

Kaposi's Sarcoma in Virally Suppressed People Living with HIV: An Emerging Condition

Romain Palich, Alain Makinson, Marianne Veyri, Amélie Guihot, Marc-Antoine Valantin, Sylvie Brégigeon-Ronot, Isabelle Poizot-Martin, Caroline Solas, Sophie Grabar, Guillaume Martin-Blondel, et al.

▶ To cite this version:

Romain Palich, Alain Makinson, Marianne Veyri, Amélie Guihot, Marc-Antoine Valantin, et al.. Kaposi's Sarcoma in Virally Suppressed People Living with HIV: An Emerging Condition. Cancers, 2021, 13 (22), pp.5702. 10.3390/cancers13225702 . hal-03456567

HAL Id: hal-03456567 https://hal.sorbonne-universite.fr/hal-03456567v1

Submitted on 30 Nov 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.





Kaposi's Sarcoma in Virally Suppressed People Living with HIV: An Emerging Condition

Romain Palich ^{1,*}, Alain Makinson ², Marianne Veyri ³, Amélie Guihot ⁴, Marc-Antoine Valantin ¹, Sylvie Brégigeon-Ronot ⁵, Isabelle Poizot-Martin ⁵, Caroline Solas ⁶, Sophie Grabar ⁷, Guillaume Martin-Blondel ⁸, and Jean-Philippe Spano ³

- ¹ Department of Infectious Diseases, Pitié-Salpêtrière Hospital, AP-HP, Pierre Louis Epidemiology and Public Health Institute (iPLESP), INSERM U1136, Sorbonne University, 75013 Paris, France; marc-antoine.valantin@aphp.fr
- ² Infectious Diseases Department, INSERM U1175, University Hospital of Montpellier, 34000 Montpellier, France; a-makinson@chu-montpellier.fr
- ³ Department of Medical Oncology, Pitié Salpêtrière Hospital, AP-HP, Institut Universitaire de Cancérologie (IUC), CLIP² Galilée, Pierre Louis Epidemiology and Public Health Institute (iPLESP), INSERM U1136, Sorbonne University, 75013 Paris, France; marianne.veyri@aphp.fr (M.V.); jean-philippe.spano@aphp.fr (J.-P.S.)
- ⁴ Department of Immunology, Pitié-Salpêtrière Hospital, AP-HP, Centre d'Immunologie et des Maladies Infectieuses, INSERM U1135, Sorbonne University, 75013 Paris, France; amelie.guihot@aphp.fr
- ⁵ Service d'Immuno-hématologie Clinique, Sciences Economiques & Sociales de la Santé & Traitement de l'Information Médicale, AP-HM, INSERM, IRD, Sciences Economiques et Sociales de la Santé et Traitement de l'Information Médicale, Hôpital Sainte-Marguerite, Aix Marseille University, 13007 Marseille, France; sylvie.ronot@ap-hm.fr (S.B.-R.); isabelle.poizot@ap-hm.fr (I.P.-M.)
- Department of Pharmacology Toxicology, AP-HM, Centre de Recherche en Cancérologie de Marseille, Hospital de la Timone, INSERM, Aix-Marseille University, 13005 Marseille, France; caroline.solas@ap-hm.fr
- ⁷ Pierre Louis Epidemiology and Public Health Institute (iPLESP), INSERM U1136, Sorbonne University,
- 75013 Paris, France; sophie.grabar@aphp.fr
- ⁸ Department of Infectious and Tropical Diseases, Centre de Physiopathologie Toulouse-Purpan, Toulouse University Hospital, INSERM U1043, CNRS UMR 5282, 31300 Toulouse, France; martin-blondel.g@chu-toulouse.fr
- * Correspondence: romain.palich@aphp.fr; Tel.: +33-142-160-171

Simple Summary: Kaposi's sarcoma (KS) in people living with HIV (PLHIV) occurs in the vast majority of cases when viral replication is not controlled and when CD4 immunosuppression is important. However, clinicians are observing more and more cases of KS in PLHIV with suppressed viremia on antiretroviral treatment. These clinical forms seem less aggressive, but cause therapeutic dead ends. Indeed, despite repeated chemotherapy, recurrences are frequent. Immunotherapy and specific treatment regimens should be evaluated in this population.

Abstract: Since the advent of highly effective combined antiretroviral treatment (cART), and with the implementation of large HIV testing programs and universal access to cART, the burden of AIDS-related comorbidities has dramatically decreased over time. The incidence of Kaposi's sarcoma (SK), strongly associated with HIV replication and CD4 immunosuppression, was greatly reduced. However, KS remains the most common cancer in patients living with HIV (PLHIV). HIV physicians are increasingly faced with KS in virally suppressed HIV-patients, as reflected by increasing description of case series. Though SK seem less aggressive than those in PLHIV with uncontrolled HIV-disease, some may require systemic chemotherapy. Persistent lack of specific anti-HHV-8 cellular immunity could be involved in the physiopathology of these KS. These clinical forms are a real therapeutic challenge without possible short-term improvement of anti-HHV-8 immunity, and no active replication of HIV to control. The cumulative toxicity of chemotherapies repeatedly leads to a therapeutic dead end. The introduction or maintenance of protease inhibitors in cART does not seem to have an impact on the evolution of these KS. Research programs in this emerging condition are important to consider new strategies.



Citation: Palich, R.; Makinson, A.; Veyri, M.; Guihot, A.; Valantin, M.-A.; Brégigeon-Ronot, S.; Poizot-Martin, I.; Solas, C.; Grabar, S.; Martin-Blondel, G.; et al. Kaposi's Sarcoma in Virally Suppressed People Living with HIV: An Emerging Condition. *Cancers* 2021, 13, 5702. https://doi.org/10.3390/ cancers13225702

Academic Editor: Naoki Mori

Received: 19 October 2021 Accepted: 9 November 2021 Published: 15 November 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Keywords: Kaposi's sarcoma; HIV; AIDS; antiretroviral; cancer

1. Introduction

Thanks to the efficacy and wide access of combined antiretroviral treatment (cART) for controlling HIV replication, the risk of Kaposi's sarcoma (KS) in people living with HIV (PLHIV) has greatly declined over the past 25 years in resource rich settings [1–4]. However, KS still remains, with non-Hodgkin lymphoma (NHL) the most common cancer among PLHIV [2,3]. KS frequently occurs in uncontrolled HIV infection, but there are increasing descriptions of KS occurring in virally suppressed patients, even in those with apparent T CD4 immune restoration. In virally suppressed patients, KS may present as a de novo complication, or as a recurrent disease, with repeated episodes over time.

There are very few published case series of virologically controlled patients presenting with KS (Table 1) [5–8]. There is also no standardized definition of cases in terms of HIV-RNA duration of suppression and HIV-RNA and CD4 thresholds, hindering comparability between studies. Our group assumes that KS in virally suppressed patients is an emerging complication in PLHIV, with specific clinical, physiopathological and therapeutic difficulties, justifying a specific review to encourage further research. Thus, we aim to provide an overview of KS in virally suppressed patients on the (1) latest available epidemiological data, (2) reported clinical features, (3) immunopathological pathways, (4) therapeutic issues, including the impact of protease inhibitors.

| Reference | Country | Number of Subjects | HIV-RNA Value Restrictions for Cases to Be Included | CD4 Count Restrictions for Cases to Be Included | Median CD4 Count | Median CD4 Nadir | cART Conditions to Be Included | Median Duration on cART | Types of cART |
|-----------|---------|-----------------------|---|--|---|---------------------------------------|-----------------------------------|----------------------------|---|
| [5] | USA | 9 | <300 copies/mL for at least 2 years | \geq 300/mm ³ | - | 340/mm ³ (range: 90–455) | - | 7 years (range: <1–19) | PI-based or NNRTI-based therapy |
| [6] | USA | 20 | <75 c opies/mL | \geq 300/mm ³ | 483/mm ³ (range: 300–625) | 216/mm ³ (range: 4–431) | - | 5 years (range: 1–12) | PI-based ($n = 11, 55\%$) or NNRTI-based ($n = 9, 45\%$) therapy |
| [7] | France | 21 | <50 copies/mL | - | 449/mm ³ (IQR: 241–625) | 196/mm ³ (IQR: 84–329) | \geq 12 months | - | PI-based (<i>n</i> = 4, 19%) or NNRTI-based (<i>n</i> = 7, 33%) or INSTI-based (<i>n</i> = 10, 48%) therapy |
| [8] | France | 12 | <50 copies/mL for at least 12 months | - | 723/mm ³ (range: 520–881) | - | - | - | - |

Table 1. Main published series including Kaposi's sarcoma in HIV-patients with suppressed viremia.

PI, protease inhibitor. NNRTI, non-nucleoside reverse transcriptase inhibitor. INSTI, integrase stand transfer inhibitor.

2. Epidemiology of KS in Virally Suppressed HIV-Patients

Few observational cohorts have specifically studied the risk of KS in PLHIV with suppressed viremia. In the French ANRS CO4 FHDH cohort, PLHIV with undetectable plasma HIV-RNA and restored immunity (i.e., $CD4 \ge 500/mm^3$ for at least 2 years) still had a 35-fold higher risk of KS compared to the general population (standardized incidence ratio (SIR) = 35.4; 95% CI 18.3–61.9) [3]. Interestingly, in the same population, this high risk was in contrast to the NHL risk, another virus-related AIDS defining cancer (SIR = 1; 95% CI 4.0–1.8), suggesting a specific susceptibility to KS in PLHIV despite apparent immune and virological control. However, despite a follow-up of 55,633 person-years, observed events of KS were rare (n = 12) in PLHIV with controlled disease. More recently, the US Veteran cohort examined the risk of cancers from 1999 to 2015, and found that the risk of KS as compared to uninfected patients was still more than 50-fold in PLHIV despite long-term viral suppression (≥ 2 years) and more than 500-fold in those with early suppression (< 2 years) [9].

KS has traditionally been described as occurring at low CD4 levels but the clinical context in which KS can occur have dramatically changed over the years and have impacted the pattern of presentation of KS. In the large collaboration of European cohorts, based on 1323 KS occurring at CD4 count \geq 200/mm³, the incidence rate was 1.2 per 1000 (95%CI 1.1–1.2) and gradually decreased with increasing CD4 levels [10]. However, an increasing number of studies have reported greater proportion of KS occurring at higher CD4 cell counts [1,11,12] and some case studies reported KS occurring in aviremic patients (Table 1). In a study gathering data from 8 American cohorts, the authors showed that across 1996– 2011 KS occurred at higher CD4 levels and lower VL [12]. Between 2007 and 2011, 15% of the KS occurred at CD4 count \geq 500/mm³ and less than one half <200/mm³. They also showed that this trend was mainly explained by the increasing proportion of the underlying HIV population on effective cART who exhibited higher CD4 and suppressed plasma HIV-RNA, and not by an increased risk of KS within each stratum. Taken together, epidemiological data describes rare but possible KS occurrence in PLHIV with immune restoration and virological control, though numbers remain low. However, the increasing number of publications describing these cases underscore a potential increase of this condition with the aging of PLHIV.

3. Clinical Presentation of KS in Virally Suppressed HIV-Patients

Data on disease severity are scarce, and probably biased by reporting of most severe cases. However, a 2006 report of nine PLHIV with controlled viremia (<300 copies/mL) and a sustained CD4 count \geq 300 cells/mm³ reported indolent KS cases, as no eruptive cutaneous lesions nor visceral involvement or other AIDS defining illnesses were described [5]. In contrast, a retrospective study from the French CancerVIH group of 21 PLHIV and KS with a median viral suppression of 3 years (IQR 2-5), and a median CD4 level count of 449/mm³ reported frequent severe disease [7]. Eight PLHIV experienced a first episode of KS, all had skin lesion, six (27%) had lymph node involvement and eleven had visceral invasion (bronchial, bone and/or gastric lesions). These cases, referred to an expert national panel for cancer treatment advise for PLHIV, probably represented the more severe cases of KS in aviremic HIV-patients in France at that time. A monocentric study from France described all consecutive diagnosed cases of KS in aviremic (12 cases) and viremic (97 cases) HIV-patients diagnosed in a tertiary referral hospital between 2000 and 2017, comparing their clinical presentations with classic KS (also named "Mediterranean KS", in HIV-negative individuals) consecutively diagnosed in the same area and extracted from the Francim cancer databank [8]. Locally skin indolent presentation was the main clinical presentation in 10 of the KS in aviremic patients, although one patient had visceral involvement and one a disseminated mucocutaneous form. KS in aviremic patients had similar semiology than the 62 classic KS, and, as expected, significantly more indolent presentations than KS in viremic patients. Thus, taking together, and as summarized in

Table 2, most cases of aviremic may be more commonly indolent, though aggressive forms are not exceptional.

| Reference | Number of Subjects: Total (Male/Female) | Median Age of Subjects: Years | Skin or Palatine Involvement | Lymph Node Involvement | Visceral Involvement Oedema or Ulceration or Nodular Oral Lesions |
|-----------|--|----------------------------------|-------------------------------------|---------------------------|---|
| [5] | 9 (-/-) | 51 (range: 41–74) | 9 (100%) | 0 | 0 |
| [6] | 20 (19/1) | 42 (range: 25–59) | 12 (6 | 60%) | 8 (40%) |
| [7] | 21 (17/4) | 54 (interquartile: 35–61) | Skin, 21 (100%) Palatine, 1 (5%) | 6 (27%) | Bronchi, 4 (18%) Bone, 4 (18%) Stomach/esophagus, 3 (14%) |
| [8] | 12 (12/0) | 54 (range: 38–60) | Skin 10 (83.3%) | 1 (8.3%) | Lung, 1 (8.3%) |

Table 2. Clinical presentation of Kaposi's sarcoma in HIV-patients with suppressed viremia in main published series.

There is no classification for KS risk evaluation specific to HIV-patients with suppressed viremia. Thus, classifications used for KS in viremic patients may not be adequate in aviremic patients, and studies should determine whether these classifications predict survival, and/or necessity of systemic therapy in this population. Classifications of AIDS-related KS severity emerged with the AIDS epidemic. The Krigel score-from an initial clinical description of 49 men who have sex with men with KS-proposed four severity stages: locally indolent KS cutaneous lesions (stage I); locally invasive and aggressive form (stage II); disseminated mucocutaneous form (stage III), often with lymph node involvement; and disseminated, mucocutaneous form with visceral involvement (stage IV), further subtyped according to systemic signs of unexplained fever and/or weight loss [13]. There is no validation of this score, and clinicians and researchers felt that the four-stage classification systematically assigned most AIDS-associated cases to stage III or IV, and poorly predicted clinical outcomes [14]. Later, the classification developed by the AIDS Clinical Trial Group (ACTG) of the National Institute of Allergy and Infectious Diseases scored tumor localization and semiology (T), immune deficiency (I), and systemic illness (S) (i.e., fever) to classify subjects in poor or good risk groups [14]. Its predictive value was confirmed in a pre- cART cohort of 294 consecutive patients enrolled in eight ACTG therapeutic trials, with overall survival significantly shorter for patients in the poor-risk categories [15]. This classification remained in use after the advent of cART era. A study from the Swiss HIV cohort showed that staging T1 and a CD4 level <200 cells/mm³ were correlated with death (hazard ratio: 5.22 and 2.33, respectively) in 144 patients KS patients recruited between January 1996 and December 2004 [16]. The classification was also shown to be predictive of Immune Reconstitution Inflammatory Syndrome (IRIS)-associated KS (IRIS-KS) [17]. However, its prognosis value in PLHIV with controlled viremia and KS is unknown.

As KS in aviremic patients may be more commonly indolent [5,8], the use of alternative classifications from non-HIV populations is appealing. A proposed staging system derived from 300 CKS patients based on disease progression was developed [18]: stage I defined by small macules and nodules primarily confined to the lower extremities, stage II by infiltrative plaques mainly involving the lower extremities, sometimes associated with a few nodules, stage III with florid multiple angiomatous plaques and nodules involving the lower extremities that are often ulcerated, and a stage IV with disseminated disease. However, this classification is prone to subjective clinical assessment, and its prognosis value has not been independently validated in CKS studies. An approach based on localized disease (limited cutaneous disease) and advanced disease (advanced cutaneous, oral, visceral or nodal disease), as advocated by the National Comprehensive Cancer Network Guidelines (https://www.nccn.org/professionals/physician_gls/pdf/kaposi. pdf, accessed on 5 November 2021) are probably the most pragmatic guidelines to guide indications of therapeutic interventions, and could probably be also applied for KS in HIV-patients with controlled viremia despite the lack of clinical trials. According to these guidelines, limited cutaneous disease if symptomatic or cosmetically unacceptable is to be

treated with topicals (alitretinoin 0.1% gel, imiquimod 5% cream), and for small lesions (i.e., ≤ 1 cm) intralesional chemotherapy (i.e., vinblastine), radiation therapy, local excision, or cryotherapy. For progressive disease, in case advanced cutaneous, oral, visceral, or nodal disease, first line systemic therapy with liposomal doxorobucin or paclitaxel are preferred options (please refer to guidelines for further details on indications and doses, and below for further discussion on therapeutic options).

4. Physiopathological Hypothesizes

We postulate that the pathological presentation of KS in HIV-patients optimally treated by cART constitutes a specific pattern, due to HHV-8 chronic antigen exposure, immune modulation by viral proteins, and local immune exhaustion. Several immunopathological hypothesis could explain the persistence of HHV-8 related KS during optimally controlled HIV infection with correct immune restoration, implicating persistent immune activation linked to HIV.

Alterations of the innate immunity and inflammation are strongly associated with KS [19], but the activation/inflammation status has not been described in KS patients with controlled plasmatic HIV-RNA so far. However, a decrease in CD4/CD8 ratio has been observed in these patients [20], suggesting that immune activation persists in periphery in this context. Indeed, a low CD4/CD8 ratio has been linked to T cell activation [21]. In vitro, various cytokines such as IL-1, IL-6, TNF-alpha and also Interferon-gamma induce proliferation of HHV-8 infected spindle cells [22]. Concordantly, basic fibroblast growth factor (bFGF) and HIV-1 Tat protein synergize in inducing angiogenic KS-like lesions in mice. Besides, the bFGF, extracellular Tat and Tat receptors are present in HIV-associated KS [23].

Other data suggest a loss of immune control by NK cells and T cells during HIVrelated KS. Indeed, NK-cells have been reported to play a role in the anti-HHV-8 immune response, notably because HHV-8 down-regulates the MHC class I molecules expression on the HHV-8-infected tumor cells. Moreover, the strong expression of NKG2D ligands by tumor cells suggests a defect in NK-cell homing or survival in the KS microenvironment. Furthermore, NK-cells show functional exhaustion in KS patients with lack of response following direct triggering of NKp30, NKp46 or CD16 activating receptors [24]. Another study confirmed this hypo functional profile of NK cells in HIV-infected patients [19]. NK cells were not present in the tumor, suggesting a defect in NK cell cytotoxicity to tumor cells in the context of KS.

Specific T cell responses to HHV-8 were also studied during HIV-related KS. We and others have shown that peripheral blood HHV-8-specific CD8 T-cell responses were much lower in HIV-infected patients with KS than in asymptomatic HHV-8-positive patients, regardless plasma HIV-RNA level [25,26]. Several hypotheses can be made to explain this defect. First HHV-8 encodes two gene products, K3 and K5 (also termed MIR1 and MIR2, respectively), which act in concert to efficiently downregulate the expression of MHC class I molecule on the surface of infected cells, thus preventing antiviral CTL responses [27]. Second, because KS lesions express PD-L1 [28], it is possible that T cells against HHV-8 undergo immune exhaustion. To date, data are not robust enough to conclude about the efficacy of anti-immune checkpoints for treating HIV-related KS. Finally, another hypothesis to the lack of T cell responses in peripheral blood would be the migration of these cells to KS tissues: this hypothesis has not been confirmed so far [25]. Taken together, data suggest that immunodominant HHV-8 proteins are probably not appropriately presented to the immune system during KS, and that T cell exhaustion could also be at play.

5. Therapeutic Challenges for Treating KS in Virally Suppressed HIV-Patients

KS in virally suppressed PLHIV is challenging, as control of HIV replication after introduction of cART and immune improvement are no longer part of the strategy [29].

Since the mid-1990s, highly effective cART has been available, which allow optimal suppression of viral replication. Still currently, cART is based in the vast majority of cases

on a combination of two NRTIs and a third agent, to be chosen from among PIs, NNRTIs or INSTIs. From 1996, PIs and NNRTIs were available, but INSTIs were not marketed until the end of the 2000s. The four series of virally suppressed PLHIV with KS we report in this article (Table 1) include patients treated in the last 10–15 years, and during this period, antiretroviral drugs have changed. In particular, INSTIs now occupy a central position. There is no data in the literature supporting the impact of this change on the incidence of KS. However, the protective role of PIs has been the subject of several works, reported below in this review.

In patients already on cART, KS—once the options of surveillance and local treatment are lapsed—will inevitably require systemic chemotherapy, such as anthracyclines or taxanes (Table 3) [30]. The difference in the proportion of patients having received chemotherapy in the different series reported here could be explained by more or less severe clinical presentations. If recurrences occur despite initial systemic chemotherapy, reiterative cure with inherent risk of cumulative toxicities commonly leads to KS treatment dead ends. As an example, the maximum cumulative dose of liposomal anthracycline is limited to ~550 mg/m² to preserve cardiac functionality, under regular cardiac monitoring [31]. For paclitaxel, it is not uncommon for peripheral neuropathies to prevent continued treatment after a high number of perfusions [32].

| Reference | Mani, J Int Assoc Physicians AIDS Care, 2009 [6] (n = 20) | Palich, <i>Clin Infect Dis</i> , 2019 [7] (<i>n</i> = 21) | Severin, <i>AIDS</i> , 2021 [8] (<i>n</i> = 12) |
|---|--|--|---|
| All cytotoxic treatments | 14 (70%) | 21 (100%) | 2 (17%) |
| Liposomal doxorubicine | 13 (65%) | 19 (90%) | - |
| Paclitaxel | 3 (15%) | 10 (48%) | - |
| Bleomycin | - | 2 (10%) | - |
| Vincristine | - | 1 (5%) | - |
| Antivascular endothelial growth factors | - | 5 (24%) | - |
| Interferon-alfa | - | 2 (10%) | - |
| All local treatments | 4 (20%) | 1 (5%) | 4 (33%) |
| Radiotherapy | 4 (20%) | 1 (5%) | 1 (8%) |
| Cryotherapy | - | - | 1 (8%) |
| Retinoid | - | - | 1 (8%) |
| Laser | - | - | 1 (8%) |
| Unspecified other treatment | 5 (25%) | - | - |
| Clinical surveillance only | 1 (5%) | - | 5 (42%) |
| Clinical outcomes | | | |
| Follow-up duration, months, median | 39 (range: 6–120) | 17 (interquartile: 9–20) | - |
| Complete or partial regression | 13/20 (65%) | 6/16 (37%) | - |
| Stable disease | 4/20 (20%) | 6/16 (38%) | - |
| Progression | 3/20 (15%) | 4/16 (25%) | - |

Table 3. Treatments used for the management of Kaposi's sarcoma in HIV-patients with suppressed viremia, in addition to combined antiretroviral therapy, and clinical outcomes, in the main published series.

Alternative therapies exist, but none are fully satisfactory or validated, and further clinical trials are warranted. Moreover, all strategies have been studies in KS in PLHIV with uncontrolled viral replication. Bleomycin monotherapy can be used, as a suboptimal option, this chemotherapy being less effective than anthracyclines and taxanes for AIDS-related KD, with a risk of pulmonary toxicity [33]. Beside cytotoxic drugs, other treatments have been

evaluated in small studies, such as antivascular endothelial growth factors (lenalidomide and pomalidomide [34,35]) and peginterferon alfa-2a [36], but these are limited by toxicities, limited efficacy. Moreover, most of these chemotherapies present a high potential risk of a drug–drug interaction with cART. In a limited number of patients, topical treatments may also be offered, such as radiotherapy, cryotherapy or topical retinoids, but often systemic treatment is required in case of recurrence despite these treatments.

The management of these patients is based on the general guidelines for AIDSassociated KS, while the therapeutic needs and tolerance are undoubtedly different. The cases reported in the literature support more indolent forms in virally suppressed patients, with, probably, visceral involvement which are rarely life-threatening. Chemotherapies with lower doses could be discussed, as well as a spacing of cures, for better tolerability over time. Induction treatment with chemotherapy, followed by maintenance or suspensive treatment with oral drugs to avoid recurrences could be evaluated, However, drug candidates for maintenance treatment remain to be defined.

Immunotherapy could be an interesting therapeutic option in patients with AIDSrelated KS despite suppressed viremia on cART. Immunotherapy relies on monoclonal antibodies blocking immune checkpoints, such as ipilimumab, blocking CTLA-4, nivolumab and pembrolizumab, blocking PD-1, or atezolizumab, blocking PD-L1. The lack of specific anti-HHV-8 cellular immunity has been shown to participate in the occurrence of KS [26]. Blocking immune checkpoints could restore anti-HHV-8 immunity and help control the tumor process. The expression of PD-1 and PD-L1 within tumor tissue has been shown to be a marker strongly correlated with the effectiveness of immunotherapy [37]. Some histological works show the expression of these immune checkpoints from KS biopsies [28,38–42], supporting possible therapeutic effects of immune checkpoints inhibitors on KS. Only one observational study reported AIDS-related KS treated by anti-PD-1 therapy [43]. Among the 9 reported cases, 6 were on cART for at least 12 months, with HIV-RNA <50 copies/mL. All had received previous treatments, included liposomal anthracycline, paclitaxel, lenalidomide and bortezomib. All had cutaneous involvement, with lymph node (n = 3), gastro-intestinal (n = 2) and lung (n = 1) extension. The authors concluded that there was partial remission in 4 patients, and stable disease in 2 patients, with a median follow-up of 5 months. Regarding adverse events, several cohorts have shown similar tolerance of immunotherapy in PLHIV than in people without HIV [44–47]. In particular, no immunological and virological effects were detected in these patients. Immune-related adverse events are commonly mild to moderate, included skin, musculoskeletal, gastrointestinal and endocrine impairment. However, some rare and serious side effects can occur with a real risk of death, including immune-induced pneumonia or cardiomyopathy. Overall, although there is no marketing approval for anti-immune checkpoint antibodies for KS treatment today, this therapeutic way could be an interesting option for PLHIV with suppressed viremia. Nevertheless, indications for immunotherapy should be balanced against the risk of serious adverse reactions, especially when KS does not endanger the patient's life.

6. Role of Protease Inhibitors for Treatment

HIV PIs were shown in vitro to have through their action on cellular proteases a wide range of effects on pathways that are important for tumorigenesis, including reducing angiogenesis and cell invasion, inhibition of the Akt pathway, induction of autophagy, and promotion of apoptosis [48]. These properties, in addition but independently of the effect of PIs on inhibition of the HIV protease and HIV replication, were suggested in vitro and in experimental models to be potentially beneficial on KS prevention and management.

The first-generation PIs ritonavir and saquinavir inhibit in vitro activation and proliferation of primary endothelial cells and KS cell lines through induction of apoptosis of tumor cells by modulating proteasomal proteolysis without affecting proliferation or survival of noncancerous cell. They also decrease production of vascular endothelial growth factor (VEGF) and inflammatory cytokines (tumor necrosis factor- α , interleukin-6 and -8), which are critical to KS development and proliferation [49,50]. Ritonavir inhibited tumor formation and progression by KS-derived cells in a KS mouse xenotransplantation model [50]. Systemic administration of indinavir and saquinavir to nude mice blocked the development and induced regression of angioproliferative KS-like lesions established by primary human KS cells by the inhibition of matrix metalloproteinase-2 proteolytic activation at concentrations present in plasma of treated individuals [51]. Moreover, nelfinavir, but no other PIs, was shown to be a potent inhibitor of HHV-8 replication in vitro through interference with the production of infectious virus [52]. In a small uncontrolled study that aimed to assess the effectiveness of indinavir as a therapy for classical KS in HIVseronegative patients, favorable clinical outcome was achieved in 16/26 patients (61.5%), and was associated with higher plasmatic indinavir concentrations, reduced plasmatic levels of basic fibroblast growth factor, lower numbers of circulating endothelial cells, and decreased antibody titers against HHV-8, compared to patients with unfavorable course, although HHV-8 viral load was not monitored [53]. These data were followed by case series and case reports describing onset or relapses of KS despite long-term remission under PI-based regimen after being switched to a non-nucleoside reverse transcriptase inhibitor (NNRTI)- or integrase strand transfer inhibitor (INSTI)-based ART [54-56]. A retrospective study performed on the Veterans Affairs HIV Clinical Case Registry showed that longer duration on ritonavir boosted PI-based regimen significantly reduced KS incidence among male Veterans, in comparison with other regimens, after accounting for potential confounders including HIV viral load, CD4 T cell count and IRIS effect, with a most pronounced protective effect after at least one year on ART. This effect was not demonstrated for nelfinavir, non-boosted PIs and NNRTI-based regimen [57].

However, the potential effect of PIs on KS has not obviously translated into the clinic. Onset of KS was reported in patients with controlled HIV replication while on PI-based ART regimen [5,58]. Several cohorts (EuroSIDA, COHERE in EuroCoord, French Hospital Database on HIV, Chelsea and Westminster HIV cohort) assessing incidence and risk factors for KS stated that boosted PI-based regimens were not associated with a lower risk of developing KS than NNRTI-based regimens in the overall population or in men who have sex with men [1,10,59–61]. Of note these studies usually did not adjust for possible KS-IRIS effect after ART initiation representing a potential confounding factor in the interpretation of their results [62]. In cohorts of KS patients that aimed to identify predictive factors of KS remission, response rate was not associated with NNRTI-based, PI-based, or boosted-PI based regimen [63,64]. Furthermore, an analysis of the prospectively collected Dat'AIDS database focusing on PLHIV with history of KS and controlled HIV replication, switching from a PI-based to a PI-free regimen was not associated with an increased risk of KS relapse [65].

Therefore, while being a relevant issue in the setting of current recommendations of ART optimization with PI-free regimens [66], and despite a tempting pre-clinical rationale, no clinical data supports a beneficial direct effect of PIs on HHV-8 and KS. Both PI- and NNRTI-based ART were shown to be equally effective in protecting against KS. This suggest that reduction in KS incidence on ART is related to improved immune recovery and HIV control, and that KS relapses in the setting of controlled HIV replication could be mainly mediated by permanent loss of the anti-HHV-8 T cell immune responses despite CD4 T cell quantitative restoration under ART [26], rather than by a specific anti-HHV-8 or antiangiogenic effect of PI-containing regimen.

7. Conclusions

KS in virally suppressed HIV-patients represents a clinical and biological entity that is still poorly understood. Exploration of immunopathological pathways is important to develop more effective therapeutic strategies, by avoiding reiterative cytotoxic chemotherapy. The search for genetic abnormalities by high-throughput sequencing of tumor tissue could also reveal previously unidentified therapeutic targets. We believe that it is necessary to constitute a prospective cohort including HIV-patients with KS despite suppressed HIV viremia on cART, in order to collect exhaustive data on these cases, and to describe the evolution of the disease according to the treatments received, with their potential associated adverse effects, and factors associated with negative outcomes. Such a cohort is now ongoing on all patients presented at the expert panel. We plead also for the implementation of clinical trials specifically enrolling virally suppressed PLHIV, in order to evaluate innovative approaches, such as induction/maintenance schemes, based on drug-reduces cytotoxic chemotherapy, immunotherapy and oral new targeted therapies.

Author Contributions: R.P., A.M., M.V., A.G., M.-A.V., S.B.-R., I.P.-M., C.S., S.G., G.M.-B. and J.-P.S. have participated in the preparation and the writing of the original draft, as well as the writing, the review and the editing of the final version. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: All authors have no conflict of interest to declare.

References

- 1. Grabar, S.; Abraham, B.; Mahamat, A.; del Giudice, P.; Rosenthal, E.; Costagliola, D. Differential impact of combination antiretroviral therapy in preventing Kaposi's sarcoma with and without visceral involvement. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* 2006, 24, 3408–3414. [CrossRef]
- 2. Hernández-Ramírez, R.U.; Shiels, M.S.; Dubrow, R.; Engels, E.A. Cancer risk in HIV-infected people in the USA from 1996 to 2012: A population-based, registry-linkage study. *Lancet HIV* **2017**, *4*, e495–e504. [CrossRef]
- Hleyhel, M.; Belot, A.; Bouvier, A.M.; Tattevin, P.; Pacanowski, J.; Genet, P.; de Castro, N.; Berger, J.-L.; Dupont, C.; Lavolé, A.; et al. Risk of AIDS-defining cancers among HIV-1-infected patients in France between 1992 and 2009: Results from the FHDH-ANRS CO4 cohort. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2013, *57*, 1638–1647. [CrossRef] [PubMed]
- Luo, Q.; Satcher Johnson, A.; Hall, H.I.; Cahoon, E.K.; Shiels, M. Kaposi sarcoma rates among persons living with human immunodeficiency virus in the United States: 2008–2016. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2020, 73, e2226–e2233. [CrossRef] [PubMed]
- 5. Maurer, T.; Ponte, M.; Leslie, K. HIV-Associated Kaposi's sarcoma with a High CD4 count and a low viral load. *N. Engl. J. Med.* **2007**, *357*, 1352–1353. [CrossRef] [PubMed]
- Mani, D.; Neil, N.; Israel, R.; Aboulafia, D.M. A retrospective analysis of AIDS-associated Kaposi's sarcoma in patients with undetectable HIV viral loads and CD4 counts greater than 300 Cells/Mm(3). J. Int. Assoc. Physicians AIDS Care 2009, 8, 279–285. [CrossRef]
- Palich, R.; Veyri, M.; Valantin, M.-A.; Marcelin, A.-G.; Guihot, A.; Pourcher, V.; Jary, A.; Solas, C.; Makinson, A.; Poizot-Martin, I.; et al. Recurrence and occurrence of Kaposi's sarcoma in patients living with human immunodeficiency virus (HIV) and on antiretroviral therapy, despite suppressed HIV viremia. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2020, 70, 2435–2438. [CrossRef]
- 8. Séverin, D.; Bessaoud, F.; Meftah, N.; Du Thanh, A.; Tretarre, B.; Guillot, B.; Makinson, A. A comparative study of classic and HIV-viremic and aviremic AIDS Kaposi sarcoma. *AIDS* **2021**, *35*, 399–405. [CrossRef]
- Park, L.S.; Tate, J.P.; Sigel, K.; Brown, S.T.; Crothers, K.; Gibert, C.; Goetz, M.B.; Rimland, D.; Rodriguez-Barradas, M.C.; Bedimo, R.J.; et al. Association of viral suppression with lower AIDS-defining and non-AIDS-defining cancer incidence in HIV-infected veterans: A prospective cohort study. *Ann. Intern. Med.* 2018, 169, 87–96. [CrossRef]
- Cancer Project Working Group for the Collaboration of Observational HIV Epidemiological Research Europe (COHERE). Study in EuroCoord Changing Incidence and Risk Factors for Kaposi Sarcoma by Time Since Starting Antiretroviral Therapy: Collaborative Analysis of 21 European Cohort Studies. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 2016, 63, 1373–1379. [CrossRef]
- Crum-Cianflone, N.F.; Hullsiek, K.H.; Ganesan, A.; Weintrob, A.; Okulicz, J.F.; Agan, B.K.; Infectious Disease Clinical Research Program HIV Working Group. Is Kaposi's sarcoma occurring at higher CD4 cell counts over the course of the HIV epidemic? *AIDS* 2010, 24, 2881–2883. [CrossRef] [PubMed]
- Yanik, E.L.; Achenbach, C.J.; Gopal, S.; Coghill, A.E.; Cole, S.R.; Eron, J.J.; Moore, R.D.; Mathews, W.C.; Drozd, D.R.; Hamdan, A.; et al. Changes in clinical context for Kaposi's sarcoma and non-Hodgkin lymphoma among people with HIV infection in the United States. J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol. 2016, 34, 3276–3283. [CrossRef]
- 13. Krigel, R.L.; Laubenstein, L.J.; Muggia, F.M. Kaposi's sarcoma: A new staging classification. Cancer Treat. Rep. 1983, 67, 531–534.
- 14. Krown, S.E.; Metroka, C.; Wernz, J.C. Kaposi's sarcoma in the acquired immune deficiency syndrome: A proposal for uniform evaluation, response, and staging criteria. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* **1989**, *7*, 1201–1207. [CrossRef] [PubMed]
- 15. Krown, S.E.; Testa, M.A.; Huang, J. AIDS-related Kaposi's sarcoma: Prospective validation of the AIDS clinical trials group staging classification. J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol. 1997, 15, 3085–3092. [CrossRef]
- El Amari, E.B.; Toutous-Trellu, L.; Gayet-Ageron, A.; Baumann, M.; Cathomas, G.; Steffen, I.; Erb, P.; Mueller, N.J.; Furrer, H.; Cavassini, M.; et al. Predicting the evolution of Kaposi sarcoma, in the highly active antiretroviral therapy era. *AIDS* 2008, 22, 1019–1028. [CrossRef] [PubMed]

- 17. Letang, E.; Lewis, J.J.; Bower, M.; Mosam, A.; Borok, M.; Campbell, T.B.; Naniche, D.; Newsom-Davis, T.; Shaik, F.; Fiorillo, S.; et al. Immune reconstitution inflammatory syndrome associated with kaposi sarcoma: Higher incidence and mortality in Africa than in the UK. *AIDS* **2013**, *27*, 1603–1613. [CrossRef]
- 18. Brambilla, L.; Boneschi, V.; Taglioni, M.; Ferrucci, S. Staging of classic Kaposi's sarcoma: A useful tool for therapeutic choices. *Eur. J. Dermatol.* **2003**, *13*, 83–86.
- 19. Ensoli, B.; Sgadari, C.; Barillari, G.; Sirianni, M.C.; Stürzl, M.; Monini, P. Biology of Kaposi's Sarcoma. *Eur. J. Cancer* 2001, 37, 1251–1269. [CrossRef]
- Caby, F.; Guiguet, M.; Weiss, L.; Winston, A.; Miro, J.M.; Konopnicki, D.; le Moing, V.; Bonnet, F.; Reiss, P.; Mussini, C.; et al. CD4/CD8 ratio and the risk of Kaposi sarcoma or non-Hodgkin lymphoma in the context of efficiently treated human immunodeficiency virus (HIV) infection: A collaborative analysis of 20 European cohort studies. *Clin. Infect. Dis.* 2020, *73*, 50–59. [CrossRef]
- Serrano-Villar, S.; Sainz, T.; Lee, S.A.; Hunt, P.W.; Sinclair, E.; Shacklett, B.L.; Ferre, A.L.; Hayes, T.L.; Somsouk, M.; Hsue, P.Y.; et al. HIV-infected individuals with low CD4/CD8 ratio despite effective antiretroviral therapy exhibit altered T cell subsets, heightened CD8+ T cell activation, and increased risk of non-AIDS morbidity and mortality. *PLoS Pathog.* 2014, 10, e1004078. [CrossRef] [PubMed]
- Barillari, G.; Buonaguro, L.; Fiorelli, V.; Hoffman, J.; Michaels, F.; Gallo, R.C.; Ensoli, B. Effects of cytokines from activated immune cells on vascular cell growth and HIV-1 gene expression. Implications for AIDS-Kaposi's sarcoma pathogenesis. *J. Immunol.* 1992, 149, 3727. [PubMed]
- Ensoli, B.; Gendelman, R.; Markham, P.; Fiorelli, V.; Colombini, S.; Raffeld, M.; Cafaro, A.; Chang, H.-K.; Brady, J.N.; Gallo, R.C. Synergy between basic fibroblast growth factor and HIV-1 tat protein in induction of Kaposi's sarcoma. *Nature* 1994, 371, 674–680. [CrossRef] [PubMed]
- Beldi-Ferchiou, A.; Lambert, M.; Dogniaux, S.; Vély, F.; Vivier, E.; Olive, D.; Dupuy, S.; Levasseur, F.; Zucman, D.; Lebbé, C.; et al. PD-1 mediates functional exhaustion of activated NK cells in patients with Kaposi sarcoma. *Oncotarget* 2016, 7, 72961–72977. [CrossRef] [PubMed]
- Lambert, M.; Gannagé, M.; Karras, A.; Abel, M.; Legendre, C.; Kerob, D.; Agbalika, F.; Girard, P.-M.; Lebbe, C.; Caillat-Zucman, S. Differences in the frequency and function of HHV8-specific CD8 T cells between asymptomatic HHV8 infection and Kaposi sarcoma. *Blood* 2006, *108*, 3871–3880. [CrossRef]
- Guihot, A.; Dupin, N.; Marcelin, A.-G.; Gorin, I.; Bedin, A.-S.; Bossi, P.; Galicier, L.; Oksenhendler, E.; Autran, B.; Carcelain, G. Low T cell responses to human herpesvirus 8 in patients with AIDS-related and classic Kaposi sarcoma. *J. Infect. Dis.* 2006, 194, 1078–1088. [CrossRef]
- Coscoy, L.; Ganem, D. Kaposi's sarcoma-associated herpesvirus encodes two proteins that block cell surface display of MHC class I chains by enhancing their endocytosis. *Proc. Natl. Acad. Sci. USA* 2000, *97*, 8051–8056. [CrossRef]
- Mletzko, S.; Pinato, D.J.; Robey, R.C.; Dalla Pria, A.; Benson, P.; Imami, N.; Bower, M. Programmed death ligand 1 (PD-L1) expression influences the immune-tolerogenic microenvironment in antiretroviral therapy-refractory Kaposi's sarcoma: A pilot study. *Oncoimmunology* 2017, 6, e1304337. [CrossRef]
- 29. Tam, H.K.; Zhang, Z.-F.; Jacobson, L.P.; Margolick, J.B.; Chmiel, J.S.; Rinaldo, C.; Detels, R. Effect of highly active antiretroviral therapy on survival among HIV-infected men with Kaposi sarcoma or non-Hodgkin lymphoma. *Int. J. Cancer* **2002**, *98*, 916–922. [CrossRef]
- 30. Gbabe, O.F.; Okwundu, C.I.; Dedicoat, M.; Freeman, E.E. Treatment of severe or progressive Kaposi's sarcoma in HIV-infected adults. *Cochrane Database Syst. Rev.* 2014, 9, CD003256. [CrossRef]
- 31. Safra, T. Cardiac safety of liposomal anthracyclines. *Oncologist* 2003, *8*, 17–24. [CrossRef]
- 32. Marupudi, N.I.; Han, J.E.; Li, K.W.; Renard, V.M.; Tyler, B.M.; Brem, H. Paclitaxel: A review of adverse toxicities and novel delivery strategies. *Expert Opin. Drug Saf.* 2007, *6*, 609–621. [CrossRef] [PubMed]
- 33. Stewart, S.; Jablonowski, H.; Goebel, F.D.; Arasteh, K.; Spittle, M.; Rios, A.; Aboulafia, D.; Galleshaw, J.; Dezube, B.J. Randomized comparative trial of pegylated liposomal doxorubicin versus bleomycin and vincristine in the treatment of AIDS-related Kaposi's sarcoma. International pegylated liposomal doxorubicin study group. J. Clin. Oncol. 1998, 16, 683–691. [CrossRef] [PubMed]
- 34. Martinez, V.; Tateo, M.; Castilla, M.-A.; Melica, G.; Kirstetter, M.; Boué, F. Lenalidomide in treating AIDS-related Kaposi's sarcoma. *AIDS* 2011, 25, 878–880. [CrossRef] [PubMed]
- Polizzotto, M.N.; Uldrick, T.S.; Wyvill, K.M.; Aleman, K.; Peer, C.J.; Bevans, M.; Sereti, I.; Maldarelli, F.; Whitby, D.; Marshall, V.; et al. Pomalidomide for symptomatic Kaposi's sarcoma in people with and without HIV infection: A phase I/II study. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* 2016, 34, 4125–4131. [CrossRef]
- 36. Rokx, C.; van der Ende, M.E.; Verbon, A.; Rijnders, B.J.A. Peginterferon Alfa-2a for AIDS-associated Kaposi sarcoma: Experience with 10 patients. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* **2013**, 57, 1497–1499. [CrossRef]
- 37. Gibney, G.T.; Weiner, L.M.; Atkins, M.B. Predictive biomarkers for checkpoint inhibitor-based immunotherapy. *Lancet Oncol.* **2016**, *17*, e542–e551. [CrossRef]
- 38. Chen, J.; del Valle, L.; Lin, H.-Y.; Plaisance-Bonstaff, K.; Forrest, J.C.; Post, S.R.; Qin, Z. Expression of PD-1 and PD-Ls in Kaposi's sarcoma and regulation by oncogenic herpesvirus lytic reactivation. *Virology* **2019**, *536*, 16–19. [CrossRef]
- Paydas, S.; Bagir, E.K.; Deveci, M.A.; Gonlusen, G. Clinical and prognostic significance of PD-1 and PD-L1 expression in sarcomas. *Med. Oncol.* 2016, 33, 93. [CrossRef]

- 40. Genovese, G.; Venegoni, L.; Fanoni, D.; Tourlaki, A.; Brambilla, L.; Berti, E. PD-L1 expression in tumour microenvironment supports the rationale for immune checkpoint blockade in classic Kaposi's sarcoma. *J. Eur. Acad. Dermatol. Venereol.* **2019**, *33*, e269–e271. [CrossRef]
- Joest, B.; Kempf, W.; Berisha, A.; Peyk, P.; Tronnier, M.; Mitteldorf, C. Stage-related PD-L1 expression in Kaposi sarcoma tumor microenvironment. J. Cutan. Pathol. 2020, 47, 888–895. [CrossRef]
- 42. Kim, Y.J.; Jung, C.J.; Won, C.H.; Chang, S.E.; Lee, M.W.; Choi, J.H.; Lee, W.J. PD-1 and PD-L1 expression in Kaposi sarcoma: A comparative study according to the pathological stage and clinical characteristics. *J. Cutan. Pathol.* 2021, 48, 221–228. [CrossRef]
- 43. Galanina, N.; Goodman, A.M.; Cohen, P.R.; Frampton, G.M.; Kurzrock, R. Successful treatment of HIV-associated Kaposi sarcoma with immune checkpoint blockade. *Cancer Immunol. Res.* **2018**, *6*, 1129–1135. [CrossRef] [PubMed]
- 44. Puronen, C.E.; Ford, E.S.; Uldrick, T.S. Immunotherapy in people with HIV and cancer. *Front. Immunol.* **2019**, *10*, 2060. [CrossRef] [PubMed]
- Cook, M.R.; Kim, C. Safety and efficacy of immune checkpoint inhibitor therapy in patients with HIV infection and advanced-stage cancer: A systematic review. JAMA Oncol. 2019, 5, 1049–1054. [CrossRef] [PubMed]
- Uldrick, T.S.; Gonçalves, P.H.; Abdul-Hay, M.; Claeys, A.J.; Emu, B.; Ernstoff, M.S.; Fling, S.P.; Fong, L.; Kaiser, J.C.; Lacroix, A.M.; et al. Assessment of the safety of pembrolizumab in patients with HIV and Advanced cancer-A phase 1 study. *JAMA Oncol.* 2019, 5, 1332–1339. [CrossRef] [PubMed]
- Spano, J.-P.; Veyri, M.; Gobert, A.; Guihot, A.; Perré, P.; Kerjouan, M.; Brosseau, S.; Cloarec, N.; Montaudié, H.; Helissey, C.; et al. Immunotherapy for cancer in people living with HIV: Safety with an efficacy signal from the series in real life experience. *AIDS* 2019, 33, F13–F19. [CrossRef]
- 48. Gantt, S.; Casper, C.; Ambinder, R.F. Insights into the broad cellular effects of nelfinavir and the HIV protease inhibitors supporting their role in cancer treatment and prevention. *Curr. Opin. Oncol.* **2013**, *25*, 495–502. [CrossRef]
- Gaedicke, S.; Firat-Geier, E.; Constantiniu, O.; Lucchiari-Hartz, M.; Freudenberg, M.; Galanos, C.; Niedermann, G. Antitumor effect of the human immunodeficiency virus protease inhibitor ritonavir: Induction of tumor-cell apoptosis associated with perturbation of proteasomal proteolysis. *Cancer Res.* 2002, *62*, 6901–6908. [PubMed]
- 50. Pati, S.; Pelser, C.B.; Dufraine, J.; Bryant, J.L.; Reitz, M.S.; Weichold, F.F. Antitumorigenic effects of HIV protease inhibitor ritonavir: Inhibition of Kaposi sarcoma. *Blood* 2002, *99*, 3771–3779. [CrossRef]
- 51. Sgadari, C.; Barillari, G.; Toschi, E.; Carlei, D.; Bacigalupo, I.; Baccarini, S.; Palladino, C.; Leone, P.; Bugarini, R.; Malavasi, L.; et al. HIV protease inhibitors are potent anti-angiogenic molecules and promote regression of Kaposi sarcoma. *Nat. Med.* **2002**, *8*, 225–232. [CrossRef]
- Gantt, S.; Carlsson, J.; Ikoma, M.; Gachelet, E.; Gray, M.; Geballe, A.P.; Corey, L.; Casper, C.; Lagunoff, M.; Vieira, J. The HIV protease inhibitor nelfinavir inhibits Kaposi's sarcoma-associated herpesvirus replication in vitro. *Antimicrob. Agents Chemother.* 2011, 55, 2696–2703. [CrossRef] [PubMed]
- Monini, P.; Sgadari, C.; Grosso, M.G.; Bellino, S.; di Biagio, A.; Toschi, E.; Bacigalupo, I.; Sabbatucci, M.; Cencioni, G.; Salvi, E.; et al. Clinical course of classic Kaposi's sarcoma in HIV-negative patients treated with the HIV protease inhibitor indinavir. *AIDS* 2009, 23, 534–538. [CrossRef]
- Bani-Sadr, F.; Fournier, S.; Molina, J.M. Relapse of Kaposi's sarcoma in HIV-infected patients switching from a protease inhibitor to a non-nucleoside reverse transcriptase inhibitor-based highly active antiretroviral therapy regimen. *AIDS* 2003, *17*, 1580–1581. [CrossRef] [PubMed]
- 55. Simonetti, F.R.; Ricaboni, D.; Cattaneo, D.; Micheli, V.; Rusconi, S.; Gervasoni, C. Relapse of Kaposi's sarcoma and HHV-8 Viremia in an HIV-infected patient switching from protease inhibitor to integrase inhibitor-based antiretroviral therapy. *J. Clin. Virol. Off. Publ. Pan Am. Soc. Clin. Virol.* **2016**, *74*, 75–77. [CrossRef]
- 56. Philibert, P.; Chiche, L.; Caillères, S.; Allemand, J.; Rebaudet, S.; Delord, M.; Stavris, C.; Retornaz, F.; Khiri, H.; Halfon, P. HHV8 and Kaposi's sarcoma: Should We really give up protease inhibitors in All HIV-infected patients? *AIDS* **2017**, *31*, 2167–2169. [CrossRef] [PubMed]
- 57. Kowalkowski, M.A.; Kramer, J.R.; Richardson, P.R.; Suteria, I.; Chiao, E.Y. Use of boosted protease inhibitors reduces Kaposi sarcoma incidence among male veterans with HIV infection. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* **2015**, *60*, 1405–1414. [CrossRef]
- Carleo, M.A.; di Martino, F.; del Giudice, A.; Gargiulo, M.; Parrella, G.; Rosario, P.; Sangiovanni, V.; Viglietti, R.; Esposito, V.; Chirianni, A. Different impact of anti-retroviral regimen containing protease inhibitors on development of HIV-related Kaposi sarcoma. *In Vivo* 2015, *29*, 133–136.
- Portsmouth, S.; Stebbing, J.; Gill, J.; Mandalia, S.; Bower, M.; Nelson, M.; Bower, M.; Gazzard, B. A comparison of regimens based on non-nucleoside reverse transcriptase inhibitors or protease inhibitors in preventing Kaposi's sarcoma. *AIDS* 2003, 17, F17–F22. [CrossRef]
- 60. Stebbing, J.; Portsmouth, S.; Nelson, M.; Mandalia, S.; Kandil, H.; Alexander, N.; Davies, L.; Brock, C.; Bower, M.; Gazzard, B. The efficacy of ritonavir in the prevention of AIDS-related Kaposi's sarcoma. *Int. J. Cancer* **2004**, *108*, 631–633. [CrossRef]
- Mocroft, A.; Kirk, O.; Clumeck, N.; Gargalianos-Kakolyris, P.; Trocha, H.; Chentsova, N.; Antunes, F.; Stellbrink, H.-J.; Phillips, A.N.; Lundgren, J.D. The changing pattern of kaposi sarcoma in patients with HIV, 1994–2003: The EuroSIDA study. *Cancer* 2004, 100, 2644–2654. [CrossRef]

- 62. Bower, M.; Nelson, M.; Young, A.M.; Thirlwell, C.; Newsom-Davis, T.; Mandalia, S.; Dhillon, T.; Holmes, P.; Gazzard, B.G.; Stebbing, J. Immune reconstitution inflammatory syndrome associated with Kaposi's sarcoma. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* 2005, 23, 5224–5228. [CrossRef] [PubMed]
- Martinez, V.; Caumes, E.; Gambotti, L.; Ittah, H.; Morini, J.-P.; Deleuze, J.; Gorin, I.; Katlama, C.; Bricaire, F.; Dupin, N. Remission from Kaposi's sarcoma on HAART is associated with suppression of HIV replication and is independent of protease inhibitor therapy. *Br. J. Cancer* 2006, *94*, 1000–1006. [CrossRef] [PubMed]
- 64. Nguyen, H.Q.; Magaret, A.S.; Kitahata, M.M.; van Rompaey, S.E.; Wald, A.; Casper, C. Persistent Kaposi sarcoma in the era of highly active antiretroviral therapy: Characterizing the predictors of clinical response. *AIDS* **2008**, *22*, 937–945. [CrossRef]
- 65. Lajaunie, R.; Cuzin, L.; Palich, R.; Makinson, A.; Bani-Sadr, F.; Duvivier, C.; Arvieux, C.; Rey, D.; Poizot-Martin, I.; Delpierre, C.; et al. No increased risk of Kaposi sarcoma relapse in patients with controlled HIV-1 infection after switching protease inhibitor-based antiretroviral therapy. *HIV Med.* **2021**. [CrossRef] [PubMed]
- Saag, M.S.; Gandhi, R.T.; Hoy, J.F.; Landovitz, R.J.; Thompson, M.A.; Sax, P.E.; Smith, D.M.; Benson, C.A.; Buchbinder, S.P.; del Rio, C.; et al. Antiretroviral drugs for treatment and prevention of HIV infection in adults: 2020 recommendations of the international antiviral society-USA panel. *JAMA* 2020, *324*, 1651–1669. [CrossRef] [PubMed]