

Diagnostic approach in adult-onset neurometabolic diseases

Gorka Fernández-Eulate, Christophe Carreau, Jean-François Benoist, Foudil Lamari, Benoit Rucheton, Natalia Shor, Yann Nadjar

To cite this version:

Gorka Fernández-Eulate, Christophe Carreau, Jean-François Benoist, Foudil Lamari, Benoit Rucheton, et al.. Diagnostic approach in adult-onset neurometabolic diseases. Journal of Neurology, Neurosurgery and Psychiatry, 2022, jnnp-2021-328045. 10.1136/jnnp-2021-328045. hal-03571929

HAL Id: hal-03571929 <https://hal.sorbonne-universite.fr/hal-03571929v1>

Submitted on 14 Feb 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Review

Diagnostic approach in adult-onset neurometabolic diseases

Gorka Fernández-Eulate (a ,^{1,2} Christophe Carreau (a ,³ Jean-François Benoist,⁴ Foudil Lamari,⁵ Benoit Rucheton,⁵ Natalia Shor,⁶ Yann Nadjar ^{● 1}

ABSTRACT

► Additional supplemental material is published online only. To view, please visit the journal online [\(http://dx.](http://dx.doi.org/10.1136/jnnp-2021-328045) [doi.org/10.1136/jnnp-2021-](http://dx.doi.org/10.1136/jnnp-2021-328045) [328045](http://dx.doi.org/10.1136/jnnp-2021-328045)).

1 Neuro-Metabolism Unit, Reference Center for Lysosomal Diseases, Neurology Department, Pitié-Salpêtrière University Hospital, APHP, Paris, France 2 Reference Center for Neuromuscular Diseases, Neuromyology Department, Pitié-Salpêtrière University Hospital, APHP, Paris, France ³Neurology Department, Saint-Louis University Hospital, APHP, Paris, France 4 Metabolic Biochemistry Laboratory, Necker Enfants Malades University Hospital, APHP, Paris-Saclay University, Paris, France 5 Department of Biochemistry of Neurometabolic Diseases, Pitié-Salpêrière University Hospital, APHP, Paris, Fance ⁶Neuroradiology Department, Pitié-Salpêtrière University Hospital, APHP, Sorbonne University, Paris, France

Correspondence to

Dr Yann Nadjar, Neurology, Assistance Publique, Hopitaux de Paris, Paris, France; yann. nadjar@aphp.fr

Received 18 September 2021 Accepted 2 January 2022

Check for updates

© Author(s) (or their employer(s)) 2022. Re-use permitted under CC BY-NC. No commercial re-use. See rights and permissions. Published by BMJ.

To cite: Fernández-Eulate G, Carreau C, Benoist J-F, et al. J Neurol Neurosurg Psychiatry Epub ahead of print: [please include Day Month Year]. doi:10.1136/jnnp-2021- 328045

unlike most neurogenetic diseases, many neurometabolic diseases are treatable, with both conservative and more recent innovative therapeutics. However, the phenotypical complexity of this group of diseases and the growing number of specialised biochemical tools account for a significant diagnostic delay and underdiagnosis. We reviewed all series and case reports of patients with a confirmed neurometabolic disease and a neurological onset after the age of 10 years, with a focus on the 36 treatable ones, and classified these diseases according to their most relevant clinical manifestations. The biochemical diagnostic approach of neurometabolic diseases lays on the use of numerous tests studying a set of metabolites, an enzymatic activity or the function of a given pathway; and therapeutic options aim to restore the enzyme activity or metabolic function, limit the accumulation of toxic substrates or substitute the deficient products. A quick diagnosis of a treatable neurometabolic disease can have a major impact on patients, leading to the stabilisation of the disease and cease of repeated diagnostic investigations, and allowing for familial screening. For the aforementioned, in addition to an exhaustive and clinically meaningful review of these diseases, we propose a simplified diagnostic approach for the neurologist with the aim to help determine when to suspect a neurometabolic disease and how to proceed in a rational manner. We also discuss the place of next-generation sequencing technologies in the diagnostic process, for which deep phenotyping of patients (both clinical and biochemical) is necessary for improving their diagnostic yield. **INTRODUCTION** Inborn errors of metabolism with neurological

Neurometabolic diseases are a group of individually rare but numerous and heterogeneous genetic diseases best known to paediatricians. The more recently reported adult forms may present with phenotypes strikingly different from paediatric ones and may mimic other more common neurological disorders in adults. Furthermore,

manifestations, or neurometabolic diseases, are a group of heterogeneous genetic disorders that share in common the alteration of specific aspects of the cellular metabolism, ultimately leading to disease. Often first described by paediatricians, the more recently reported adult-onset forms have phenotypes sometimes considerably different from paediatric ones, which may mimic other more common neurological disorders in adults, thus justifying a specific approach.¹⁻³ Adult-onset neurometabolic diseases are individually rare, numerous, heterogeneous and frequently show complex clinical presentations. These reasons, in addition to the myriad of specialised biochemical diagnostic tools available,⁴ account for a significant diagnostic delay and underdiagnosis.⁵ However, unlike many other neurogenetic diseases, a substantial part of neurometabolic diseases can be successfully treated, with both conservative and more recently approved innovative therapeutics. Early recognition and diagnosis of a treatable neurometabolic disease can have a major impact for patients, leading to the stabilisation of the disease or even the regression of some signs and symptoms, halting unnecessary diagnostic investigations, and allowing for family screening and treatment of presymptomatic carriers.

For all of the aforementioned, an overview of adult-onset neurometabolic diseases will be outlined, from important general considerations to phenotypical descriptions focused on treatable diseases. Furthermore, a simplified diagnostic approach for the adult neurologist will be presented, with the aim to help determine when to suspect a neurometabolic disease and how to further proceed in a rational manner.

METHODS

Search strategy and selection criteria

Neurometabolic diseases are a group of genetic disorders that share in common the alteration of the cellular metabolism, ultimately leading to disease including neurological manifestations. This definition has its limitations, given that many neurological diseases may be associated to changes in cellular metabolism that participate in some degree to the pathogenesis of the disease. Within the aim of a pragmatic clinical approach, this review will consider as a neurometabolic disease those that can be either diagnosed on characteristic biochemical abnormalities, indicating an alteration of an specific metabolic pathway, and/or those that may respond to treatments aimed at correcting a given metabolic dysfunction.6 In order to review all adult-onset neurometabolic diseases, we first searched PubMed for article abstracts published in English, French and Spanish using the search terms "inborn errors of metabolism", "metabolic disease" and "neurometabolic disease", and reviewed the book by Hollak and Lachmann in $2016⁷$ to assemble all adult-onset neurometabolic diseases, including patients from any ethnic origin (see [online supplemental material](https://dx.doi.org/10.1136/jnnp-2021-328045) [1\)](https://dx.doi.org/10.1136/jnnp-2021-328045). Furthermore, we consider as 'adult onset' those

Figure 1 Neurological and extraneurological manifestations of treatable adult-onset neurometabolic diseases. Neurometabolic diseases are classified according to their neurological systematisation: brain: encephalic involvement; spinal cord 'plus': associated spinal cord involvement; nerve plus: associated nerve involvement; muscle plus: associated muscle involvement. The boxes have been coloured according to the topography of the manifestation: red for cortical, blue for subcortical, green for spinal cord, yellow for peripheral nerve and pink for muscle manifestations. Dark colours correspond to 'major' manifestations, whereas dull colours correspond to 'minor' manifestations (see text). When the box is framed in bold, it means that the manifestation in question may be acute/subacute in presentation. Extrapyramidal involvement is hypokinetic when it corresponds to parkinsonism and hyperkinetic in the case of dystonia chorea or ballismus. Cobalamin metabolism defect: predominantly clbC and cblB deficiency. *Acute or subacute change in mental status (confusion or coma). †Compressive neuropathy. ‡Pseudo-syringomyelic syndrome, multifocal demyelinating polyneuropathy with conduction blocks. §Cranial neuropathy. The authors have created and have permission to use the image. A, axonal; AD GTP-CH1,autosomal dominant GTP cyclohydrolase 1; AMACR, alpha-methylacyl-CoA racemase; AN, auditory neuropathy; AVED, ataxia due to vitamin E deficiency; BG, basal ganglia signal abnormality; C, cataract; CA, cardiac arrythmia; CBS, cystathionine beta-synthase; CM, cardiomyopathy; CO,corneal opacities; CTX, cerebrotendinous xanthomatosis; CV, cornea verticillata; D, demyelinating; ERT, enzyme replacement therapy; HSCT, haematopoietic stem cell transplantation; HSGP, horizontal supranuclear gaze palsy; hyper, hyperkinetic; hypo, hypokinetic; I, insufficiency; KF, Keyser-Fleisher ring; LCFA, long-chain fatty acid; LCHAD, long-chain L-3 hydroxyacyl-CoA dehydrogenase deficiency; LD, lens dislocation; LDL, low-density lipoprotein; LE, leucoencephalopathy; LHON, Leber hereditary optic neuropathy; M, myelitis (spinal MRI T2 hyperintensities); MADD,multiple acyl-CoA dehydrogenase deficiency; MCAD, medium-chain acyl-coenzyme A dehydrogenase deficiency; MCT, medium-chain triglyceride; MNN, motor neuronopathy; MS, multiple sclerosis; MTHFR, methylenetetrahydrofolate reductase; MTP, mitochondrial trifunctional protein deficiency; ON, optic neuropathy; OTC, ornithine transcarbamylase; PBA, phenylbutyrate; PDC, pyruvate dehydrogenase complex; PKD, paroxysmal kinesigenic dyskinesia; RD, retinal dystrophy; Rhabdo, rhabdomyolysis; SFN, small-fibre neuropathy; SNN, sensory neuronopathy; SRT, substrate reduction therapy; TBRBGD, thiamin and biotin responsive basal ganglia disease; TCP, thrombocytopenia; UCD, urea cycle disorder; VSGP, vertical supranuclear gaze palsy; X-ALD, X-linked adrenoleucodystrophy.

patients with a genetically confirmed neurometabolic disease and an onset of neurological symptoms after 10 years since patients with a given neurometabolic disease and an onset of progressive neurological symptoms after this age usually present a rather homogeneous phenotype, which differs from earlieronset forms. $8-10$ We did not exclude as adult onset those patients with subtle and non-progressive symptoms before the age of 10 years of age that could go overlooked, such as mild intellectual disability.

Subsequently, we undertook a focused search in PubMed for articles in those same languages, using each previously identified adult-onset neurometabolic disease as a search term, with the aim to refine the adult-onset neurometabolic diseases selection and to identify those where a disease-modifying therapy is currently available (figure 1 and [online supplemental material](https://dx.doi.org/10.1136/jnnp-2021-328045) [2\)](https://dx.doi.org/10.1136/jnnp-2021-328045). We collected all clinical, MRI, biochemical and therapy data concerning these diseases. Although numerous and different classifications exist, these treatable adult-onset neurometabolic diseases were classified and presented in a clinically relevant

manner, meaning that disorders with similar clinical characteristics and common biochemical tests were grouped together.

REVIEW

KEY CONCEPTS IN NEUROMETABOLIC DISEASES Pathophysiology

Neurometabolic diseases are often due to an enzyme deficiency or dysfunction, potentially causing substrate accumulation upstream of the enzymatic block and a lack of downstream product synthesis (figure 2). If the accumulation of substrates is toxic at abnormally high concentrations, it may be responsible for clinical manifestations.⁶ These manifestations may be acute, as in ammonium accumulation in urea cycle disorders $(UCDs)^{11}$ or haeme precursors' accumulation in acute intermittent porphyria, 12 and/or progressive, as in the case of lysosomal storage disorders (LSD) .¹³ Furthermore, the toxic accumulation may also be due to a transport defect (as in Wilson and

Neurogenetics

Figure 2 Schematic representation of the pathophysiology and potential therapeutic strategies in neurometabolic diseases. In blue squares are specific treatments within the three main therapeutic strategies and the diseases where the treatment is indicated. *****Treatment is also a free-radical scavenger. **Treatment will also decrease substrate accumulation by downregulating its synthesis. The authors have created and have permission to use the image. A, substrate; AD GTP-CH1, autosomal dominant GTP cyclohydrolase 1; AMACR: alpha-methylacyl-CoA racemase; AVED, ataxia due to vitamin E deficiency; B, product; cblC, cobalamin C; CBS, cystathionine beta-synthase; CTX, cerebrotendinous xanthomatosis; E, enzyme; HSCT, haematopoietic stem cell transplantation; LCHAD, long-chain L-3 hydroxyacyl-CoA dehydrogenase; LHON, Leber hereditary optic neuropathy; MADD, multiple acyl-CoA dehydrogenase deficiency; MCAD, medium-chain acyl-coenzyme A dehydrogenase; MTHFR, methylenetetrahydrofolate reductase; MTP, mitochondrial trifunctional protein deficiency; OTC, ornithine transcarbamylase; PDC, pyruvate dehydrogenase complex; TBRBGD, thiamin and biotin responsive basal ganglia disease; X-ALD, X-linked adrenoleucodystrophy.

aceruloplasminaemia). In other neurometabolic diseases, it is the deficiency in a given enzymatic product that leads to disease, either directly, as in neurotransmitter synthesis disorders, or indirectly, due to impaired downstream cellular function like in pyruvate dehydrogenase complex (PDC) deficiency (in this case, in addition to lactic acidosis).15 Moreover, due to the complexity of metabolic networks, a deficiency in a given enzyme can also lead to less schematic metabolic derangements with both indirect increased toxic compounds as well as deficient ones. The observed enzyme deficiency is not always due to an anomaly of the enzyme itself, but it can also be explained by a defect in substrate or enzyme transport to the catabolic site (as in X-linked adrenoleucodystrophy (X-ALD) where peroxysomal transport of very long chain fatty acids is impaired)¹⁶; by a deficiency in a protein with the function of bringing in proximity the enzyme and its substrate (as in saposin deficiency, responsible for a metachromatic leucodystrophy with arylsulfatase normo-function) 17 ; or by an alteration in the metabolism of a vitamin required for the synthesis of a cofactor essential for the correct functioning of the enzyme (as the impaired synthesis of the 5-methyltetrahydrofolate, the active form of folate occurring in methyltetrahydrofolate reductase (MTHFR) deficiency and leading to the subsequent altered methionine synthase activity responsible for hyperhomocysteinaemia and hypomethioninaemia).10

Biochemical tools

Compared with other neurogenetic diseases, the diagnostic approach in neurometabolic diseases relies in the use of very numerous biochemical tests.⁴ These may be the measurement of one or a set of metabolites through a single test (such as total homocysteine or amino acid chromatography, respectively), of an enzymatic activity or of a functional test such as the mitochondrial respiration. Those tests may be performed in different fluids or tissues (including blood, urine, CSF, muscle or cultured cells, mainly skin-derived fibroblasts). Some biochemical studies are broad and point towards an overall alteration of a metabolic pathway; thus, additional analysis may be needed to identify the specific location of the metabolic block. These should also help guide and/or interpret the genetic study, whether it is Sanger sequencing of a specific gene or next-generation sequencing (NGS) of a metabolically oriented panel of genes.¹⁸ For example, an abnormal profile of blood acylcarnitines is suggestive of an altered mitochondrial beta-oxidation of fatty acids, and the nature of the specific abnormal acylcarnitines can point toward a specific enzyme within this pathway. Other biochemical studies can only screen for a specific neurometabolic disease, such as blood cholestanol in cerebrotendinous xanthomatosis (CTX), or the different specific enzymatic activities in LSDs, among which only the subgroups of mucopolysaccharidoses and oligosaccharidoses, almost exclusively of paediatric onset, can be screened

as a whole by analysing mucopolysaccharids and oligosaccharids in urine.¹⁹

Therapeutic strategies

Unlike most non-metabolic neurogenetic diseases, many neurometabolic diseases are treatable⁶; that is, the natural history of the disease can be modified by a specific therapeutic intervention. Some innovative treatments have been very recently proved to be efficient, and many therapeutic trials are ongoing in the field of neurometabolic diseases. Several and sometimes complementary therapeutic approaches exist (figure 2):

- 1. Restoration of a minimal enzymatic activity. It can be achieved by iterative intravenous infusion of the deficient enzyme as in Fabry and Pompe disease, known as *enzyme replacement therapy (*ERT).20 21 However, ERT is frequently unable to cross the blood–brain barrier and therefore has little impact on neurological symptoms. Various strategies are currently being studied to overcome this caveat, and recently, the intraventricular administration of ERT has been shown to be effective in neuronal ceroid lipofuscinosis type 2^{22} Depending on the neurometabolic disease, increasing the enzyme activity may be possible through other strategies than ERT: (1) the administration of a *high-dose vitamin or an enzymatic cofactor*, such as vitamin B6 (pyridoxine) in certain forms of classic homocystinuria (cystathionine beta-synthase deficiency)23; (2) the administration of a *molecule with a chaperone effect* that will limit enzyme degradation, like migalastat in Fabry disease or arimoclomol in Niemann-Pick C disease (that increases heat shock protein expression)^{24, 25}; (3) *haematopoietic stem cell transplant* of cells expressing a normal activity of the deficient enzyme, whether allogenous (classically used in X-ALD, Krabbe disease and metachromatic leucodystrophy) $26-28$ or ex vivo genetically modified autologous stem cells expressing the deficient enzyme in a supraphysiological manner (recent trials have demonstrated the efficacy of this latter strategy in children with X-ALD and metachromatic leucodystrophy $)^{29\,30}$; and (4) liver and/or kidney transplant with restricted indications in organic acidurias and UCDs. Without directly acting on enzyme activity, some treatments similarly aim to support the deficient metabolic function: this mainly concerns defects of energy metabolism (Glut1 deficiency, PDC deficiency and mitochondrial betaoxidation disorders) for which specific ketogenic diets can restore energy production, bypassing the altered energetic pathway.
- 2. Limiting or decreasing the accumulation of substrates upstream of the enzymatic block. This can be achieved by a specific *diet or regime*, such as the limitation of protein intake in UCDs and the consequent reduction of the ammonemia¹¹; by a *detoxifying drug* which binds to these substrates and limit their toxicity or facilitate their elimination, as is the case for sodium benzoate in $UCDs³¹$; or, still in UCDs, by extrarenal depuration to decrease ammoniaemia in the context of severe disease decompensation. In LSDs, drugs that inhibit the endogenous synthesis of these accumulated substrates can be used, which are known as *substrate reduction therapy*, like miglustat in Niemann-Pick C disease, which inhibits the ganglioside synthesis pathway.³² Similarly, a novel RNA interference therapeutic targeting hepatic delta-aminolevulinate synthetase 1 messenger RNA prevents the accumulation of haeme precursors in acute intermittent porphyria.³³
- 3. The *supply or substitution of the deficient product* downstream of the deficient enzymatic reaction, such as L-dopa

supplementation in autosomal dominant GTP cyclohydrolase I deficiency, also known as Segawa's disease or dopasensitive dystonia.³⁴

WHEN TO SUSPECT A NEUROMETABOLIC DISEASE IN ADULTS AND HOW TO PROCEED

As mentioned previously, early recognition and diagnosis of a treatable neurometabolic disease is particularly important. We will follow the algorithm presented in figure 3, which is the result of the exhaustive review of the literature added to the clinical experience of the authors, and should therefore be interpreted as such as it will not replace clinical reasoning for each individual patient. In practice, the adult neurologist can encounter three situations.

Characteristic presentation of a given neurometabolic disease In some situations, the presenting phenotype may be *immediately suggestive of a given adult-onset neurometabolic disease* (as described in figure 1 for treatable diseases): the specific biochemical and/or genetic test should be performed directly.

Genetic presentation or atypical acquired presentation

More frequently, *the clinical picture suggests a genetic disease* (family history of similar symptoms, parental consanguinity or a very chronic course), *or a commonly acquired aetiology is suspected in the first place but there are atypical features* leading to consider other less common disease mimics. In these situations, a neurometabolic aetiology should be considered if some of the following 'red flags' are present:

- 1. *Multiple neurological manifestations*, either involving several different neuroanatomical structures (eg, the association of epilepsy, indicating supratentorial disfunction, and cerebellar ataxia, indicating infratentorial involvement), or associated extraneurological signs and symptoms (visual, auditory, cardiac, hepatobiliary or endocrine). This is usually the case for most neurometabolic diseases. However, some more specific features can be found in particular subgroups of adult-onset neurometabolic diseases: in mitochondrial diseases (progressive external ophthalmoplegia, ptosis, stroke-like episodes, sensory neuronopathy, optic neuropathy, retinopathy, cataracts, sensorineural deafness and diabetes), 35 in LSDs (supranuclear gaze palsy, hepatosplenomegaly, facial dysmorphia and osteoarticular deformities) 13 and in peroxisomal disorders (demyelinating polyneuropathy, retinopathy, cataract, sensorineural deafness, facial dysmorphia and osteoarticular deformities). 36
- 2. *Acute or subacute neurological manifestations occurring in a context of basal metabolism modification* and/or increased energy demand, such as weight loss, prolonged fasting, drastic modification of diet, surgery, infection or drugs. This can occur in aminoacidopathies,³⁷ disorders of energy metabolism (including mitochondrial respiratory chain and betaoxidation disorders), 38 disorders of vitamin metabolism²³ and porphyrias.¹²
- 3. *Principal types of brain MRI findings* (figure 4):
	- A demyelinating cerebral white matter disorder with confluent, bilateral and more or less symmetrical T2 hyperintensities, in the absence of vascular risk factors, can suggest certain LSDs (Krabbe disease with involvement of the cortico-spinal tracts, 39 metachromatic leucodystrophy with frontal-predominant periventricular hyperintensities),⁴⁰ a specific peroxisomal disorder: X-ALD (with contrast enhancement in the inflammatory phase

Figure 3 Recommended algorithm to identify and evaluate adult patients with suspected neurometabolic diseases. *Movement disorders include extrapyramidal manifestations as well as isolated ataxia. **Genetic hyperhomocystinaemias include CBS deficiency, MTHFR deficiency and cobalamin metabolism defects. ***In parallel with diagnostic NGS if available or followed by confirmatory genetic testing if positive diagnostic work-up. The authors have created and have permission to use the image. CBS, cystathionine beta-synthase; CSF, cerebrospinal fluid; CTX, cerebrotendinous xanthomatosis; DWI, diffusion-weighted imaging; EMNG, electromyoneurography; GAG, glycosaminoglycan; HyperHcy, hyperhomocysteinaemia; LCHAD, long-chain L-3 hydroxyacyl-CoA dehydrogenase; LHON, Leber hereditary optic neuropathy; MADD, multiple acyl-CoA dehydrogenase deficiency; MTP, mitochondrial trifunctional protein deficiency; NGS, next-generation-sequencing; NP-C, Niemann-Pick type C; OS, oligosaccharides; PDE, pyridoxine-dependent epilepsy; UCD, urea cycle disorder; X-ALD, X-linked adrenoleucodystrophy; VLCFA, very long chain fatty acids.

of the disease), 41 several 'cerebral' organic acidurias (with periventricular hyperintensities and possible involvement of the U-fibres), $42\overline{43}$ as well as other diseases (see figure 1 for treatable ones) including CTX (with involvement of the dentate nuclei or the peridentate cerebellar white matter).⁴⁴ Furthermore, the presence of a bilateral and symmetrical often longitudinally extensive involvement of specific spinal cord tracts (such as subacute combined degeneration), associated or not with cerebral involvement, may also suggest a diagnosis of a treatable neurometabolic disease such as CTX, disorders of homocysteine remethylation (MTHFR deficiency and cobalamin C deficiency) 45 and biotinidase deficiency. 46

– A bilateral and symmetrical involvement of the deep grey matter or basal ganglia. These abnormalities can be seen in several disorders of energy metabolism including mitochondrial respiratory chain disorders (producing a Leigh syndrome if associated with encephalopathy), 47 PDC deficiency, 48 and thiamin and biotin responsive basal ganglia disease (TBRBGD)⁴⁹; in cerebral organic acidurias

(associated or not with white matter T2 hyperintensities), 42 and in Wilson's disease.⁵⁰

– Frequently, the brain MRI may be completely normal, and this should not rule out a neurometabolic disease if other clinical and paraclinical findings are present.

In these situations, the neurologist should first search for *subclinical signs* that may guide the diagnosis: mainly a spinal MRI in the context of a pathological brain MRI, a nerve conduction study with electromyography, an ophthalmological evaluation to detect any corneal, lens, retinal and optic nerve alterations, and an audiological evaluation including an audiogram and auditory evoked potentials (that can be abnormal in the absence of audiogram abnormalities). Second, depending on the context, perform other *complementary exams*, including *specific biochemical analysis*, according to two possible parallel diagnostic strategies:

We will follow a "metabolic" strategy if the context is suggestive of a particular subgroup of neurometabolic diseases as previously mentioned: in this case diagnostic tests exploring

Figure 4 Brain MRI illustrating various treatable adult-onset neurometabolic diseases. All MRIs originate from adult patients (19–60 years old). Krabbe disease (A1–2): T2-FLAIR signal abnormalities involving the periventricular and deep white matter with frequent involvement of the corticospinal tract. Metachromatic leucodystrophy (B1–2): T2-FLAIR showing a diffuse leucoencephalopathy in particular around the frontal horns with sparing of subcortical U fibres leading to a 'butterfly pattern'. X-linked adrenoleucodystrophy (C1–2): posterior leucoencephalopathy on T2-FLAIR (C1) and a peripheral border of active demyelination on gadolinium enhancement on T1W sequence (C2, arrow). Glutaric aciduria type 1 (D1–4): T2-FLAIR demonstrating a diffuse periventricular and deep white matter hyperintensity with lenticular (D2) and optic pathway (D3, arrow) signal abnormalities; enlargement of the convexity of subarachnoid spaces and sylvian fissures (D2) as well as bilateral temporal hypoplasia on T1W (D4). Cerebrotendinous xanthomatosis (E1–2): T2-FLAIR hyperintensity in the posterior limbs of both internal capsules (E1, arrow) and signal abnormalities within the dentate nuclei and the deep cerebellar white matter (E2). Methylenetetrahydrofolate reductase deficiency (F1–2): periventricular white matter signal abnormalities on T2-FLAIR (F1) and T1W (F2) sequences. Cerebrospinal fluid folate deficiency (G1–4): bilateral and symmetric leucoencephalopathy on T2-FLAIR sequences involving the supratentorial white matter and middle cerebellar peduncles. Spectroscopic analysis (G4) shows a rather low level of choline. Phenylketonuria (H1–3): periventricular and deep white matter leucoencephalopathy on T2-FLAIR predominantly located within posterior regions. Acute intermittent porphyria (I1–4) can sometimes be associated with posterior reversible encephalopathy syndrome (PRES) (I1–2). Complete disappearance of the PRES syndrome (I3–4) afterwards. Leber's hereditary optic neuropathy 'plus' disease (J1–2): T2-FLAIR MS-like signal abnormalities (J1) and T2 central hyperintensity of the optic chiasma (J2, arrow). Mitochondrial encephalomyopathy, lactic acidosis, and stroke-like episodes (K1–3): T2-FLAIR (K1–2) showing multifocal stroke-like cortical lesions in different stages of evolution involving multiple vascular territories. Increased signal on apparent diffusion coefficient MAP (K3, arrow) represents vasogenic rather than cytotoxic oedema. Wilson's disease (L1–2): bilateral and symmetrical midbrain (L2, arrow), heads of the caudate and putamen T2-FLAIR hyperintensities. Thiamin and biotin responsive basal ganglia disease (TBRBGD) (M1–3): acute episode of a diffuse vasogenic oedema with numerous hyper T2-FLAIR (M1,3) and diffusion cortical lesions (M2). The heads of the caudate and putamen (M2) are also involved. Fabry's disease (N1–N3): T2-FLAIR periventricular and deep white matter signal abnormalities. Compared with the baseline MRI (N1), the cerebral small vessel disease progresses on the 4-year follow-up MRI (N2) in a patient with no known cardiovascular disease. Ischaemic stroke sequelae of the posterior circulation (N3, arrow) can also be seen. The authors have created and have permission to use the image.T1W, T1 weighted.

Table 1 Key biochemical diagnostic tests for the diagnosis of treatable adult-onset neurometabolic diseases (clustered according to the explored

AASA, α-aminoadipic-δ-semialdehyde; AD GTP-CH1, autosomal dominant GTP cyclohydrolase 1; AMACR, alpha-methylacyl-CoA racemase; AVED, ataxia due to vitamin E deficiency; b, blood; CBS, cystathionine beta-synthase; CSF, cere xanthomatosis; DHCA dihytoxycholestanoic acid, LGHA). long-chain L-3 hydroxycy-l-CoA dehydrogenae deficiency, HOM Leer hereditary optine mempaty; LSM509, Jso-sphinogenyelin 509; byosob3, golootnaosysphinoshous MADD, multip basal ganglia disease; THCA, trihydroxycholestanoic acid; u, urine; X-ALD, X-linked adrenol

the suspected metabolic pathways will be performed (see figure 3 and table 1).

In parallel, we will follow a "neurological" strategy where the biochemical diagnostic work up will be performed in accordance with the combination of neurological manifestations (see figure 1 for treatable diseases). Three main criteria are to be used to judge if a biochemical test can be useful for the patient: the consistency of the clinical presentation as a whole with the tested disease, the invasiveness and reliability (in terms of sensitivity and specificity) of the test, and the existence of a treatment. In this line figure 1 shows the different clinical manifestations of all potentially treatable adult-onset neurometabolic diseases categorised as major (if they are frequent or predominant within the clinical picture, and potentially be found isolated) or minor (if infrequent, even subclinical and rarely found isolated), as well as acute or chronic onset. Table 1 specifies the limitations of some screening tests. In the case of a treatable neurometabolic disease, it is probably legitimate to test not only if the presentation is reminiscent of the disease but also if it is merely compatible.

Unexplained isolated manifestation

Finally, some neurometabolic diseases may have an *unspecific isolated neurological manifestation* without the aforementioned clinical and paraclinical hints, compatible with either a genetic or acquired aetiology. For example, adrenomyeloneuropathy (the spinal form of X-ALD) can present as an isolated spastic paraparesis, 51 and rarely, this is may also be the case for CTX. 52 Niemann-Pick type C disease may present with isolated psychosis or a movement disorder.⁸ Certain neurometabolic diseases, like Tangier (possibly mimicking chronic idiopathic demyelinating polyneuropathy) or Pompe disease, may present with isolated neuromuscular signs and symptoms.^{53 54} In these cases, if the diagnosis is not obvious after first-line investigations, it may be useful to carry out a biochemical analysis of the possible neurometabolic diseases, in parallel to the relevant genetic study if necessary (eg, a 'spastic paraplegia' panel).

Enseignement Superieur (ABES). Protected by copyright. J Neurol Neurosurg Psychiatry: first published as 10.1136/jnnp-2021-328045 on 9 February 2022. Downloaded from <http://jnnp.bmj.com/> on February 14, 2022 at Agence Bibliographique de

Neurogenetics

The recommended diagnostic approach is summarised in figure 3. If a biochemical test was to be positive, an evaluation by a specialist in the field should be considered to interpret the result within the clinical context and to guide the subsequent diagnostic process, whether further biochemical studies are needed or a genetic study can be undertaken. It should be noted that for some neurometabolic diseases, no reliable biochemical diagnostic test is available, and it is the genetic study alone that can confirm the disease. In these cases, it may be possible to perform a therapeutic test, of outmost importance in the case of neurometabolic diseases with possible acute/subacute presentations where benefit from early treatment can be dramatic. This is the case for TBRBGD and riboflavin transporter deficiency.²³ Finally, the clinician should take into account the prevalence of individual neurometabolic diseases in their region, as well as the frequency of consanguinity and the presence of founding mutations.

PLACE FOR NGS GENE TECHNOLOGIES AND METABOLOMICS IN THE DIAGNOSIS OF NEUROMETABOLIC DISEASES

Neurometabolic diseases are predominantly, but not exclusively, single-gene recessive disorders. NGS technologies now allow for rapid and inexpensive large-scale genomic analysis of rare diseases.⁵⁵ The first and most widespread technology is that of a panel of genes, allowing for the sequencing of hundreds of known genes implicated in neurometabolic diseases. If available, an alternative to gene panels is the sequencing of the entire set of protein-coding sequences or exome, known as wide exome sequencing or to directly sequence the entire 6billion base pair human genome through wide genome sequencing. NGS has facilitated the detection of new gene–phenotype associations, expanding the clinical spectrum of already established Mendelian disorders.¹⁸ All of the aforementioned has increased our ability to correctly diagnose patients, has decreased diagnostic delay and reduced overall diagnostic expenses.⁵⁶ Furthermore, NGS will also allow to detect genetic modifiers that could explain the phenotypical heterogeneity encountered within patients suffering from the same disease, even among siblings carrying the same pathogenic variants. 57

However, the increased genetic resolution and complexity of the data comes with an increased detection of variants of unknown significance. This is why a combined approach with deep phenotyping of patients is needed. Concerning intellectual deficiency with a suspected metabolic origin, both clinical and biochemical phenotyping reached a high diagnostic yield of 68% ,¹⁸ whereas similar exome studies without deep phenotyping usually result in a lower diagnostic yield of around 16%.58 To fit with this strategy, NGS, if easily available, could be performed in parallel to targeted biochemical phenotyping in patients with suspected neurometabolic diseases in the context of our proposed diagnostic approach. Complementary biochemical tests may also be needed once NGS results are available to help in the interpretation of variants of interest.

In addition, biochemical phenotyping could also be enlarged by metabolomics, establishing a comprehensive biochemical profile.⁵⁹ The high-dimensional nature of the data amassed through metabolomics requires the use of dedicated preprocessing platforms and multivariate analysis or even machine learning tools to classify the findings.⁶⁰ This untargeted approach has the potential to direct the diagnosis among varied and unspecific manifestations and to diminish the number of specific biochemical tests, thus reducing the overall cost. 61 In addition,

it could provide complementary biochemical data to assist in the interpretation of genetic variants from NGS gene studies. 62 Although these metabolomics approaches are powerful, they have not yet been translated into clinical practice.

CONCLUSIONS

Adult neurometabolic diseases encompass a heterogeneous set of conditions for which we have presented a synthetic overview of the different phenotypes, especially for those with a currently available treatment, as well as a simplified diagnostic approach. Nevertheless, this practical knowledge is bound to change over the next years, with the identification of new neurometabolic diseases, the report of new phenotypes for known ones, the accessibility of untargeted diagnostic tools (genomics and metabolomics) and the discovery of new treatments for currently untreatable neurometabolic diseases. We hope that this review will increase awareness of this group of diseases and allow for an efficient and rational use of the biochemical tests available in the diagnosis of neurometabolic diseases, ultimately leading to prompt diagnosing and early treatment of patients.

Twitter Gorka Fernández-Eulate [@Gorkaeulate](https://twitter.com/Gorkaeulate)

Contributors Concept and design, drafting of the manuscript: GF-E, CC YN. Administrative, technical, or material support: CC, NS and FL. Supervision: GF-E and YN. Acquisition, analysis or interpretation of data, critical revision of the manuscript for important intellectual content: all authors.

Funding The authors have not declared a specific grant for this research from any funding agency in the public, commercial or not-for-profit sectors.

Competing interests YN reported grants from Takeda and Orchard therapeutics and has received honoraria from Sanofi Genzyme and Orphazyme. No other disclosures were reported.

Patient consent for publication Not applicable.

Ethics approval This study does not involve human participants.

Provenance and peer review Not commissioned; externally peer reviewed.

Supplemental material This content has been supplied by the author(s). It has not been vetted by BMJ Publishing Group Limited (BMJ) and may not have been peer-reviewed. Any opinions or recommendations discussed are solely those of the author(s) and are not endorsed by BMJ. BMJ disclaims all liability and responsibility arising from any reliance placed on the content. Where the content includes any translated material, BMJ does not warrant the accuracy and reliability of the translations (including but not limited to local regulations, clinical guidelines, terminology, drug names and drug dosages), and is not responsible for any error and/or omissions arising from translation and adaptation or otherwise.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: [http://creativecommons.org/licenses/by-nc/4.0/.](http://creativecommons.org/licenses/by-nc/4.0/)

ORCID iDs

Gorka Fernández-Eulate<http://orcid.org/0000-0003-1112-7464> Christophe Carreau <http://orcid.org/0000-0001-7000-9367> Yann Nadjar<http://orcid.org/0000-0003-1466-6957>

REFERENCES

- 1 Carreau C, Benoit C, Ahle G, et al. Late-onset riboflavin transporter deficiency: a treatable mimic of various motor neuropathy aetiologies. J Neurol Neurosurg [Psychiatry](http://dx.doi.org/10.1136/jnnp-2020-323304) 2021;92:27–35.
- 2 Sirrs SM, Lehman A, Stockler S, et al. Treatable inborn errors of metabolism causing neurological symptoms in adults. [Mol Genet Metab](http://dx.doi.org/10.1016/j.ymgme.2013.10.002) 2013;110:431–8.
- 3 Masingue M, Benoist J-F, Roze E, et al. Cerebral folate deficiency in adults: a heterogeneous potentially treatable condition. [J Neurol Sci](http://dx.doi.org/10.1016/j.jns.2018.11.014) 2019;396:112-8.
- 4 Guerrero RB, Salazar D, Tanpaiboon P. Laboratory diagnostic approaches in metabolic disorders. [Ann Transl Med](http://dx.doi.org/10.21037/atm.2018.11.05) 2018;6:470.
- 5 Gahl WA, Markello TC, Toro C, et al. The national institutes of health undiagnosed diseases program: insights into rare diseases. [Genet Med](http://dx.doi.org/10.1038/gim.0b013e318232a005) 2012;14:51-9.
- 6 Sedel F, Lyon-Caen O, Saudubray J-M. Therapy insight: inborn errors of metabolism in adult neurology--a clinical approach focused on treatable diseases. Nat Clin Pract [Neurol](http://dx.doi.org/10.1038/ncpneuro0494) 2007;3:279–90.
- sciences 1103 clinical sciences. Orphanet J Rare Dis 2018;13. 9 Masingue M, Dufour L, Lenglet T, et al. Natural history of adult patients with GM2 gangliosidosis. [Ann Neurol](http://dx.doi.org/10.1002/ana.25689) 2020;87:609–17.
- 10 Gales A, Masingue M, Millecamps S, et al. Adolescence/adult onset MTHFR deficiency may manifest as isolated and treatable distinct neuro-psychiatric syndromes. [Orphanet J Rare Dis](http://dx.doi.org/10.1186/s13023-018-0767-9) 2018;13:1–8.
- 11 Matsumoto S, Häberle J, Kido J, et al. Urea cycle disorders-update. [J Hum Genet](http://dx.doi.org/10.1038/s10038-019-0614-4) 2019;64:833–47.
- 12 Puy H, Gouya L, Deybach J-C. Porphyrias. [Lancet](http://dx.doi.org/10.1016/S0140-6736(09)61925-5) 2010;375:924-37.

[Hum Genet](http://dx.doi.org/10.1038/ejhg.2017.35) 2017;25:788.

- 13 Platt FM, d'Azzo A, Davidson BL, et al. Lysosomal storage diseases. Nat Rev Dis [Primers](http://dx.doi.org/10.1038/s41572-018-0025-4) 2018;4.
- 14 Trender-Gerhard I, Sweeney MG, Schwingenschuh P, et al. Autosomal-dominant GTPCH1-deficient DRD: clinical characteristics and long-term outcome of 34 patients. [J Neurol Neurosurg Psychiatry](http://dx.doi.org/10.1136/jnnp.2008.155861) 2009;80:839–45.
- 15 Patel KP, O'Brien TW, Subramony SH, et al. The spectrum of pyruvate dehydrogenase complex deficiency: clinical, biochemical and genetic features in 371 patients. [Mol](http://dx.doi.org/10.1016/j.ymgme.2011.09.032) [Genet Metab](http://dx.doi.org/10.1016/j.ymgme.2011.09.032) 2012;105:34–43.
- 16 Turk BR, Theda C, Fatemi A, et al. X-linked adrenoleukodystrophy: pathology, pathophysiology, diagnostic testing, newborn screening and therapies. Int J Dev [Neurosci](http://dx.doi.org/10.1002/jdn.10003) 2020;80:52–72.
- 17 Fenu S, Castellotti B, Farina L, et al. Saposin B deficiency as a cause of adult-onset metachromatic leukodystrophy. [Neurology](http://dx.doi.org/10.1212/WNL.0000000000007951) 2019;93:310-2.
- 18 Tarailo-Graovac M, Shyr C, Ross CJ, et al. Exome sequencing and the management of neurometabolic disorders. [N Engl J Med](http://dx.doi.org/10.1056/NEJMoa1515792) 2016;374:2246–55.
- 19 Sedel F. Inborn errors of metabolism in adults: A diagnostic approach to neurological and psychiatric presentations. In: [Inborn metabolic diseases: diagnosis and treatment](http://dx.doi.org/10.1007/978-3-642-15720-2_2). Springer Berlin Heidelberg, 2012: 57–74.
- 20 Eng CM, Guffon N, Wilcox WR, et al. Safety and efficacy of recombinant human alpha-galactosidase a replacement therapy in Fabry's disease. [N Engl J Med](http://dx.doi.org/10.1056/NEJM200107053450102) 2001;345:9-16.
- 21 van der Ploeg AT, Clemens PR, Corzo D, et al. A randomized study of alglucosidase alfa in late-onset Pompe's disease. [N Engl J Med](http://dx.doi.org/10.1056/NEJMoa0909859) 2010;362:1396-406.
- 22 Schulz A, Ajayi T, Specchio N, et al. Study of intraventricular Cerliponase alfa for CLN2 disease. [N Engl J Med](http://dx.doi.org/10.1056/NEJMoa1712649) 2018;378:1898–907.
- 23 Mandia D, Shor N, Benoist J-F, et al. Adolescent-onset and adult-onset vitamin-responsive neurogenetic diseases: a review. [JAMA Neurol](http://dx.doi.org/10.1001/jamaneurol.2020.4911) 2021;78:483.
- 24 Germain DP, Hughes DA, Nicholls K, et al. Treatment of Fabry's disease with the pharmacologic chaperone Migalastat. [N Engl J Med](http://dx.doi.org/10.1056/NEJMoa1510198) 2016;375:545-55.
- 25 Mengel E, Patterson MC, Da Riol RM. Efficacy and safety of arimoclomol in Niemann-Pick disease type C: results from a double-blind, randomised, placebo-controlled, multinational phase 2/3 trial of a novel treatment. [J Inherit Metab Dis](http://dx.doi.org/10.1002/jimd.12428) 2021:1–18.
- 26 Peters C, Charnas LR, Tan Y, et al. Cerebral X-linked adrenoleukodystrophy: the international hematopoietic cell transplantation experience from 1982 to 1999. [Blood](http://dx.doi.org/10.1182/blood-2003-10-3402) 2004;104:881–8.
- 27 Krivit W, Shapiro E, Kennedy W, et al. Treatment of late infantile metachromatic leukodystrophy by bone marrow transplantation. [N Engl J Med](http://dx.doi.org/10.1056/NEJM199001043220106) 1990;322:28-32.
- 28 Krivit W, Shapiro EG, Peters C, et al. Hematopoietic stem-cell transplantation in globoid-cell leukodystrophy. [N Engl J Med](http://dx.doi.org/10.1056/NEJM199804163381605) 1998;338:1119-27.
- 29 Eichler F, Duncan C, Musolino PL, et al. Hematopoietic stem-cell gene therapy for cerebral adrenoleukodystrophy. [N Engl J Med](http://dx.doi.org/10.1056/NEJMoa1700554) 2017;377:1630-8.
- 30 Biffi A, Montini E, Lorioli L, et al. Lentiviral hematopoietic stem cell gene therapy benefits metachromatic leukodystrophy. [Science](http://dx.doi.org/10.1126/science.1233158) 2013;341:1233158.
- 31 Enns GM, Berry SA, Berry GT, et al. Survival after treatment with phenylacetate and benzoate for urea-cycle disorders. [N Engl J Med](http://dx.doi.org/10.1056/NEJMoa066596) 2007;356:2282–92.
- 32 Patterson MC, Vecchio D, Prady H, et al. Miglustat for treatment of Niemann-Pick C disease: a randomised controlled study. [Lancet Neurol](http://dx.doi.org/10.1016/S1474-4422(07)70194-1) 2007;6:765-72.
- 33 Balwani M, Sardh E, Ventura P, et al. Phase 3 trial of RNAi therapeutic Givosiran for acute intermittent porphyria. [N Engl J Med](http://dx.doi.org/10.1056/NEJMoa1913147) 2020;382:2289-301.
- 34 Nygaard TG, Marsden CD, Fahn S. Dopa-responsive dystonia: long-term treatment response and prognosis. [Neurology](http://dx.doi.org/10.1212/wnl.41.2_part_1.174) 1991;41:174-81.
- 35 Gorman GS, Chinnery PF, DiMauro S, et al. Mitochondrial diseases. Nat Rev Dis
- 36 Waterham HR, Ferdinandusse S, Wanders RJA. Human disorders of peroxisome metabolism and biogenesis. [Biochim Biophys Acta - Mol Cell Res](http://dx.doi.org/10.1016/j.bbamcr.2015.11.015) 1863;2016:922–33.
- 37 Wasim M, Awan FR, Khan HN, et al. Aminoacidopathies: prevalence, etiology, screening, and treatment options. [Biochem Genet](http://dx.doi.org/10.1007/s10528-017-9825-6) 2018;56:7-21.
- 38 Feillet F, Steinmann G, Vianey-Saban C, et al. Adult presentation of MCAD deficiency revealed by coma and severe arrythmias. [Intensive Care Med](http://dx.doi.org/10.1007/s00134-003-1871-3) 2003;29:1594–7.
- 39 Cousyn L, Law-Ye B, Pyatigorskaya N, et al. Brain MRI features and scoring of leukodystrophy in adult-onset Krabbe disease. [Neurology](http://dx.doi.org/10.1212/WNL.0000000000007943) 2019;93:e647–52.
- 40 Groeschel S, Kehrer C, Engel C, et al. Metachromatic leukodystrophy: natural course of cerebral MRI changes in relation to clinical course. *[J Inherit Metab Dis](http://dx.doi.org/10.1007/s10545-011-9361-1)* 2011;34:1095–102.
- 41 Loes DJ, Fatemi A, Melhem ER, et al. Analysis of MRI patterns AIDS prediction of progression in X-linked adrenoleukodystrophy. [Neurology](http://dx.doi.org/10.1212/01.WNL.0000079050.91337.83) 2003;61:369–74.
- 42 Garbade SF, Greenberg CR, Demirkol M, et al. Unravelling the complex MRI pattern in glutaric aciduria type I using statistical models-a cohort study in 180 patients. [J Inherit](http://dx.doi.org/10.1007/s10545-014-9676-9) [Metab Dis](http://dx.doi.org/10.1007/s10545-014-9676-9) 2014;37:763–73.
- 43 Kölker S, Christensen E, Leonard JV, et al. Diagnosis and management of glutaric aciduria type I – revised recommendations. [J Inherit Metab Dis](http://dx.doi.org/10.1007/s10545-011-9289-5) 2011;34:677-94.
- 44 Barkhof F, Verrips A, Wesseling P, et al. Cerebrotendinous xanthomatosis: the spectrum of imaging findings and the correlation with neuropathologic findings. [Radiology](http://dx.doi.org/10.1148/radiology.217.3.r00dc03869) 2000;217:869–76.
- 45 Hao M, Zhang Y, Hou S, et al. Spinal cord demyelination combined with hyperhomocysteinemia: a case report. [Neuropsychiatr Dis Treat](http://dx.doi.org/10.2147/NDT.S70963) 2014;10:2057-9.
- 46 Bottin L, Prud'hon S, Guey S, et al. Biotinidase deficiency mimicking neuromyelitis optica: initially exhibiting symptoms in adulthood. [Mult Scler](http://dx.doi.org/10.1177/1352458515596457) 2015;21:1604-7.
- 47 Baertling F, Rodenburg RJ, Schaper J, et al. A guide to diagnosis and treatment of Leigh syndrome. [J Neurol Neurosurg Psychiatry](http://dx.doi.org/10.1136/jnnp-2012-304426) 2014;85:257-65.
- 48 Wijburg FA, Barth PG, Bindoff LA, et al. Leigh syndrome associated with a deficiency of the pyruvate dehydrogenase complex: results of treatment with a ketogenic diet. [Neuropediatrics](http://dx.doi.org/10.1055/s-2008-1071331) 1992;23:147–52.
- 49 Tabarki B, Al-Shafi S, Al-Shahwan S, et al. Biotin-responsive basal ganglia disease revisited: clinical, radiologic, and genetic findings. [Neurology](http://dx.doi.org/10.1212/WNL.0b013e31827deb4c) 2013;80:261-7.
- 50 Zhong W, Huang Z, Tang X. A study of brain MRI characteristics and clinical features in 76 cases of Wilson's disease. [J Clin Neurosci](http://dx.doi.org/10.1016/j.jocn.2018.10.096) 2019;59:167-74.
- 51 Ciarlariello VB, de Freitas JL, Pedroso JL, et al. X-Linked adrenoleukodystrophy mimicking hereditary spastic paraplegia. [Mov Disord Clin Pract](http://dx.doi.org/10.1002/mdc3.12858) 2020;7:109-10.
- 52 Nicholls Z, Hobson E, Martindale J, et al. Diagnosis of spinal xanthomatosis by next-generation sequencing: identifying a rare, treatable mimic of hereditary spastic paraparesis. [Pract Neurol](http://dx.doi.org/10.1136/practneurol-2015-001117) 2015;15:280-3.
- 53 Chan J, Desai AK, Kazi ZB, et al. The emerging phenotype of late-onset Pompe disease: a systematic literature review. [Mol Genet Metab](http://dx.doi.org/10.1016/j.ymgme.2016.12.004) 2017;120:163-72.
- 54 Mercan M, Yayla V, Altinay S, et al. Peripheral neuropathy in Tangier disease: a literature review and assessment. [J Peripher Nerv Syst](http://dx.doi.org/10.1111/jns.12265) 2018;23:88-98.
- 55 Rexach J, Lee H, Martinez-Agosto JA, et al. Clinical application of next-generation sequencing to the practice of neurology. [Lancet Neurol](http://dx.doi.org/10.1016/S1474-4422(19)30033-X) 2019;18:492-503.
- 56 Soden SE, Saunders CJ, Willig LK, et al. Effectiveness of exome and genome sequencing guided by acuity of illness for diagnosis of neurodevelopmental disorders. [Sci Transl Med](http://dx.doi.org/10.1126/scitranslmed.3010076) 2014;6:265ra168.
- 57 Rahit KMTH, Tarailo-Graovac M. Genetic modifiers and rare Mendelian disease. [Genes](http://dx.doi.org/10.3390/genes11030239) 2020;11. doi:10.3390/genes11030239. [Epub ahead of print: 25 02 2020].
- 58 de Ligt J, Willemsen MH, van Bon BWM, et al. Diagnostic exome sequencing in persons with severe intellectual disability. [N Engl J Med](http://dx.doi.org/10.1056/NEJMoa1206524) 2012;367:1921-9.
- Mordaunt D, Cox D, Fuller M. Metabolomics to improve the diagnostic efficiency of inborn errors of metabolism. [Int J Mol Sci](http://dx.doi.org/10.3390/ijms21041195) 2020;21:1195.
- 60 Mendez KM, Reinke SN, Broadhurst DI. A comparative evaluation of the generalised predictive ability of eight machine learning algorithms across ten clinical metabolomics data sets for binary classification. [Metabolomics](http://dx.doi.org/10.1007/s11306-019-1612-4) 2019;15:150.
- 61 Miggiels P, Wouters B, van Westen GJP. Novel technologies for metabolomics: More for less. TrAC - Trends Anal. [Chem](http://dx.doi.org/10.1016/j.trac.2018.11.021) 2019;120:115323.
- 62 Stark Z, Tan TY, Chong B, et al. A prospective evaluation of whole-exome sequencing as a first-tier molecular test in infants with suspected monogenic disorders. Genet [Med](http://dx.doi.org/10.1038/gim.2016.1) 2016;18:1090–6.

Supplementary material 1: List of all adult-onset neurometabolic diseases. In bold, those with a disease-modifying therapy currently available in clinical practice. Adapted from: *Hollak CE, Lachmann R, editors. Inherited metabolic disease in adults: A clinical guide. Oxford University Press; 2016.*

DISORDERS OF CARBOHYDRATE METABOLISM

Glycogen storage disorders 0-XIII, including **Pompe**, Cori/Forbes, Andersen or Adult polyglucosan body disease and McArdle disease Galactosemia **GLUT1 deficiency**

DISORDERS OF MITOCHONDRIAL ENERGY METABOLISM

Pyruvate dehydrogenase complex deficiency

Disorders of mitochondrial energy metabolism: Mitochondrial encephalomyopathy with lactic acidosis and stroke-like episodes (MELAS), Myoclonic epilepsy with ragged-red fibers (MERRF), Neurogenic weakness with ataxia and retinits pigmentosa (NARP), Leigh syndrome (subacute necrotizing encephalomyelopathy), **Leber hereditary optic neuropathy (LHON)**, Progressive external ophtalmoplegia (PEO), Kearns-Sayre syndrome (KSS), Sensory ataxic neuropathy, dysarthria and ophtalmoparesis (SANDO), Myoclonic epilepsy, myopathy, sensory ataxia (MEMSA), Mitochondrial neurogastrointestinal encephalomyopathy (MNGIE)

Fatty acid oxidation: Carnitine deficiency carnitine palmitoyltransferase 1A (CPT1A) deficiency, carnitine-acylcarnitine translocase (CACT) deficiency, carnitine palmitoyltransferase 2 (CPT2) deficiency, Very long-chain acyl-CoA dehydrogenase (VLCAD) deficiency, **Long-chain 3-hydroxyacyl-CoA dehydrogenase (LCHAD) deficiency, Mitochondrial trifunctional protein (MTP) deficiency**, **Medium-chain acyl CoA dehydrogenase (MCAD) deficiency**, Short-chain enoyl-CoA dehydrogenase (SCAD) deficiency, 3-hydroxyacyl-CoA dehydrogenase (HADH) deficiency Electron transfer defects: **Multiple acyl-CoA dehydrogenase (MADD) deficiency**

Riboflavin metabolism defects: **Brown-Vialetto-Van Laere (BVVL) syndrome**, FAD synthetase deficiency, MFT deficiency 81

Disorders of ketogenesis and ketolysis: 3-hydroxyl-3-methylglutaryl-CoA (HMG-CoA) synthase deficiency, HMG-CoA lyase deficiency

Disorders of creatine metabolism: AGAT deficiency, GAMT deficiency, CrT defect **Coenzyme Q10 deficiency**

DISORDERS OF PROTEIN METABOLISM

Phenylketonuria

Maple syrup urine disease, Proionic acidemia, Methlymalonic acidemia and Isovaleric acidemia

Urea cycle disorders, Hyperornithinemia-hyperammonemia-homocitrullinuria syndrome Citrin deficiency

Cystathionine beta-synthase deficiency or Homocystinuria

Cerebral organic acidurias: Glutaric aciduria type I and 2-hydrohyglutatic acidurias

Lysinuric protein intolerance and Hartnup disease

DISORDERS OF VITAMIN METABOLISM **Biotinidase deficiency Disorders of cobalamin: CblC and others Folate metabolism: Methylenetetrahydrofolate reductase (MTHFR) deficiency Disorders of thiamin metabolism: Biotin-thiamin responsive basal ganglia disease**

NEUROTRASMITTERS

Succinic semialdehyde dehydrogenase deficiency Adult-onset monoamine disorders: **Autosomal dominant GTP cyclohydrolase deficiency** Brain serotonin deficiency

DYSLIPIDEMIAS **Tangier disease Abetalipoproteinemia**

BILE ACID SYNTHESIS DEFECTS

Cerebrotendinous xanthomatosis Spastic paraplegia (SPG) type 5

DISORDERS OF PURINE METABOLISM Lesch-Nyhan disease and variants

PORPHYRIAS Acute intermittent porphyria

MINERAL AND METAL METABOLISM DISORDERS Disorders of copper and iron metabolism: Neuroferritinopathy, **Wilson disease, Acerulopasminemia** Disorders of manganese metabolism: **SLC30A10 deficiency**

LYOSOMAL STORAGE DISORDERS **Fabry disease Gaucher disease type III** GM1-gangliosidosis type II and III

GM2-gangliosidosis (Tay-Sachs and Sandhoff disease) **Krabbe disease Metachromatic leukodystrophy (ARSA and Saposin-C deficiency) Niemann type C Pompe disease** Neuronal Ceroid Lipofuscinosis (mainly CLN2, CLN3, CLN4, CLN5, CLN6, CLN7, CLN11, CLN13). Sialidosis or Mucolipidosis type 1

PEROXISOMAL DISORDERS

X-linked adrenoleukodystrophy and adrenomyeloneuropathy Refsum disease 2-methylacyl-CoA racemase (AMACR) deficiency

CONGENITAL DISORDERS OF GLYCOSYLATION (CDG)

Only some genes like *DPGAT1* congenital myasthenia

Supplementary material 2: List of references for Figure 1 and Table 1.

References

- 1 Wijemanne S, Jankovic J. Dopa-responsive dystonia Clinical and genetic heterogeneity. *Nat Rev Neurol* 2015;**11**:414–24. doi:10.1038/nrneurol.2015.86
- 2 Trender-Gerhard I, Sweeney MG, Schwingenschuh P, *et al.* Autosomal-dominant GTPCH1-deficient DRD: Clinical characteristics and long-term outcome of 34 patients. *J Neurol Neurosurg Psychiatry* 2009;**80**:839–45. doi:10.1136/jnnp.2008.155861
- 3 Hyland K. Clinical Utility of Monoamine Neurotransmitter Metabolite Analysis in Cerebrospinal Fluid. *Clin Chem* 2008;**54**:633–41. doi:10.1373/clinchem.2007.099986
- 4 Gelener P, Severino M, Diker S, *et al.* Adult-onset glutaric aciduria type I: rare presentation of a treatable disorder. *neurogenetics 2020 213* 2020;**21**:179–86. doi:10.1007/S10048-020-00610-9
- 5 Tabarestani S, Varriano B, Rawal S, *et al.* Seizures and early onset dementia: D2HGA1 inborn error of metabolism in adults. *Ann Clin Transl Neurol* 2020;**7**:2052. doi:10.1002/ACN3.51162
- 6 Külkens S, Harting I, Sauer S, *et al.* Late-onset neurologic disease in glutaryl-CoA dehydrogenase deficiency. *Neurology* 2005;**64**:2142–4. doi:10.1212/01.WNL.0000167428.12417.B2
- 7 Bähr O, Mader I, Zschocke J, *et al.* Adult onset glutaric aciduria type I presenting with a leukoencephalopathy. *Neurology* 2002;**59**:1802–4. doi:10.1212/01.WNL.0000036616.11962.3C
- 8 Fujitake J, Ishikawa Y, Fujii H, *et al.* l-2-Hydroxyglutaric aciduria: two Japanese adult cases in one family. *J Neurol 1999 2465* 1999;**246**:378–82. doi:10.1007/S004150050367
- 9 Mainka T, Ziagaki A, Koning TJ de, *et al.* The Wide Phenotypic Spectrum of L-2 Hydroxyglutaric Aciduria in Adults. *Mov Disord Clin Pract* 2020;**7**:1004–6. doi:10.1002/MDC3.13092
- 10 Tsai FC, Lee HJ, Wang AG, *et al.* Experiences during newborn screening for glutaric aciduria type 1: Diagnosis, treatment, genotype, phenotype, and outcomes. *J Chinese Med Assoc* 2017;**80**:253–61. doi:10.1016/J.JCMA.2016.07.006
- 11 Pierson TM, Nezhad M, Tremblay MA, *et al.* Adult-onset glutaric aciduria type I presenting with white matter abnormalities and subependymal nodules. *Neurogenetics* 2015;**16**:325–8. doi:10.1007/s10048-015-0456-y
- 12 Herskovitz M, Goldsher D, Sela B-A, *et al.* Subependymal mass lesions and peripheral polyneuropathy in adult-onset glutaric aciduria type I. *Neurology* 2013;**81**:849–50. doi:10.1212/WNL.0B013E3182A2CBF2
- 13 Marcel C, Mallaret M, Lagha-Boukbiza O, *et al.* L-2-hydroxyglutaric aciduria diagnosed in a young adult with progressive cerebellar ataxia and facial dyskinesia. *Rev Neurol (Paris)* 2012;**168**:187–91. doi:10.1016/J.NEUROL.2011.06.002
- 14 Karatas H, Saygi S, Bastan B, *et al.* L-2-hydroxyglutaric aciduria: Report of four Turkish adult patients. *Neurologist* 2010;**16**:44–6. doi:10.1097/NRL.0B013E31819F9556
- 15 Angle B, Burton BK. Risk of sudden death and acute life-threatening events in patients with glutaric acidemia type II. *Mol Genet Metab* 2008;**93**:36–9. doi:10.1016/j.ymgme.2007.09.015
- 16 Periasamy V, Rudwan M, Yadav G, *et al.* Epilepsy in a Young Adult Caused by L-2- Hydroxyglutaric Aciduria: A Case Report. *Med Princ Pract* 2008;**17**:258–61. doi:10.1159/000117804
- 17 Chen S, Zhu M, Hao Y, *et al.* Effect of Delayed Diagnosis of Phenylketonuria With Imaging Findings of Bilateral Diffuse Symmetric White Matter Lesions: A Case Report and Literature Review. *Front Neurol* 2019;**0**:1040. doi:10.3389/FNEUR.2019.01040
- 18 Wang C, Li J. Subacute onset leukodystrophy and visual-spatial disorders revealing phenylketonuria combined with homocysteinmia in adulthood. *Med (United States)* 2018;**97**:1–5. doi:10.1097/MD.0000000000009801
- 19 Tufekcioglu Z, Cakar A, Bilgic B, *et al.* Adult-onset phenylketonuria with rapidly progressive dementia and parkinsonism. *Neurocase* 2016;**22**:273–5. doi:10.1080/13554794.2016.1142567
- 20 Rosini F, Rufa A, Monti L, *et al.* Adult-onset phenylketonuria revealed by acute reversible dementia, prosopagnosia and parkinsonism. J. Neurol. 2014;**261**:2446–8. doi:10.1007/s00415-014-7492-7
- 21 Vockley J, Andersson HC, Antshel KM, *et al.* Phenylalanine hydroxylase deficiency: Diagnosis and management guideline. *Genet Med* 2014;**16**:188–200. doi:10.1038/gim.2013.157
- 22 Bilder DA, Burton BK, Coon H, *et al.* Psychiatric symptoms in adults with phenylketonuria. *Mol Genet Metab* 2013;**108**:155–60. doi:10.1016/J.YMGME.2012.12.006
- 23 Kasim S, Moo LR, Zschocke J, *et al.* Phenylketonuria presenting in adulthood as progressive spastic paraparesis with dementia. *J Neurol Neurosurg Psychiatry* 2001;**71**:795–7. doi:10.1136/JNNP.71.6.795
- 24 Anderson D, Jain-Ghai S, Sligl WI. Adult-onset presentation of a urea cycle disorder

necessitating intensive care unit admission. *Can J Anesth Can d'anesthésie 2020 678* 2020;**67**:1094–6. doi:10.1007/S12630-020-01618-3

- 25 Koya Y, Shibata M, Senju M, *et al.* Hyperammonemia in a woman with late-onset ornithine transcarbamylase deficiency. *Intern Med* 2019;**58**:937–42. doi:10.2169/internalmedicine.1851-18
- 26 Panza E, Martinelli D, Magini P, *et al.* Hereditary Spastic Paraplegia Is a Common Phenotypic Finding in ARG1 Deficiency, P5CS Deficiency and HHH Syndrome: Three Inborn Errors of Metabolism Caused by Alteration of an Interconnected Pathway of Glutamate and Urea Cycle Metabolism. *Front Neurol* 2019;**10**:131. doi:10.3389/FNEUR.2019.00131
- 27 Bigot A, Brunault P, Lavigne C, *et al.* Psychiatric adult-onset of urea cycle disorders : A case-series. *Mol Genet Metab Reports* 2017;**12**:103–9. doi:10.1016/j.ymgmr.2017.07.001
- 28 Maillot F, Blasco H, Lioger B, *et al.* Diagnostic et traitement des déficits du cycle de l'urée à l'âge adulte. *La Rev Médecine Interne* 2016;**37**:680–4. doi:10.1016/J.REVMED.2016.02.011
- 29 Anstey JR, Haydon TP, Ghanpur RB, *et al.* Initial presentation of a urea cycle disorder in adulthood: an under-recognised cause of severe neurological dysfunction. *Med J Aust* 2015;**203**:445–7. doi:10.5694/MJA15.00510
- 30 Atiq M, Holt AF, Safdar K, *et al.* Adult onset urea cycle disorder in a patient with presumed hepatic encephalopathy. *J Clin Gastroenterol* 2008;**42**:213–4. doi:10.1097/01.mcg.0000225628.84168.25
- 31 Maillot F, Crenn P. Maladies métaboliques Les déficits du cycle de l ' urée chez les patients adultes. 2007;:897–903.
- 32 Tuchman M, Ahrens M, Barsotti R, *et al.* Consensus statement from a conference for the management of patients with urea cycle disorders. *J Pediatr* 2001;**138**:S1–5. doi:10.1067/mpd.2001.111830
- 33 Marelli C, Lavigne C, Stepien KM, *et al.* Clinical and molecular characterization of adult patients with late-onset MTHFR deficiency. *J Inherit Metab Dis* 2021;**44**:777– 86. doi:10.1002/JIMD.12323
- 34 Vieira D, Florindo C, Almeida IT de, *et al.* Adult-onset methylenetetrahydrofolate reductase deficiency. *BMJ Case Reports CP* 2020;**13**:e232241. doi:10.1136/BCR-2019-232241
- 35 Powers JM, Rosenblatt DS, Schmidt RE, *et al.* Neurological and neuropathologic heterogeneity in two brothers with cobalamin C deficiency. *Ann Neurol* 2001;**49**:396–400. doi:10.1002/ana.78
- 36 Wei Y, Guan Y, Hao H. Late-onset cobalamin C disease presenting with acute progressive polyneuropathy. *Muscle Nerve* 2020;**61**:E37–40. doi:10.1002/MUS.26865
- 37 Hoss GRW, Poloni S, Blom HJ, *et al.* Three Main Causes of Homocystinuria: CBS, cblC and MTHFR Deficiency. What do they Have in Common? *J Inborn Errors Metab Screen* 2019;**7**. doi:10.1590/2326-4594-jiems-2019-0007
- 38 Gales A, Masingue M, Millecamps S, *et al.* Adolescence/adult onset MTHFR deficiency may manifest as isolated and treatable distinct neuro-psychiatric

syndromes. *Orphanet J Rare Dis* 2018;**13**:1–8. doi:10.1186/s13023-018-0767-9

- 39 Morris AAM, Kožich V, Santra S, *et al.* Guidelines for the diagnosis and management of cystathionine beta-synthase deficiency. J. Inherit. Metab. Dis. 2017;**40**:49–74. doi:10.1007/s10545-016-9979-0
- 40 Norris SA, Pogarcic A, Hicks M, *et al.* Adult-onset dystonia with marfanoid features. *Neurol Clin Pract* 2017;**7**:e31–4. doi:10.1212/CPJ.0000000000000297
- 41 Stabler SP, Korson M, Jethva R, *et al.* Metabolic profiling of total homocysteine and related compounds in hyperhomocysteinemia: Utility and limitations in diagnosing the cause of puzzling thrombophilia in a family. In: *JIMD Reports*. Springer 2013. 149–63. doi:10.1007/8904_2013_235
- 42 Michot JM, Sedel F, Giraudier S, *et al.* Psychosis, paraplegia and coma revealing methylenetetrahydrofolate reductase deficiency in a 56-year-old woman. J. Neurol. Neurosurg. Psychiatry. 2008;**79**:963–4. doi:10.1136/jnnp.2008.143677
- 43 Kelly PJ, Furie KL, Kistler JP, *et al.* Stroke in young patients with hyperhomocysteinemia due to cystathionine beta-synthase deficiency. *Neurology* 2003;**60**:275–9. doi:10.1212/01.WNL.0000042479.55406.B3
- 44 Sitarska D, Ługowska A. Laboratory diagnosis of the Niemann-Pick type C disease: an inherited neurodegenerative disorder of cholesterol metabolism. Metab. Brain Dis. 2019;**34**:1253–60. doi:10.1007/s11011-019-00445-w
- 45 Nadjar Y, Hütter-Moncada AL, Latour P, *et al.* Adult Niemann-Pick disease type C in France: Clinical phenotypes and long-term miglustat treatment effect 11 Medical and Health Sciences 1103 Clinical Sciences. *Orphanet J Rare Dis* 2018;**13**. doi:10.1186/s13023-018-0913-4
- 46 Di Lazzaro V, Marano M, Florio L, *et al.* Niemann–Pick type C: focus on the adolescent/adult onset form. Int. J. Neurosci. 2016;**126**:963–71. doi:10.3109/00207454.2016.1161623
- 47 Maubert A, Hanon C, Metton JP. Maladie de Niemann-Pick de type C chez l'adulte et troubles psychiatriques : revue de littérature. *Encephale* 2013;**39**:315–9. doi:10.1016/J.ENCEP.2013.04.013
- 48 Leurs A, Chepy A, Detonellaere C, *et al.* Maladie de Gaucher de type 3, une maladie également de l'adulte ? *La Rev Médecine Interne* 2018;**39**:589–93. doi:10.1016/J.REVMED.2018.03.005
- 49 El-Beshlawy A, Tylki-Szymanska A, Vellodi A, *et al.* Long-term hematological, visceral, and growth outcomes in children with Gaucher disease type 3 treated with imiglucerase in the International Collaborative Gaucher Group Gaucher Registry. *Mol Genet Metab* 2017;**120**:47–56. doi:10.1016/J.YMGME.2016.12.001
- 50 Stirnemann JÔ, Belmatoug N, Camou F, *et al.* A review of gaucher disease pathophysiology, clinical presentation and treatments. *Int J Mol Sci* 2017;**18**:1–30. doi:10.3390/ijms18020441
- 51 Mistry PK, Belmatoug N, Dahl S vom, *et al.* Understanding the natural history of Gaucher disease. *Am J Hematol* 2015;**90**:S6–11. doi:10.1002/AJH.24055
- 52 Grabowski GA, Zimran A, Ida H. Gaucher disease types 1 and 3: Phenotypic characterization of large populations from the ICGG Gaucher Registry. *Am J Hematol* 2015;**90**:S12–8. doi:10.1002/AJH.24063
- 53 Ben Rhouma F, Kallel F, Kefi R, *et al.* Adult gaucher disease in southern Tunisia: report of three cases. *Diagnostic Pathol 2012 71* 2012;**7**:1–4. doi:10.1186/1746- 1596-7-4
- 54 Vellodi A, Tylki-Szymanska A, Davies EH, *et al.* Management of neuronopathic Gaucher disease: Revised recommendations. *J Inherit Metab Dis* 2009;**32**:660–4. doi:10.1007/S10545-009-1164-2
- 55 Guimarães J, Amaral O, Miranda MCS. Adult-onset neuronopathic form of Gaucher's disease: a case report. *Parkinsonism Relat Disord* 2003;**9**:261–4. doi:10.1016/S1353-8020(02)00096-2
- 56 Charrow J, Esplin JA, Gribble TJ, *et al.* Gaucher disease: Recommendations on diagnosis, evaluation, and monitoring. Arch. Intern. Med. 1998;**158**:1754–60. doi:10.1001/archinte.158.16.1754
- 57 Neil JF, Glew RH, Peters SP. Familial Psychosis and Diverse Neurologic Abnormalities in Adult-Onset Gaucher's Disease. *Arch Neurol* 1979;**36**:95–9. doi:10.1001/ARCHNEUR.1979.00500380065007
- 58 Xia Z, Wenwen Y, Xianfeng Y, *et al.* Adult-onset Krabbe disease due to a homozygous GALC mutation without abnormal signals on an MRI in a consanguineous family: A case report. *Mol Genet Genomic Med* 2020;**8**:1–7. doi:10.1002/mgg3.1407
- 59 Cousyn L, Law-Ye B, Pyatigorskaya N, *et al.* Brain MRI features and scoring of leukodystrophy in adult-onset Krabbe disease. *Neurology* 2019;**93**:E647–52. doi:10.1212/WNL.0000000000007943
- 60 Zhang T, Yan C, Ji K, *et al.* Adult-onset Krabbe disease in two generations of a Chinese family. *Ann Transl Med* 2018;**6**:174–174. doi:10.21037/atm.2018.04.30
- 61 Escolar ML, Kiely BT, Shawgo E, *et al.* Psychosine, a marker of Krabbe phenotype and treatment effect. *Mol Genet Metab* 2017;**121**:271–8. doi:10.1016/j.ymgme.2017.05.015
- 62 Liao H-C, Spacil Z, Ghomashchi F, *et al.* Lymphocyte Galactocerebrosidase Activity by LC-MS/MS for Post–Newborn Screening Evaluation of Krabbe Disease. *Clin Chem* 2017;**63**:1363–9. doi:10.1373/clinchem.2016.264952
- 63 Adachi H, Ishihara K, Tachibana H, *et al.* Adult-onset Krabbe disease presenting with an isolated form of peripheral neuropathy. *Muscle Nerve* 2016;**54**:152–7. doi:10.1002/MUS.25067
- 64 Debs R, Froissart R, Aubourg P, *et al.* Krabbe disease in adults: phenotypic and genotypic update from a series of 11 cases and a review. *J Inherit Metab Dis* 2013;**36**:859–68. doi:10.1007/S10545-012-9560-4
- 65 Rappard DF Van, Boelens JJ, Pediatric C, *et al.* Best Practice & Research Clinical Endocrinology & Metabolism Metachromatic leukodystrophy : Disease spectrum and approaches for treatment. *Best Pract Res Clin Endocrinol Metab* 2014;**31**. doi:10.1016/j.beem.2014.10.001
- 66 Hahn AF, Gordon BA, Feleki V, *et al.* A variant form of metachromatic leukodystrophy without arylsulfatase deficiency. *Ann Neurol* 1982;**12**:33–6. doi:10.1002/ana.410120106
- 67 Ortiz A, Germain DP, Desnick RJ, *et al.* Fabry disease revisited: Management and

treatment recommendations for adult patients. *Mol Genet Metab* 2018;**123**:416– 27. doi:10.1016/j.ymgme.2018.02.014

- 68 Curiati MA, Aranda CS, Kyosen SO, *et al.* The Challenge of Diagnosis and Indication for Treatment in Fabry Disease. *J Inborn Errors Metab Screen* 2017;**5**:232640981668573. doi:10.1177/2326409816685735
- 69 Smid BE, Van der Tol L, Biegstraaten M, *et al.* Plasma globotriaosylsphingosine in relation to phenotypes of fabry disease. *J Med Genet* 2015;**52**:262–8. doi:10.1136/jmedgenet-2014-102872
- 70 Ghali J, Murugasu A, Day T, *et al.* Carpal Tunnel Syndrome in Fabry Disease. *JIMD Rep* 2011;**2**:17–23. doi:10.1007/8904_2011_37
- 71 Hegemann S, Hajioff D, Conti G, *et al.* Hearing loss in Fabry disease: data from the Fabry Outcome Survey. *Eur J Clin Invest* 2006;**36**:654–62. doi:10.1111/J.1365- 2362.2006.01702.X
- 72 Hossain A. ARTICLE IN PRESS A Case of Adult-onset Pompe Disease with Cerebral Stroke and Left Ventricular Hypertrophy D5X X Keiko. *J Stroke Cerebrovasc Dis* 2018;:1–7. doi:10.1016/j.jstrokecerebrovasdis.2018.06.043
- 73 Chan J, Desai AK, Kazi ZB, *et al.* The emerging phenotype of late-onset Pompe disease: A systematic literature review. Mol. Genet. Metab. 2017;**120**:163–72. doi:10.1016/j.ymgme.2016.12.004
- 74 Young SP, Stevens RD, An Y, *et al.* Analysis of a glucose tetrasaccharide elevated in Pompe disease by stable isotope dilution-electrospray ionization tandem mass spectrometry. *Anal Biochem* 2003;**316**:175–80. doi:10.1016/S0003-2697(03)00056- 3
- 75 Umapathysivam K, Whittle AM, Ranieri E, *et al.* Determination of acid α-glucosidase protein: Evaluation as a screening marker for Pompe disease and other lysosomal storage disorders. *Clin Chem* 2000;**46**:1318–25.
- 76 Rattay TW, Rautenberg M, Söhn AS, *et al.* Defining diagnostic cutoffs in neurological patients for serum very long chain fatty acids (VLCFA) in genetically confirmed X-Adrenoleukodystrophy. *Sci Rep* 2020;**10**:1–12. doi:10.1038/s41598-020-71248-8
- 77 Mannari A, Wiggins B, Bachuwa G. Adult-Onset Cerebral Adrenoleukodystrophy Without Adrenal Gland Involvement. *Cureus* 2020;**12**. doi:10.7759/CUREUS.9813
- 78 Shamim D, Alleyne K. X-linked adult-onset adrenoleukodystrophy: Psychiatric and neurological manifestations. *SAGE Open Med Case Reports* 2017;**5**:2050313X1774100. doi:10.1177/2050313x17741009
- 79 Stepien KM, Wierzbicki AS, Poll-The BT, *et al.* The Challenges of a Successful Pregnancy in a Patient with Adult Refsum's Disease due to Phytanoyl-CoA Hydroxylase Deficiency. *JIMD Rep* 2016;**33**:49–53. doi:10.1007/8904_2016_569
- 80 Bompaire F, Marcaud V, Trionnaire E Le, *et al.* Refsum Disease Presenting with a Late-Onset Leukodystrophy. *JIMD Rep* 2014;**19**:7–10. doi:10.1007/8904_2014_355
- 81 Wanders RJA, Komen J, Ferdinandusse S. Phytanic acid metabolism in health and disease. Biochim. Biophys. Acta - Mol. Cell Biol. Lipids. 2011;**1811**:498–507. doi:10.1016/j.bbalip.2011.06.006
- 82 Britton TC, Gibberd FB, Clemens ME, *et al.* The significance of plasma phytanic acid levels in adults. *J Neurol Neurosurg Psychiatry* 1989;**52**:891–4.

doi:10.1136/jnnp.52.7.891

- 83 Smith EH, Gavrilov DK, Oglesbee D, *et al.* An adult onset case of alpha-methyl-acyl-CoA racemase deficiency. *J Inherit Metab Dis* 2010;**33**:349–53. doi:10.1007/S10545- 010-9183-6
- 84 Kapina V, Sedel F, Truffert A, *et al.* RELAPSING RHABDOMYOLYSIS DUE TO PEROXISOMAL α-METHYLACYL-COA RACEMASE DEFICIENCY. *Neurology* 2010;**75**:1300–2. doi:10.1212/WNL.0B013E3181F612A5
- 85 Clarke CE, Alger S, Preece MA, *et al.* Tremor and deep white matter changes in αmethylacyl-CoA racemase deficiency. *Neurology* 2004;**63**:188–9. doi:10.1212/01.WNL.0000132841.81250.B7
- 86 McLean BN, Allen J, Ferdinandusse S, *et al.* A new defect of peroxisomal function involving pristanic acid: A case report. *J Neurol Neurosurg Psychiatry* 2002;**72**:396– 9. doi:10.1136/jnnp.72.3.396
- 87 Ferdinandusse S, Denis S, Clayton PT, *et al.* Mutations in the gene encoding peroxisomal α-methylacyl-CoA racemase cause adult-onset sensory motor neuropathy. *Nat Genet 2000 242* 2000;**24**:188–91. doi:10.1038/72861
- 88 Osman C, Foulds N, Hunt D, *et al.* Diagnosis of pyridoxine-dependent epilepsy in an adult presenting with recurrent status epilepticus. *Epilepsia* 2020;**61**:e1–6. doi:10.1111/EPI.16408
- 89 Xue J, Wang J, Gong P, *et al.* Simultaneous quantification of alpha-aminoadipic semialdehyde, piperideine-6-carboxylate, pipecolic acid and alpha-aminoadipic acid in pyridoxine-dependent epilepsy. *Sci Rep* 2019;**9**:1–10. doi:10.1038/s41598-019- 47882-2
- 90 Dahiphale R, Jain S, Agrawal M. Biotinidase deficiency. *Indian Pediatr* 2008;**45**:777– 9. doi:10.1212/nxg.0000000000000525
- 91 Van Winckel G, Ballhausen D, Wolf B, *et al.* Severe Distal Motor Involvement in a Non-compliant Adult With Biotinidase Deficiency: The Necessity of Life-Long Biotin Therapy. *Front Neurol* 2020;**11**:1–5. doi:10.3389/fneur.2020.516799
- 92 Iseghem V Van, Sprengers M, Zaeytijd J De, *et al.* Biotinidase de fi ciency : A treatable cause of opticospinal syndrome in young. *Mult Scler Relat Disord* 2019;**32**:64–5. doi:10.1016/j.msard.2019.04.025
- 93 Wolf B. Biotinidase deficiency should be considered in individuals thought to have multiple sclerosis and related disorders. *Mult Scler Relat Disord* 2019;**28**:26–30. doi:10.1016/j.msard.2018.11.030
- 94 Deschamps R, Savatovsky J, Vignal C, *et al.* Adult-onset biotinidase deficiency: Two individuals with severe, but reversible optic neuropathy. J. Neurol. Neurosurg. Psychiatry. 2018;**89**:1009–10. doi:10.1136/jnnp-2017-316644
- 95 Yilmaz S, Serin M, Canda E, *et al.* A treatable cause of myelopathy and vision loss mimicking neuromyelitis optica spectrum disorder: late-onset biotinidase deficiency. *Metab Brain Dis* 2017;**32**:675–8. doi:10.1007/s11011-017-9984-5
- 96 Bottin L, Prud'hon S, Guey S, *et al.* Biotinidase deficiency mimicking neuromyelitis optica: Initially exhibiting symptoms in adulthood. *Mult Scler* 2015;**21**:1604–7. doi:10.1177/1352458515596457
- 97 Cowan TM, Blitzer MG, Wolf B. Technical standards and guidelines for the diagnosis

of biotinidase deficiency. Genet. Med. 2010;**12**:464–70. doi:10.1097/GIM.0b013e3181e4cc0f

- 98 Lee J, Hegele RA. Abetalipoproteinemia and homozygous hypobetalipoproteinemia: a framework for diagnosis and management. *J Inherit Metab Dis* 2014;**37**:333–9. doi:10.1007/s10545-013-9665-4
- 99 Nagappa M, Bindu PS, Adwani S, *et al.* Clinical, hematological, and imaging observations in a 25-year-old woman with abetalipoproteinemia. *Ann Indian Acad Neurol* 2014;**17**:113. doi:10.4103/0972-2327.128574
- 100 Zamel R, Khan R, Pollex RL, *et al.* Abetalipoproteinemia: Two case reports and literature review. *Orphanet J Rare Dis* 2008;**3**:1–8. doi:10.1186/1750-1172-3-19
- 101 Euch-Fayache G El, Bouhlal Y, Amouri R, *et al.* Molecular, clinical and peripheral neuropathy study of Tunisian patients with ataxia with vitamin E deficiency. *Brain* 2014;**137**:402–10. doi:10.1093/brain/awt339
- 102 Cavalier L, Ouahchi K, Kayden HJ, *et al.* Ataxia with isolated vitamin E deficiency: Heterogeneity of mutations and phenotypic variability in a large number of families. *Am J Hum Genet* 1998;**62**:301–10. doi:10.1086/301699
- 103 Finckh B, Kontush A, Commentz J, *et al.* Monitoring of ubiquinol-10, ubiquinone-10, carotenoids, and tocopherols in neonatal plasma microsamples using highperformance liquid chromatography with coulometric electrochemical detection. *Anal Biochem* 1995;**232**:210–6. doi:10.1006/abio.1995.0009
- 104 Gotoda T, Arita M, Arai H, *et al.* Adult-Onset Spinocerebellar Dysfunction Caused by a Mutation in the Gene for the α-Tocopherol–Transfer Protein. *N Engl J Med* 1995;**333**:1313–9. doi:10.1056/nejm199511163332003
- 105 Pope S, Artuch R, Heales S, *et al.* Cerebral folate deficiency: Analytical tests and differential diagnosis. *J Inherit Metab Dis* 2019;**42**:jimd.12092. doi:10.1002/jimd.12092
- 106 Masingue M, Benoist JF, Roze E, *et al.* Cerebral folate deficiency in adults: A heterogeneous potentially treatable condition. *J Neurol Sci* 2019;**396**:112–8. doi:10.1016/j.jns.2018.11.014
- 107 Sadighi Z, Butler IJ, Koenig MK. Adult-onset cerebral folate deficiency. *Arch Neurol* 2012;**69**:778–9. doi:10.1001/archneurol.2011.3036
- 108 Tabarki B, Al-Shafi S, Al-Shahwan S, *et al.* Biotin-responsive basal ganglia disease revisited: Clinical, radiologic, and genetic findings. Neurology. 2013;**80**:261–7. doi:10.1212/WNL.0b013e31827deb4c
- 109 Kono S, Miyajima H, Yoshida K, *et al.* Mutations in a Thiamine-Transporter Gene and Wernicke's-like Encephalopathy. *N Engl J Med* 2009;**360**:1792–4. doi:10.1056/nejmc0809100
- 110 Carreau C, Benoit C, Ahle G, *et al.* Late-onset riboflavin transporter deficiency: A treatable mimic of various motor neuropathy aetiologies. *J Neurol Neurosurg Psychiatry* 2021;**92**:27–35. doi:10.1136/jnnp-2020-323304
- 111 Foley AR, Menezes MP, Pandraud A, *et al.* Treatable childhood neuronopathy caused by mutations in riboflavin transporter RFVT2. *Brain* 2014;**137**:44–56. doi:10.1093/brain/awt315
- 112 Leen WG, Wevers RA, Kamsteeg EJ, *et al.* Cerebrospinal fluid analysis in the workup

of GLUT1 deficiency syndrome: A systematic review. JAMA Neurol. 2013;**70**:1440–4. doi:10.1001/jamaneurol.2013.3090

- 113 Afawi Z, Suls A, Ekstein D, *et al.* Mild adolescent/adult onset epilepsy and paroxysmal exercise-induced dyskinesia due to GLUT1 deficiency. *Epilepsia* 2010;**51**:2466–9. doi:10.1111/J.1528-1167.2010.02726.X
- 114 Veggiotti P, Teutonico F, Alfei E, *et al.* Glucose transporter type 1 deficiency: Ketogenic diet in three patients with atypical phenotype. *Brain Dev* 2010;**32**:404–8. doi:10.1016/J.BRAINDEV.2009.04.013
- 115 Pavlu-Pereira H, Silva MJ, Florindo C, *et al.* Pyruvate dehydrogenase complex deficiency: updating the clinical, metabolic and mutational landscapes in a cohort of Portuguese patients. *Orphanet J Rare Dis* 2020;**15**:298. doi:10.1186/s13023-020- 01586-3
- 116 Rahman S, Clarke CF, Hirano M. 176th ENMC International Workshop: Diagnosis and treatment of coenzyme Q 10 deficiency. *Neuromuscul Disord* 2012;**22**:76–86. doi:10.1016/j.nmd.2011.05.001
- 117 Sedel F, Challe G, Mayer JM, *et al.* Thiamine responsive pyruvate dehydrogenase deficiency in an adult with peripheral neuropathy and optic neuropathy. J. Neurol. Neurosurg. Psychiatry. 2008;**79**:846–7. doi:10.1136/jnnp.2007.136630
- 118 Quinzii CM, Hirano M, DiMauro S. CoQ10 deficiency diseases in adults. *Mitochondrion* 2007;**7**:122–6. doi:10.1016/j.mito.2007.03.004
- 119 Mellick G, Price L, Boyle R. Late-onset presentation of pyruvate dehydrogenase deficiency. *Mov Disord* 2004;**19**:727–9. doi:10.1002/MDS.20063
- 120 Ogasahara S, Engel AG, Frens D, *et al.* Muscle coenzyme Q deficiency in familial mitochondrial encephalomyopathy. *Proc Natl Acad Sci* 1989;**86**:2379–82. doi:10.1073/PNAS.86.7.2379
- 121 Ciron J, Baron C, Boissonnot M, *et al.* Peripheral nervous system involvement in Leber's hereditary optic neuropathy. *J Neurol Sci* 2018;**388**:94–6. doi:10.1016/J.JNS.2018.03.002
- 122 Carelli V, Carbonelli M, De Coo IF, *et al.* International consensus statement on the clinical and therapeutic management of leber hereditary optic neuropathy. *J Neuro-Ophthalmology* 2017;**37**:371–81. doi:10.1097/WNO.0000000000000570
- 123 Martikainen MH, Ng YS, Gorman GS, *et al.* Clinical, Genetic, and Radiological Features of Extrapyramidal Movement Disorders in Mitochondrial Disease. *JAMA Neurol* 2016;**73**:668–74. doi:10.1001/JAMANEUROL.2016.0355
- 124 Pfeffer G, Burke A, Yu-Wai-Man P, *et al.* Clinical features of MS associated with Leber hereditary optic neuropathy mtDNA mutations. *Neurology* 2013;**81**:2073–81. doi:10.1212/01.WNL.0000437308.22603.43
- 125 J P. Multiple sclerosis associated with Leber's Hereditary Optic Neuropathy. *J Neurol Sci* 2009;**286**:24–7. doi:10.1016/J.JNS.2009.09.009
- 126 McFarland R, Chinnery PF, Blakely EL, *et al.* Homoplasmy, heteroplasmy, and mitochondrial dystonia. *Neurology* 2007;**69**:911–6. doi:10.1212/01.WNL.0000267843.10977.4A
- 127 Gilhuis HJ, Schelhaas HJ, Cruysberg JRM, *et al.* Demyelinating polyneuropathy in Leber hereditary optic neuropathy. *Neuromuscul Disord* 2006;**16**:394–5.

doi:10.1016/J.NMD.2006.03.006

- 128 Horváth R, Abicht A, Shoubridge EA, *et al.* Leber's hereditary optic neuropathy presenting as multiple sclerosis-like disease of the CNS. *J Neurol 2000 2471* 2000;**247**:65–7. doi:10.1007/S004150050015
- 129 Nikoskelainen EK, Marttila RJ, Huoponen K, *et al.* Leber's 'plus': neurological abnormalities in patients with Leber's hereditary optic neuropathy. *J Neurol Neurosurg Psychiatry* 1995;**59**:160–4. doi:10.1136/JNNP.59.2.160
- 130 Nadjar Y, Souvannanorath S, Maisonobe T, *et al.* Sensory neuronopathy as a major clinical feature of mitochondrial trifunctional protein deficiency in adults. *Rev Neurol (Paris)* 2020;**176**:380–6. doi:10.1016/j.neurol.2019.11.011
- 131 Chen W, Zhang Y, Ni Y, *et al.* Late-onset riboflavin-responsive multiple acyl-CoA dehydrogenase deficiency (MADD): Case reports and epidemiology of ETFDH gene mutations. *BMC Neurol* 2019;**19**:1–7. doi:10.1186/s12883-019-1562-5
- 132 Van Hove JLK, Zhang W, Kahler SG, *et al.* Medium-chain acyl-CoA dehydrogenase (MCAD) deficiency: Diagnosis by acylcarnitine analysis in blood. *Am J Hum Genet* 1993;**52**:958–66./pmc/articles/PMC1682046/?report=abstract (accessed 11 Mar 2021).
- 133 De Biase I, Viau KS, Liu A, *et al.* Diagnosis, treatment, and clinical outcome of patients with mitochondrial trifunctional protein/long-chain 3-hydroxy acyl-CoA dehydrogenase deficiency. In: *JIMD Reports*. Springer 2017. 63–71. doi:10.1007/8904_2016_558
- 134 Wang Z, Hong D, Zhang W, *et al.* Severe sensory neuropathy in patients with adultonset multiple acyl-CoA dehydrogenase deficiency. *Neuromuscul Disord* 2016;**26**:170–5. doi:10.1016/J.NMD.2015.12.002
- 135 Grünert SC. Clinical and genetical heterogeneity of late-onset multiple acylcoenzyme A dehydrogenase deficiency. *Orphanet J Rare Dis* 2014;**9**:1–8. doi:10.1186/s13023-014-0117-5
- 136 Rosenbohm A, Süssmuth SD, Kassubek J, *et al.* Novel ETFDH mutation and imaging findings in an adult with glutaric aciduria type II. *Muscle Nerve* 2014;**49**:446–50. doi:10.1002/MUS.23979
- 137 Liewluck T, Mundi MS, Mauermann ML. Mitochondrial trifunctional protein deficiency: a rare cause of adult-onset rhabdomyolysis. *Muscle Nerve* 2013;**48**:989– 91. doi:10.1002/mus.23959
- 138 Schatz UA, Ensenauer R. The clinical manifestation of MCAD deficiency: Challenges towards adulthood in the screened population. J. Inherit. Metab. Dis. 2010;**33**:513– 20. doi:10.1007/s10545-010-9115-5
- 139 Lang TF. Adult presentations of medium-chain acyl-CoA dehydrogenase deficiency (MCADD). J. Inherit. Metab. Dis. 2009;**32**:675–83. doi:10.1007/s10545-009-1202-0
- 140 Abdenur JE, Chamoles NA, Schenone AB, *et al.* Multiple acyl-CoA-dehydrogenase deficiency (MADD): Use of acylcarnitines and fatty acids to monitor the response to dietary treatment. *Pediatr Res* 2001;**50**:61–6. doi:10.1203/00006450-200107000- 00013
- 141 Li LY, Zhu XQ, Tao WW, *et al.* Acute onset neurological symptoms in Wilson disease after traumatic, surgical or emotional events: A cross-sectional study. *Med (United*

States) 2019;**98**. doi:10.1097/MD.0000000000015917

- 142 Guillaud O, Brunet A-S, Mallet I, *et al.* Relative exchangeable copper: A valuable tool for the diagnosis of Wilson disease. *Liver Int* 2018;**38**:350–7. doi:10.1111/liv.13520
- 143 Bandmann O, Weiss KH, Kaler SG. Wilson's disease and other neurological copper disorders. *Lancet Neurol* 2015;**14**:103–13. doi:10.1016/S1474-4422(14)70190-5
- 144 Roberts EA, Schilsky ML. Diagnosis and treatment of Wilson disease: An update. Hepatology. 2008;**47**:2089–111. doi:10.1002/hep.22261
- 145 Ferenci P, Członkowska A, Merle U, *et al.* Late-Onset Wilson's Disease. *Gastroenterology* 2007;**132**:1294–8. doi:10.1053/J.GASTRO.2007.02.057
- 146 Marchi G, Busti F, Zidanes AL, *et al.* Aceruloplasminemia: A Severe Neurodegenerative Disorder Deserving an Early Diagnosis. *Front Neurosci* 2019;**13**. doi:10.3389/FNINS.2019.00325
- 147 Kono S. Aceruloplasminemia. *Curr Drug Targets* 2012;**13**:1190–9. doi:10.2174/138945012802002320
- 148 Quadri M, Federico A, Zhao T, *et al.* Mutations in SLC30A10 cause parkinsonism and dystonia with hypermanganesemia, polycythemia, and chronic liver disease. *Am J Hum Genet* 2012;**90**:467–77. doi:10.1016/j.ajhg.2012.01.017
- 149 Tuschl K, Clayton PT, Gospe SM, *et al.* Syndrome of hepatic cirrhosis, dystonia, polycythemia, and hypermanganesemia caused by mutations in SLC30A10, a manganese transporter in man. *Am J Hum Genet* 2012;**90**:457–66. doi:10.1016/j.ajhg.2012.01.018
- 150 Gospe J, Caruso RD, Clegg MS, *et al.* Paraparesis, hypermanganesaemia, and polycythaemia: A novel presentation of cirrhosis. *Arch Dis Child* 2000;**83**:439–42. doi:10.1136/adc.83.5.439
- 151 Makary MS, Kisanuki YY, Amin NN, *et al.* Teaching NeuroImages: Cerebrotendinous xanthomatosis. Neurology. 2018;**90**:e637–8. doi:10.1212/WNL.0000000000004967
- 152 Sasamura A, Akazawa S, Haraguchi A, *et al.* Late-onset cerebrotendinous xanthomatosis with a novel mutation in the CYP27A1 gene. *Intern Med* 2018;**57**:1611–6. doi:10.2169/internalmedicine.0120-17
- 153 Degos B, Nadjar Y, Amador M, *et al.* Natural history of cerebrotendinous xanthomatosis : a paediatric disease diagnosed in adulthood. *Orphanet J Rare Dis* 2016;:1–4. doi:10.1186/s13023-016-0419-x
- 154 Nakashima N, Sakai Y, Sakai H, *et al.* A point mutation in the bile acid biosynthetic enzyme sterol 27- hydroxylase in a family with cerebrotendinous xanthomatosis. *J Lipid Res* 1994;**35**:663–8. doi:10.1016/s0022-2275(20)41180-0
- 155 Leitersdorf E, Reshef A, Meiner V, *et al.* Frameshift and splice-junction mutations in the sterol 27-hydroxylase gene cause cerebrotendinous xanthomatosis in Jews of Moroccan origin. *J Clin Invest* 1993;**91**:2488–96. doi:10.1172/JCI116484
- 156 Simon A, Pompilus F, Querbes W, *et al.* Patient Perspective on Acute Intermittent Porphyria with Frequent Attacks : A Disease with Intermittent and Chronic Manifestations. *Patient - Patient-Centered Outcomes Res* Published Online First: 2018. doi:10.1007/s40271-018-0319-3
- 157 Kevelam SH, Neeleman RA, Waisfisz Q, *et al.* Acute intermittent porphyria-related leukoencephalopathy. *Neurology* 2016;**87**:1258–65.

doi:10.1212/WNL.0000000000003129

- 158 Bonkovsky HL, Maddukuri VC, Yazici C, *et al.* Acute Porphyrias in the USA: Features of 108 Subjects from Porphyrias Consortium. *Am J Med* 2014;**127**:1233–41. doi:10.1016/J.AMJMED.2014.06.036
- 159 Puy H, Gouya L, Deybach JC. Porphyrias. *Lancet* 2010;**375**:924–37. doi:10.1016/S0140-6736(09)61925-5
- 160 Hooper AJ, Hegele RA, Burnett JR. Tangier disease. *Curr Opin Lipidol* 2020;**31**:80–4. doi:10.1097/MOL.0000000000000669
- 161 Mercan M, Yayla V, Altinay S, *et al.* Peripheral neuropathy in Tangier disease: A literature review and assessment. *J Peripher Nerv Syst* 2018;**23**:88–98. doi:10.1111/jns.12265