes Relat Dis Off J Am Soc Bariatr Surg*. 2005;1(6):517-522; discussion 522. doi:10.1016/j.soard.2005.08.003
- 32. Flancbaum L, Belsley S, Drake V, Colarusso T, Tayler E. Preoperative nutritional status of patients undergoing Roux-en-Y gastric bypass for morbid obesity. *J Gastrointest Surg Off J Soc Surg Aliment Tract*. 2006;10(7):1033-1037. doi:10.1016/j.gassur.2006.03.004
- 33. Fattal-Valevski A. Thiamine (Vitamin B1). *J Evid-Based Complement Altern Med*. 2011;16(1):12- 20. doi:10.1177/1533210110392941
- 34. Krzizek E-C, Brix JM, Herz CT, et al. Prevalence of Micronutrient Deficiency in Patients with Morbid Obesity Before Bariatric Surgery. *Obes Surg*. 2018;28(3):643-648. doi:10.1007/s11695- 017-2902-4
- 35. Diaz K, Na Z, Gupta S, et al. Prevalence of Folic Acid Deficiency and Cost Effectiveness of Folic Acid Testing: A Single Center Experience. *Blood*. 2018;132(Supplement 1):4878-4878. doi:10.1182/blood-2018-99-111607
- 36. Mechanick JI, Apovian C, Brethauer S, et al. Clinical practice guidelines for the perioperative nutrition, metabolic, and nonsurgical support of patients undergoing bariatric procedures - 2019 update: cosponsored by American Association of Clinical Endocrinologists/American College of Endocrinology, The Obesity Society, American Society for Metabolic & Bariatric Surgery, Obesity Medicine Association, and American Society of Anesthesiologists. *Surg Obes Relat Dis Off J Am Soc Bariatr Surg*. 2020;16(2):175-247. doi:10.1016/j.soard.2019.10.025
- 37. Arias PM, Domeniconi EA, García M, Esquivel CM, Martínez Lascano F, Foscarini JM. Micronutrient Deficiencies After Roux-en-Y Gastric Bypass: Long-Term Results. *Obes Surg*. 2020;30(1):169-173. doi:10.1007/s11695-019-04167-x
- 38. Al-Mutawa A, Anderson AK, Alsabah S, Al-Mutawa M. Nutritional Status of Bariatric Surgery Candidates. *Nutrients*. 2018;10(1):67. doi:10.3390/nu10010067
- 39. van der Wielen RP, Löwik MR, van den Berg H, et al. Serum vitamin D concentrations among elderly people in Europe. *Lancet Lond Engl*. 1995;346(8969):207-210. doi:10.1016/s0140- 6736(95)91266-5
- 40. Orces C, Lorenzo C, Guarneros JE. The Prevalence and Determinants of Vitamin D Inadequacy among U.S. Older Adults: National Health and Nutrition Examination Survey 2007-2014. *Cureus*. 11(8):e5300. doi:10.7759/cureus.5300
- 41. Verger EO, Aron-Wisnewsky J, Dao MC, et al. Micronutrient and Protein Deficiencies After Gastric Bypass and Sleeve Gastrectomy: a 1-year Follow-up. *Obes Surg*. 2016;26(4):785-796. doi:10.1007/s11695-015-1803-7
- 42. Ha J, Kwon Y, Kwon J-W, et al. Micronutrient status in bariatric surgery patients receiving postoperative supplementation per guidelines: Insights from a systematic review and metaanalysis of longitudinal studies. *Obes Rev*. 2021;22(7):e13249. doi:10.1111/obr.13249
- 43. Mehaffey JH, Mehaffey RL, Mullen MG, et al. Nutrient Deficiency 10 Years Following Roux-en-Y Gastric Bypass: Who's Responsible? *Obes Surg*. 2017;27(5):1131-1136. doi:10.1007/s11695- 016-2364-0
- 44. Slusher AL, McAllister MJ, Huang C-J. A therapeutic role for vitamin D on obesity-associated inflammation and weight-loss intervention. *Inflamm Res*. 2015;64(8):565-575. doi:10.1007/s00011-015-0847-4
- 45. Aron-Wisnewsky J, Clément K. A place for vitamin supplementation and functional food in bariatric surgery? *Curr Opin Clin Nutr Metab Care*. 2019;22(6):442-448. doi:10.1097/MCO.0000000000000602
- 46. O'Kane M, Parretti HM, Pinkney J, et al. British Obesity and Metabolic Surgery Society Guidelines on perioperative and postoperative biochemical monitoring and micronutrient replacement for patients undergoing bariatric surgery—2020 update. *Obes Rev*. 2020;21(11):e13087. doi:10.1111/obr.13087
- 47. Agus A, Clément K, Sokol H. Gut microbiota-derived metabolites as central regulators in metabolic disorders. *Gut*. 2021;70(6):1174-1182. doi:10.1136/gutjnl-2020-323071
- 48. Hill MJ. Intestinal flora and endogenous vitamin synthesis: *Eur J Cancer Prev*. 1997;6:S43-S45. doi:10.1097/00008469-199703001-00009
- 49. Magnúsdóttir S, Ravcheev D, de Crécy-Lagard V, Thiele I. Systematic genome assessment of Bvitamin biosynthesis suggests co-operation among gut microbes. *Front Genet*. 2015;6:148. doi:10.3389/fgene.2015.00148
- 50. Rodionov DA, Arzamasov AA, Khoroshkin MS, et al. Micronutrient Requirements and Sharing Capabilities of the Human Gut Microbiome. *Front Microbiol*. 2019;10. doi:10.3389/fmicb.2019.01316
- 51. Belzer C, Chia LW, Aalvink S, et al. Microbial Metabolic Networks at the Mucus Layer Lead to Diet-Independent Butyrate and Vitamin B12 Production by Intestinal Symbionts. *mBio*. 2017;8(5). doi:10.1128/mBio.00770-17
- 52. Sharma V, Rodionov DA, Leyn SA, et al. B-Vitamin Sharing Promotes Stability of Gut Microbial Communities. *Front Microbiol*. 2019;10. doi:10.3389/fmicb.2019.01485
- 53. Cani PD, de Vos WM. Next-Generation Beneficial Microbes: The Case of Akkermansia muciniphila. *Front Microbiol*. 2017;8. doi:10.3389/fmicb.2017.01765
- 54. Udayappan S, Manneras-Holm L, Chaplin-Scott A, et al. Oral treatment with Eubacterium hallii improves insulin sensitivity in db/db mice. *Npj Biofilms Microbiomes*. 2016;2(1):1-10. doi:10.1038/npjbiofilms.2016.9
- 55. Pompei A, Cordisco L, Amaretti A, Zanoni S, Matteuzzi D, Rossi M. Folate Production by Bifidobacteria as a Potential Probiotic Property. *Appl Environ Microbiol*. 2007;73(1):179-185. doi:10.1128/AEM.01763-06
- 56. Pompei A, Cordisco L, Amaretti A, et al. Administration of Folate-Producing Bifidobacteria Enhances Folate Status in Wistar Rats. *J Nutr*. 2007;137(12):2742-2746. doi:10.1093/jn/137.12.2742
- 57. Rossi M, Amaretti A, Raimondi S. Folate production by probiotic bacteria. *Nutrients*. 2011;3(1):118-134. doi:10.3390/nu3010118
- 58. Gustafsson BE, Daft FS, McDaniel EG, Smith JC, Fitzgerald RJ. Effects of Vitamin K-Active Compounds and Intestinal Microorganisms in Vitamin K-Deficient Germfree Rats. *J Nutr*. 1962;78(4):461-468. doi:10.1093/jn/78.4.461
- 59. Fridericia LS, Freudenthal P, Gudjonnsson S, Johansen G, Schoubye N. Refection, a Transmissible Change in the Intestinal Content, enabling Rats to grow and thrive without Vitamin B in the Food. *Epidemiology & Infection*. Published online 1927. doi:10.1017/S002217240003182X
- 60. Bender D. *Nutritional Biochemistry of the Vitamins*.; 2003. doi:10.1017/CBO9780511615191
- 61. Oppel TW. Studies of Biotin Metabolism in Man. Part I. The Excretion of Biotin in, Human Urine. *Am J Med Sci*. 1942;204(6):856-875.
- 62. Hayashi A, Mikami Y, Miyamoto K, et al. Intestinal Dysbiosis and Biotin Deprivation Induce Alopecia through Overgrowth of Lactobacillus murinus in Mice. *Cell Rep*. 2017;20(7):1513-1524. doi:10.1016/j.celrep.2017.07.057
- 63. Miki T, Goto R, Fujimoto M, Okada N, Hardt W-D. The Bactericidal Lectin RegIIIβ Prolongs Gut Colonization and Enteropathy in the Streptomycin Mouse Model for Salmonella Diarrhea. *Cell Host Microbe*. 2017;21(2):195-207. doi:10.1016/j.chom.2016.12.008
- 64. Le Chatelier E, Nielsen T, Qin J, et al. Richness of human gut microbiome correlates with metabolic markers. *Nature*. 2013;500(7464):541-546. doi:10.1038/nature12506
- 65. Cotillard A, Kennedy SP, Kong LC, et al. Dietary intervention impact on gut microbial gene richness. *Nature*. 2013;500(7464):585-588. doi:10.1038/nature12480
- 66. Aron-Wisnewsky J, Prifti E, Belda E, et al. Major microbiota dysbiosis in severe obesity: fate after bariatric surgery. *Gut*. 2019;68(1):70-82. doi:10.1136/gutjnl-2018-316103
- 67. Ellis JL, Karl JP, Oliverio AM, et al. Dietary vitamin K is remodeled by gut microbiota and influences community composition. *Gut Microbes*. 2021;13(1):1-16. doi:10.1080/19490976.2021.1887721
- 68. Hibberd MC, Wu M, Rodionov DA, et al. The effects of micronutrient deficiencies on bacterial species from the human gut microbiota. *Sci Transl Med*. 2017;9(390). doi:10.1126/scitranslmed.aal4069
- 69. Feng D, Chen B, Zeng B, et al. Faecal microbiota from children with vitamin A deficiency impairs colonic barrier function in germ-free mice: the possible role of alterative bile acid metabolites. *Nutrition*. Published online April 20, 2021:111274. doi:10.1016/j.nut.2021.111274
- 70. Tian Y, Nichols RG, Cai J, Patterson AD, Cantorna MT. Vitamin A deficiency in mice alters host and gut microbial metabolism leading to altered energy homeostasis. *J Nutr Biochem*. 2018;54:28-34. doi:10.1016/j.jnutbio.2017.10.011
- 71. Ley RE, Turnbaugh PJ, Klein S, Gordon JI. Microbial ecology: human gut microbes associated with obesity. *Nature*. 2006;444(7122):1022-1023. doi:10.1038/4441022a
- 72. Jumpertz R, Le DS, Turnbaugh PJ, et al. Energy-balance studies reveal associations between gut microbes, caloric load, and nutrient absorption in humans123. *Am J Clin Nutr*. 2011;94(1):58-65. doi:10.3945/ajcn.110.010132
- 73. Duncan SH, Lobley GE, Holtrop G, et al. Human colonic microbiota associated with diet, obesity and weight loss. *Int J Obes*. 2008;32(11):1720-1724. doi:10.1038/ijo.2008.155
- 74. Thomas RL, Jiang L, Adams JS, et al. Vitamin D metabolites and the gut microbiome in older men. *Nat Commun*. 2020;11(1):5997. doi:10.1038/s41467-020-19793-8
- 75. Ooi JH, Li Y, Rogers CJ, Cantorna MT. Vitamin D Regulates the Gut Microbiome and Protects Mice from Dextran Sodium Sulfate–Induced Colitis123. *J Nutr*. 2013;143(10):1679-1686. doi:10.3945/jn.113.180794
- 76. Jin D, Wu S, Zhang Y-G, et al. Lack of Vitamin D Receptor Causes Dysbiosis and Changes the Functions of the Murine Intestinal Microbiome. *Clin Ther*. 2015;37(5):996-1009.e7. doi:10.1016/j.clinthera.2015.04.004
- 77. Mayengbam S, Chleilat F, Reimer RA. Dietary Vitamin B6 Deficiency Impairs Gut Microbiota and Host and Microbial Metabolites in Rats. *Biomedicines*. 2020;8(11). doi:10.3390/biomedicines8110469
- 78. Fangmann D, Theismann E-M, Türk K, et al. Targeted Microbiome Intervention by Microencapsulated Delayed-Release Niacin Beneficially Affects Insulin Sensitivity in Humans. *Diabetes Care*. 2018;41(3):398-405. doi:10.2337/dc17-1967
- 79. Forslund K, Hildebrand F, Nielsen T, et al. Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota. *Nature*. 2015;528(7581):262-266. doi:10.1038/nature15766
- 80. Sun L, Xie C, Wang G, et al. Gut microbiota and intestinal FXR mediate the clinical benefits of metformin. *Nat Med*. Published online November 5, 2018:1. doi:10.1038/s41591-018-0222-4
- 81. Yang Q, Liang Q, Balakrishnan B, Belobrajdic DP, Feng Q-J, Zhang W. Role of Dietary Nutrients in the Modulation of Gut Microbiota: A Narrative Review. *Nutrients*. 2020;12(2). doi:10.3390/nu12020381
- 82. Jiang S, Zhu Q, Mai M, Yang W, Du G. Vitamin B and vitamin D as modulators of gut microbiota in overweight individuals. *Int J Food Sci Nutr*. 2020;71(8):1001-1009. doi:10.1080/09637486.2020.1748580
- 83. Almuzara M, Cittadini R, Estraviz ML, Ellis A, Vay C. First report of Comamonas kerstersii causing urinary tract infection. *New Microbes New Infect*. 2018;24:4-7. doi:10.1016/j.nmni.2018.03.003
- 84. Giannelli V, Di Gregorio V, Iebba V, et al. Microbiota and the gut-liver axis: Bacterial translocation, inflammation and infection in cirrhosis. *World J Gastroenterol WJG*. 2014;20(45):16795-16810. doi:10.3748/wjg.v20.i45.16795
- 85. Clavel T, Duck W, Charrier C, Wenning M, Elson C, Haller D. Enterorhabdus caecimuris sp. nov., a member of the family Coriobacteriaceae isolated from a mouse model of spontaneous colitis, and emended description of the genus Enterorhabdus Clavel et al. 2009. *Int J Syst Evol Microbiol*. 2010;60(Pt 7):1527-1531. doi:10.1099/ijs.0.015016-0
- 86. Wu Y, Chi X, Zhang Q, Chen F, Deng X. Characterization of the salivary microbiome in people with obesity. *PeerJ*. 2018;6:e4458. doi:10.7717/peerj.4458
- 87. Kameyama K, Itoh K. Intestinal colonization by a Lachnospiraceae bacterium contributes to the development of diabetes in obese mice. *Microbes Environ*. 2014;29(4):427-430. doi:10.1264/jsme2.ME14054
- 88. Xie J, Peters BM, Li B, et al. Clinical features and antimicrobial resistance profiles of important Enterobacteriaceae pathogens in Guangzhou representative of Southern China, 2001-2015. *Microb Pathog*. 2017;107:206-211. doi:10.1016/j.micpath.2017.03.038
- 89. Naderpoor N, Mousa A, Fernanda Gomez Arango L, Barrett HL, Dekker Nitert M, de Courten B. Effect of Vitamin D Supplementation on Faecal Microbiota: A Randomised Clinical Trial. *Nutrients*. 2019;11(12). doi:10.3390/nu11122888
- 90. Biyong EF, Alfos S, Dumetz F, et al. Dietary vitamin A supplementation prevents early obesogenic diet-induced microbiota, neuronal and cognitive alterations. *Int J Obes*. Published online November 22, 2020:1-11. doi:10.1038/s41366-020-00723-z
- 91. Pham VT, Fehlbaum S, Seifert N, et al. Effects of colon-targeted vitamins on the composition and metabolic activity of the human gut microbiome– a pilot study. *Gut Microbes*. 2021;13(1):1-20. doi:10.1080/19490976.2021.1875774
- 92. LeBlanc JG, Chain F, Martín R, Bermúdez-Humarán LG, Courau S, Langella P. Beneficial effects on host energy metabolism of short-chain fatty acids and vitamins produced by commensal and probiotic bacteria. *Microb Cell Factories*. 2017;16(1):79. doi:10.1186/s12934-017-0691-z
- 93. Strozzi GP, Mogna L. Quantification of Folic Acid in Human Feces After Administration of Bifidobacterium Probiotic Strains: *J Clin Gastroenterol*. 2008;42:S179-S184. doi:10.1097/MCG.0b013e31818087d8
- 94. Engevik MA, Morra CN, Röth D, et al. Microbial Metabolic Capacity for Intestinal Folate Production and Modulation of Host Folate Receptors. *Front Microbiol*. 2019;10:2305. doi:10.3389/fmicb.2019.02305
- 95. Gokhale S, Bhaduri A. Provitamin D 3 modulation through prebiotics supplementation: simulation based assessment. *Sci Rep*. 2019;9(1):19267. doi:10.1038/s41598-019-55699-2
- 96. World Health Organization, Food and Agriculture Organization of the United Nations, eds. *Vitamin and Mineral Requirements in Human Nutrition*. 2nd ed. World Health Organization ; FAO; 2004.