

## Spatio-temporal beta diversity of plankton species and their interactions in permanent and temporal waterholes in a semi-arid savannah

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Spatio-temporal beta diversity of plankton species and their interactions in permanent and temporal waterholes in a semi-arid savannah 6 7 8 Florence D. Hulot <sup>a,b\*</sup>, S. Msiteli-Shumba<sup>c</sup>, Mathias Iung <sup>a,e</sup>, Camille Noûs <sup>a,d</sup>, Elisa Thébault <sup>e</sup> <sup>a</sup>Ecologie Systématique Evolution, Univ. Paris-Sud, CNRS, AgroParisTech, Université Paris-Saclay, Orsay, France; <sup>b</sup>Zone Atelier Hwange - Hwange LTER, CNRS-CIRAD, Dete, Zimbabwe; <sup>c</sup>University of Zimbabwe, Biological Sciences Department, Harare, Zimbabwe; <sup>d</sup>https://www.cogitamus.fr/; <sup>e</sup>Sorbonne Université, CNRS, IRD, INRAE, Université Paris Est Créteil, Institute of Ecology and Environmental Sciences of Paris (iEES-Paris), Paris, France \*Corresponding author: Florence D. Hulot (florence.hulot@universite-paris-saclay.fr) 

## **Abstract**

Freshwater communities can vary greatly across space and time. Studying these variations,
that is spatial and temporal beta diversity, provides fundamental information on the processes
that maintain diversity and on the consequences of environmental changes on communities.
Recently, drying events have been shown to strongly affect the spatial and temporal beta
diversity of temperate freshwater ecosystems, but the effects of such events are mostly
unknown for freshwater communities in semi-arid climates that are frequently submitted to
drying up. In addition, studies have so far focused on variations in species composition that is
species beta diversity, while variations in species trophic interactions that is food-web beta
diversity can give additional insights on how community functioning vary in space and time.
Here, we combine species and food-web perspectives to explore the spatiotemporal beta
diversity of plankton species and their trophic interactions in waterholes undergoing different
water-level regimes: either an alternation between dry and water-full states, or a permanent
water-full state due to water pumping. Our results show that waterholes with artificial water
pumping do not differ from natural waterholes in their contribution to spatiotemporal beta
diversity. Instead, beta diversity is strongly driven by temporal variations of species
composition and food-web structure during the dry season, which is characterized by
degraded planktonic communities with a low richness, diversity and connectance. Species-
and interaction-based approaches give complementary information on the spatiotemporal beta
diversity, as they highlight different planktonic communities with contrasted functioning.

#### Keywords

Food-web beta diversity, food web connectance, trophic level, Hwange National Park, planktonic food web, temporary ponds

#### 1 Introduction

 The study of the variations of species composition across space and time (i.e. beta diversity) is of major importance to explain diversity patterns on earth and to provide fundamental information on the processes that maintain this diversity (Whittaker 1972). Planktonic communities and their dynamics have been widely studied in isolated freshwater ecosystems. However these ecosystems are connected by biological processes such as individual active and passive dispersal (Kappes et al. 2014; Incagnone et al. 2015; Juracka et al. 2016; Slusarczyk et al. 2017), and as for other types of ecosystems, their diversity depends both on stochastic processes related with colonization and extinction dynamics, and on deterministic processes associated with differences in species niches to environmental conditions (Chase 2007). While the variations of communities across space (i.e. spatial beta diversity) have been generally studied separately from the variations across time (i.e. temporal beta diversity), recent studies have emphasized the importance of considering spatial and temporal dimensions at the same time (Cook et al. 2018; Musters et al. 2019; Crabot et al. 2020; Sarremejane et al. 2020). Indeed, the spatial and temporal beta diversity of freshwater ecosystems are together affected by environmental perturbations such as drying events or eutrophication. For instance, Sarremejane et al. (2020) showed that the temporal beta diversity of aquatic invertebrates in streams in UK increased with longer drought events while the spatial beta diversity decreased as flow permanence increased. So far, the study of spatiotemporal variations in freshwater communities has been mainly restricted to a few temperate stream ecosystems. Aquatic ecosystems in semi-arid areas display a wide range of dynamics from temporary to permanent systems, which may vary from year to year. The alternation between completely dry state and water-full state is a strong selective pressure on aquatic species, which have consequences on aquatic systems functioning (Thomas et al. 2000; Coops et al. 2003; Thomaz et al. 2006; Bazzanti et al. 2009;

 Zohary and Ostrovsky 2011; Michaloudi et al. 2012; Teferi et al. 2014; Medeiros et al. 2015). Such alternation between dry and water-full states is expected to strongly affect both spatial and temporal beta diversity. Repeated drying events might decrease spatial beta diversity as a result of increasing environmental filtering in harsh conditions (Chase 2007) but it could also increase spatial beta diversity if it leads to greater habitat heterogeneity (Sarremejane et al. 2020). Meanwhile, strong seasonal changes in water level may lead to very dissimilar species assemblages at different times of the year, and thus to high temporal beta diversity, as is expected in highly seasonal and predictable environments (Tonkin et al. 2017). Our understanding of the diversity of planktonic communities in semi-arid aquatic ecosystems should thus gain from a spatiotemporal perspective of beta diversity. The study of beta diversity has been classically considered from a species-based taxonomical point of view, but other aspects of biodiversity such as functional trait diversity or diversity of species interactions in ecological networks can offer complementary perspectives on this issue (Pellissier et al. 2018; Crabot et al. 2019; Ohlmann et al. 2019). As for species diversity, species interaction diversity may be explored at the local level (which species interactions occur in a given place at a given time) but also at the spatial regional level (which species interactions are common between communities or specific to some communities) and at the temporal level (which species interactions persist or vanish in time) (Poisot et al. 2012; Pellissier et al. 2018). Because the changes in abundance and occurrence of species across space and time can depend on species traits and/or trophic groups, the drivers of beta diversity can differ depending on the diversity metrics considered (Pellissier et al. 2018; Crabot et al. 2019). By building on recent developments on the study of the beta diversity of ecological networks, linking species-based and network-based approaches should allow to better characterize how the functioning of planktonic food webs in freshwater ecosystems vary across time and space. Likewise, species interaction beta diversity assessed in a set of aquatic

ecosystems may help to identify the interactions that have a pivotal role and to assess the impacts of drying up on ecosystem functioning. We study here the spatiotemporal beta diversity of plankton species and plankton interactions in waterholes located in a semi-arid area in Hwange National Park and its periphery, Zimbabwe. Thirty waterholes, located in the park and the neighboring forestry and communal areas, were monitored during a period of 10 months covering the rainy and dry seasons. While some of these waterholes were temporal as they dried up during the dry season, other waterholes remained permanent thanks to their large water stock or to artificial water pumping from the water table (Msiteli-Shumba et al. 2018). Artificially pumped waterholes (hereafter artificial waterholes) were developed as a management measure to provide water for wildlife during the dry season (Chamaillé-Jammes, Valeix, et al. 2007). This study system offers an opportunity to investigate the potential consequences of the alternation between dry and water-full states on spatial and temporal beta diversity of freshwater ecosystems in tropical semi-arid areas. More specifically, we address the following questions: (1) do the location and pumping of water affect species and interaction-based beta diversity of waterholes? (2) How do temporal changes in community composition in the waterholes contribute to overall beta diversity? (3) What is the degree of uniqueness of the waterholes in

waterholes and decrease seasonal variations in environmental conditions in these waterholes.

Artificial waterholes are then expected to contribute less than natural and temporal waterholes to overall beta diversity. We also expect that temporal beta diversity plays a significant part of overall beta diversity in this highly seasonal system. Finally, we expect that interaction beta

space and time and how does it relate with the diversity and food web structure of the

waterhole communities? (4) What are the characteristics of plankton species and interactions

that distinguish waterholes? We hypothesize that artificial pumping decreases both spatial and

temporal beta diversity as it might homogenize environmental conditions among artificial

- diversity will give a complementary perspective to species beta diversity. While both are
- 2 expected to be related with species diversity because species poor communities often tend to
- 3 be more dissimilar (Chase and Myers 2011; da Silva Brito et al. 2020), food web complexity
- 4 is also expected to matter for interaction beta diversity.

#### **Methods**

7 Study site

8 Hwange National Park (HNP) covers an area of about 14 651 km² and is situated in

Matabeleleland North Province, Western Zimbabwe. It is located in a semi-arid area where the

rainfall is highly variable with an average of 650 mm and a coefficient of variation of 25%

(Arraut et al. 2018). The vegetation in most of the park, especially in the area where this study

was carried out, is typical of a highly heterogeneous dystrophic wooded savanna, dominated by

mixed woodlands and bushlands on Kalahari sands, with open grasslands along drainage lines

(Chamaillé-Jammes, Fritz, et al. 2007). The rainy season lasts from December to April. The

cool dry winter months extend from May to July and the hot dry months from August to

November.

The waterholes investigated in this study are located in HNP and the adjacent Sikumi Forest

and communal areas situated in the periphery of the park (Figure 1). The HNP is devoted to

wildlife conservation and there are no inhabitants. Domestic and wildlife species coexist in the

forest reserve and in the communal area but the latter is inhabited. Numerous depressions are

fed by rainwater runoff during the rainy season and, according to their size and depth, these

natural waterholes may dry out completely during the course of the dry season. To supply

wildlife with water during the dry season, underground water is pumped to maintain water in

 some permanent artificial waterholes. Natural and artificial waterholes are found in the park and forestry areas, while only natural waterholes are found in the communal area. Rainfalls may be restricted to some areas, especially in the beginning and end of the rainy season, leading to heterogeneity in waterholes filling and drying up.

Field sampling and laboratory analyses

identified to the lowest taxonomic level possible.

Thirty waterholes were sampled between February and December 2013 over 7 months, covering the wet, cool dry and hot dry seasons. The waterholes were selected to represent a set of artificial and natural waterholes, located inside and outside the park. These comprised 5 waterholes in the communal area, 5 waterholes in the Sikumi Forest, and 20 waterholes in the Hwange Main Camp area in HNP. All waterholes could not be sampled at each sampling date because of impossible access during the wet season or because the waterholes had dried up during the dry season. Please note that some artificial waterholes were also sampled occasionally as some of the pumps may have been out of order causing the artificial waterholes to dry up. In short, 28 waterholes were sampled in February and March, 26 in April, 21 in July, 14 in September, 12 in November and December. Samples were collected with the use of a 1L polyethene bottle fastened to a 3 m pole. Water samples were taken between 8:00 and 12:00 to minimize variability in the sampling time of the day. About 10L of water was filtrated first through a 100 µm filter and then through a 30 µm filter. Samples were collected in 50 ml falcon tubes and preserved in 4% formalin. Taxonomic identification was carried out under an Olympus CK40 inverted microscope with the assistance of taxonomic keys (Msiteli-Shumba et al. 2017). All plankton species were

 1 Species beta diversity

2 We quantified species beta diversity following the method of Legendre and Caceres (2013).

3 Because the waterholes were not all sampled at all dates, we could not compare the total beta

4 diversity among waterholes only, or among dates only. Thus the total beta diversity was

computed over all waterholes and sampling dates, and it was expressed as the variance of the

Hellinger transformed species-by-sampling unit matrix. Each sampling unit corresponds here

to a waterhole-month combination. We then calculated the contribution of each sampling unit

to beta diversity (hereafter LCBD<sub>Sp</sub>) following Legendre and Caceres (2013). The LCBD<sub>Sp</sub>

values represent the uniqueness of the sampling units, i.e. the waterhole a given month, in

terms of species composition. LCBD<sub>Sp</sub> values were tested for significance against the null

hypothesis of random species distributions among waterholes and months. We also computed

the Species Contributions to Beta Diversity (hereafter SCBD<sub>Sp</sub>) to identify the species that

contribute the most to species beta diversity. For instance, species with high abundances in

only a few sites have high SCBD<sub>Sp</sub> values. The LCBD<sub>Sp</sub> and SCBD<sub>Sp</sub> indices were computed

using the beta.div function of the adespatial package in R (Dray et al. 2019).

*Interaction beta diversity* 

18 Building the meta-food web

The first step to analyze food-web beta diversity is to describe all the potential trophic

interactions among all the observed species in the waterholes, which constitutes the meta-food

web. To build the corresponding prey-predator interaction matrix, we looked into the

22 literature for data on the predation of the zooplankton species on phytoplankton and

potentially other zooplankton species. Not all species could be found in the literature.

According to personal expertise, allometric relationships between planktonic prey and

 predators (Brooks and Dodson 1965; Burns 1968) and species taxonomy (Gauzens et al. 2013), we hypothesized that species of the same size and belonging to the same genus have the same diet. With this information, we were able to construct the meta-food web matrix where all species observed in the waterholes are in lines and columns. The predation of the species in the line by the species in the column was coded by a "1". The absence of predation was represented by a "0". In the matrix, except for copepod nauplii, species with ontogenetic omnivory appear in the matrix only once in the column with the multiple prey that they eat during their lifetime, even though the species may be prey as a juvenile and a predator as an adult. Therefore, cannibalism is represented by a "1" at the intersection of the same species in the column and line. Copepod nauplii was represented in the matrix as a tropho-species. We added in the matrix the groups "bacteria", "small phytoplankton" (phytoplankton <30 µm), "protists" and "POM and DOM" (particulate and dissolved organic matter) as some species fed on these groups. Small phytoplankton, protists and POM and DOM are considered to be basal species. These groups are essential in the network construction. Otherwise small filterfeeders such as small rotifers and nauplii would be at the same trophic level than primary producers and the diet of generalist filter-feeders such as cladocerans would be narrowed. Once the meta-food web was built, we extracted local food webs for each waterhole at each sampling month resulting in 141 food webs.

Calculating interaction beta diversity

To follow the same approach as detailed above for beta diversity on community composition but on food webs, we adapted the method of Legendre and Caceres (2013) to food web data. First, we calculated food web based LCBD values (hereafter LCBD<sub>FW</sub>) using the interaction-by-sampling unit matrix instead of the species-by-sampling unit matrix as used previously

(Figure S1, Appendix 2). Because we do not have data on interaction strengths but only on interaction presence or absence, the LCBD<sub>FW</sub> values were derived from the Jaccard dissimilarities in interactions among all pairs of sampling units. More specifically, the food web dissimilarity  $D_{bi}$  between sampling units b and i is defined by  $D_{bi} = \frac{B+C}{A+B+C}$  where B (resp. C) corresponds to the number of interactions only present in the food web of sampling unit b (resp. i), while A corresponds to the number of interactions present in the food webs of both sampling units b and i. Second, LCBD<sub>FW</sub> values were tested for significance against the null hypothesis of random species distributions among waterholes and months (i.e. same null hypothesis as for LCBD<sub>Sp</sub> values). To do so, we built 999 permutations of the species-by-sampling unit matrix (Legendre and De Caceres 2013), on which we derived 999 random interaction-by-sampling unit matrices. Waterhole-month combinations with significant LCBD<sub>FW</sub> values correspond to sampling observations with trophic interactions that differ from food webs in other months and waterholes. Third, we also proposed a measure of Species contribution to beta diversity for food webs (SCBD<sub>FW</sub>) adapted from the approach of Legendre and Caceres (2013). We first suggest a calculation of the contribution of each interaction of the meta-food web to overall beta diversity following the same philosophy as Legendre and Caceres (2013) but using the Jaccard dissimilarity instead of the variance of the composition matrix as previously defined. Then we defined the  $SCBD_{FWi}$  of species i as half of the sum of the contributions to beta diversity of the interactions in which species i participates as prey or predator. By doing so, the sum of the SCBD<sub>FW</sub> values is equal to 1 as required. The precise definition and calculation of SCBD<sub>FW</sub> is detailed in Appendix 2. Please note that by construction species that are not connected in the meta- food web (i.e. inedible basal phytoplankton species) do not contribute 

to food web beta diversity because the beta diversity calculation is based on the interactionby-sampling unit matrix.

4 Describing local communities and species positions in the meta-food web

- 5 Species and trophic groups were first described by their frequency of occurrence in the
- 6 sampling units. The frequency of occurrence was defined by the number of sampling units in
- 7 which a given species occurs divided by the total number of sampling units. In addition,
- 8 species were characterized by their positions in the meta-food web. Species degree
- 9 corresponds to the number of interactions of a species in the meta-food web (i.e. total number
- of potential prey and predators). Species trophic level is defined by the average trophic
- position of its prey plus 1.
- We also characterized the local diversity and food web structure of each waterhole at each
- sampled month by the following measures: (i) total species richness; (ii) Shannon diversity as
- a function of species abundances; (iii) connectance as the number of interactions in the food
- web divided by the number of total possible interactions; (iv) average trophic level as the
- average of the trophic levels of species present in the food web.

 Statistical analyses

- 19 All statistical analyses were done with R. We checked for spatial autocorrelation in LCBD<sub>Sp</sub>
- and in LCBD<sub>FW</sub> at each sampling month. To do so, we assessed the significance of Moran's *I*
- using the R package ape. Both LCBD<sub>Sp</sub> and LCBD<sub>FW</sub> showed no sign of spatial
- autocorrelation in any of the months studied (Table S1). We tested the additive effects of
- waterhole management (i.e. pump presence), sampling month and location (i.e. HNP, forest or

communal area) on LCBD<sub>Sp</sub> and LCBD<sub>FW</sub> using a mixed linear model with the waterhole as a random effect using the lmer function of the lmerTest package (Kuznetsova et al. 2017). We then selected the more parsimonious model using the Akaike information criteria. We also tested the effects of species diversity, Shannon diversity, average trophic level and connectance on LCBD<sub>Sp</sub> and LCBD<sub>FW</sub> using a mixed linear model with the waterhole and the sampling month as random effects. Finally, we tested the effects of the species occurrence frequency, the species degree and its trophic level in the meta-food web on SCBD<sub>Sp</sub> and

#### **Results**

Species beta diversity

SCBD<sub>FW</sub> with a linear model.

The calculation of the LCBD<sub>Sp</sub> reveals dissimilarities between waterholes (Fig. 2A). Within the natural waterholes, among those that are significantly dissimilars from the others, only two are in HNP: Ballaballa (April) and Ngwenya (February). The others are either in the forestry area (Jwapi in February) or in the communal area (Mambanje in February and March, Marist in April, July and November, Nengasha in February, March and April, and Silewu in March and September). Among artificial waterholes, significantly dissimilar waterholes are mostly in HNP (Caterpillar in November, Kennedy 1, Livingi and Tchabema in February, Kennedy 2 in November, Mabuyamabema in February and March, Makwa in March, Ngweshla artificial in March, November and December, and Nyamandlovu and Tshebetshebe in April). Two waterholes from the forestry area are distinguishable: Ganda in November and December and Safari Lodge in February. We note that some waterholes are consistently similar (Jambile and Ngweshla natural, Sedina and Magoli for natural waterholes and Caterpillar, Dom, Dopi, Foster, Guvalala and Jambile artificial for artificial ones). We tested

 as additive factors the effects of the pump presence, the sampling month and the location of the waterholes to explain the LCBD<sub>Sp</sub>. The results of the model show that the pump presence has no effect but the sampling month and, marginally, the location affect the LCBD<sub>Sp</sub> (Table 1): the LCBD<sub>Sp</sub> values are significantly higher in November (p<0.001) and marginally higher in February (P=0.09) and they are significantly lower in the forestry (p < 0.03) and communal (p < 0.04) areas than in HNP. To analyse the SCBD<sub>Sp</sub>, we first looked at the species SCBD<sub>Sp</sub> rank (Figure S2), which reveals several groups of species according to their contribution to beta diversity (Figure 3B, black diamonds and inverted grey triangles). Tetraedron regulare and total nauplii explain 9% of the total SCBD<sub>Sp</sub>. With an additional set of 3 species, T. assymetricum, Euglena sp. and Anabaena sp, 20 % of the total SCBD<sub>Sp</sub> is explained. With 15 more species, that is a total of 20 species, up to 60% of the total SCBD<sub>Sp</sub> is explained. These species are by decreasing contribution order: Microcystis aeruginosa, T. limnetica, Phacus curvicauda, Aphanocapsa sp., Oscillatoria sp., Spyrogira sp., Melosira sp., Diatoma sp., Polyarthra vulgaris, Pediastrum simplex, Cymbella sp., Chroococcus sp., Brachionus calyciflorus, Cyclotella sp. and Camphylodiscus sp. We note that out of these 20 taxa, only three are herbivorous consumers (nauplii, P. vulgaris and B. calyciflorus), the others are primary producers. Among the phytoplankton taxa, we find mostly Bacillariophyta and Cyanobacteria. Food-web beta diversity The LCBD<sub>FW</sub> shows slightly different results than the LCBD<sub>Sp</sub>. Within natural waterholes, Nengasha, located in the communal area differentiates itself from other waterholes in March, July and November and Ngewshla natural, located in HNP, in March (Figure 2B). Within

artificial waterholes, most dissimilarities between waterholes are observed during the dry

- season in July, September and November and apply to waterholes located in HNP (Dom, Dopi, Jambile artificial, Kennedy 2 and Ngewshla artificial) or in the forestry area (Ganda). Only the sampling month has a significant effect on the LCBD<sub>FW</sub> (Table 1). The LCBD<sub>FW</sub> are higher in November (p = 0.008) and September (p = 0.04) and marginally higher in February (p = 0.04) =0.05). The analysis of the SCBD<sub>FW</sub> reveals that the two species that have the higher SCBD<sub>FW</sub> are two copepods, *Macrocyclops varicans* and *Eudiaptomus gracilis* (Figure 3B, grey triangles). They contribute to 9% of the total SCBD<sub>FW</sub>. The next three higher contributions explaining up to 21% of the total SCBD<sub>FW</sub> are *Daphnia pulex*, *B. calyciflorus* and the microphytoplankton group. To draw a parallel with the SCBD<sub>Sp</sub>, 60% of the SCBD<sub>FW</sub> is explained by an additional set of 16 species or group of species by order of decreasing contribution: bacteria, T. assymetricum, protists, P. vulgaris, T. limnetica, T. regulare, Filinia pejleri, nauplii, Cyclotella sp., Daphnia longispina, Euglena sp., Macrocyclops albidus, particulate and dissolved organic matter, Diaphanosoma tropicum, Afrocyclops gibsoni and Brachionus patulus (Figure 3B, black diamonds and grey triangles). Unlike SCBD<sub>Sp</sub>, this set of species gather together consumer species and primary producers. As species and food-web based LCBD and SCBD do not provide the same information about waterhole community and functioning dissimilarities, we plotted LCBD<sub>FW</sub> and SCBD<sub>FW</sub> as a function of species LCBD<sub>Sp</sub> and SCBD<sub>Sp</sub> respectively (Figure 3). Linear regressions show that they are positively correlated (adjusted  $R^2 = 0.11$  and p < 0.0001 for LCBD and adjusted  $R^2$ =0.77 and p <0.0001 for SCBD). The waterholes that have high contribution to both
  - LCDB<sub>Sp</sub> and LCBD<sub>FW</sub> are Jwapi and Mabuyamabema in February, Nengasha in March and Kennedy 2, Ganda and Ngweshla artificial in November (Figure 3A, black diamonds). The

species that have the higher contributions to both SCBD<sub>FW</sub> and SCBD<sub>Sp</sub> are the phytoplankton

 species *T. assymetricum*, *T. limnetica*, *T. regulare*, *Euglena* sp., *Cyclotella* sp. and the small herbivorous zooplankton *B. calyciflorus*, *P. vulgaris* and nauplii (Figure 3B, black diamonds).

4 Effects of diversity and food-web indices on LCBD and SCBD values

We explored the relationships between diversity, food-web indices, and species and food-web beta diversity. The LCBD<sub>Sp</sub> is positively correlated with the average trophic level of the waterhole food-webs and negatively correlated with Shannon diversity (Figure 4B and Table 2). The local contribution of a waterhole to beta diversity is thus higher when the average trophic level of the planktonic food-web it hosts is high and the species abundances are unbalanced. The LCBD<sub>FW</sub> decreases significantly with species richness and food-web connectance (Figure 4A and Table 2). Put another way, the more dissimilar waterholes from the interaction perspective are waterholes with few species and simple planktonic food-webs. The SCBD<sub>Sp</sub> increases significantly with species occurrence frequency and decreases significantly with species degree in the meta-food web (Figure 5). In other words, species that have high SCBD<sub>Sp</sub> are also widespread species, even if they might be in small abundances in numerous waterholes, and have a low number of connections in the meta-food web. As for the SCBD<sub>FW</sub>, it increases significantly with species occurrence frequency, and with species degree and trophic level in the meta-food web (Figure 5). In contrast with the SCBD<sub>Sp</sub>, species that most contribute to the SCBD<sub>FW</sub> are highly connected with other species of the meta- food web and have a high trophic level. These species may be either top-consumers, such as carnivorous copepods, or intermediate consumers with numerous prey and predators,

**Discussion** 

such as filter-feeder rotifers, cladoceran and nauplii.

Contrary to our first expectation, waterholes with artificial water pumping did not contribute less to spatiotemporal beta diversity than natural waterholes. Instead, the contribution of waterholes to species and interaction beta diversity was mostly related to monthly changes in their community composition. Species- and interaction-based approaches also gave complementary information on the spatiotemporal beta diversity of this freshwater ecosystem. While inedible phytoplankton species contributed importantly to species beta diversity, highly connected species in the food web contributed the most to interaction beta diversity. We discuss these different results in detail below. No significant effects of the presence of pumps on the contribution of waterholes to beta diversity Depending on the drought intensity and on the waterhole size and depth, most of the natural waterholes dry up during the dry season while the artificial ones only undergo water level fluctuations thanks to water pumping. While we expected that artificial pumping would decrease both spatial and temporal beta diversity by homogenizing environmental conditions across space and time, we show it has no significant impact on the contribution of the waterholes to species and interaction spatiotemporal beta diversity (LCDB<sub>Sp</sub> and LCBD<sub>FW</sub> respectively). This result is in agreement with the previous analyses of species community composition in this system. Indeed, Msiteli-Shumba et al (2017) showed that ground water pumping had no effect on the phytoplankton and zooplankton community structure, except in July for zooplankton (Msiteli-Shumba et al. 2017). The absence of difference in the beta diversity contribution between artificial and natural

waterholes, and the relatively weak spatial beta diversity observed, might be explained by

several complementary mechanisms. First, it might be explained by the fact that

 environmental conditions become harsher during the dry season in the artificial waterholes too and not only in the natural waterholes. Such harsh conditions in both artificial and natural waterholes might lead to similarly strong environmental filtering in both types of waterholes, consequently decreasing spatial beta diversity (Chase 2007). Indeed, in waterholes that do not dry, the water level strongly decreases and fluctuates from day to day. The surface area varies from 21 903 m<sup>2</sup> to 531 m<sup>2</sup> in the park from the rainy to the dry season (Msiteli-Shumba et al. 2017). In addition, large herbivores such as elephants, zebras, kudus and giraffes tend to aggregate around waterholes for drinking water (Chamaillé-Jammes, Valeix, et al. 2007; Ferry et al. 2016). Attendance favors dung deposition around waterholes which may induce water nutrient enrichment (Hulot et al. 2019). As a consequence, water conductivity, hardness, turbidity and ammonium concentration strongly increase during the dry season, while the macrophyte abundance decreases as well as water level (Msiteli-Shumba et al. 2018). Ammonium can reach concentrations that are toxic to plankton species, leading to strong differences in community composition between the dry and wet seasons in artificial waterholes, as found in natural waterholes. Our analysis is based on species that easily form resting stages and allow them to withstand adverse environmental conditions (Incagnone et al. 2015; Ellegaard and Ribeiro 2018). Such resting stage strategy may be developed by these species to avoid unfavorable habitats in permanent waterholes as much as in nature waterholes, resetting plankton communities during the dry season. At the onset of the rainy season, plankton communities could thus build up from the sediment egg bank in all waterholes whatever their dynamics during the dry season. Second, dispersal among waterholes might also contribute to reduced spatial beta diversity and to the lack of difference among natural and artificial waterholes. Plankton species are efficient passive dispersers (Incagnone et al. 2015) provided there is no barrier between favorable habitats (Juracka et al. 2016). The savannah landscape studied here is quite

 homogeneous and we did not find any sign of spatial autocorrelation in the contribution to species and interaction beta diversity. The mean distance between two waterholes is 23.1 km (SD: 12.9 km), the longest distance among waterholes being 59 km (*i.e.* distance between Mambanje and Jambile Artificiel). These distances are easily covered by potential vectors of dispersion such as wildlife and birds, which could contribute to connect waterholes and decrease spatial beta diversity of plankton communities. In summary, the similarity of species and interaction spatiotemporal beta diversity in natural and artificial ponds might rely on plankton resting stages that allow to endure the dry season and on dispersion. However, we do not know the weight of resting stages responding similarly to harsh conditions vs. migration in driving the observed spatial beta diversity, and more generally the relative importance of species sorting and mass effects on the spatial dynamics of these waterholes.

The contribution of waterholes to beta diversity depends on seasonal community changes. Our results show that temporal beta diversity can be an important component of the beta diversity of aquatic ecosystems in semi-arid areas, which are characterized by harsh environmental conditions during the dry season. Both the LCDB<sub>Sp</sub> and the LCBD<sub>FW</sub> of the waterholes depend significantly on the sampling month, the waterholes sampled in November during the dry season showing larger contributions to beta diversity. The importance of the seasonal dynamics for understanding the structure of these aquatic communities is consistent with previous findings (Msiteli-Shumba et al. 2017; Msiteli-Shumba et al. 2018; Hulot et al. 2020). Previous studies on this system indeed showed the planktonic community composition as well as food web structure varied mostly between the wet and the dry seasons rather than in relation with water pumping. Our results also highlight the importance of considering temporal beta diversity along with spatial beta diversity. Different waterholes indeed contribute significantly to beta diversity at different times. For instance, Nyamandlovu

 contributes significantly more than expected at random to species beta diversity in April only, whereas Ganda contributes significantly to species beta diversity in November and December. Although we expect to find a similar importance of temporal beta diversity in driving the beta diversity of waterholes in other years than the one considered here, we should acknowledge that our study was restricted to seven months and that the interannual variability should be addressed in future studies. In particular, the drought intensity changes from one year to the other and affects water level changes in ponds and, as a consequence, may affect planktonic community dynamics. Considering this study system over several years would allow to investigate how spatiotemporal beta diversity is affected by severe events such as prolonged droughts, and whether differences between natural and artificial waterholes occur in this context. The degree of uniqueness of the waterholes a given month depends on both diversity and food web structure The LCBD has been shown to distinguish ecosystems that are either very rich or degraded (Legendre and De Caceres 2013): high LCBD values might indeed indicate strongly different communities with either rare species combination or communities missing otherwise abundant and common species. In our study, higher values of LCBD<sub>Sp</sub> and LCBD<sub>FW</sub> are associated with plankton communities with lower species richness and diversity as well as lower connectance. Common species contribute the most to both species and food-web beta diversity, and highly connected species and top consumers are also the main contributors to food-web beta diversity. These results are consistent with the fact that our analysis of beta diversity highlights the waterholes sampled during the driest months. Several studies showed that catastrophic events such as drought may initiate losses of species and interactions, especially top predators inducing a food-web simplification (Ledger, Brown, Edwards,

Hudson, et al. 2013; Ledger, Brown, Edwards, Milner, et al. 2013; Jackson et al. 2020). In a

study analyzing the planktonic food-webs of roughly the same set of waterholes sampled in November 2012, February and July 2013, Hulot et al (2020) also showed that the food-webs during the dry season have lower connectance, less intermediate species and more basal species, lower omnivory index and lower mean trophic level than during the rainy season. Our present results of higher LCBD values in dry months are in line with these findings except for the mean trophic level, which increases with the LCDB<sub>Sp</sub>. This relationship may rely on the rich planktonic communities found during the rainy season where the top-consumers are more abundant. It might also relate with strong top-down effects of consumers on phytoplankton community composition in food webs with high mean trophic level, as phytoplankton species are the main contributors to species-level beta diversity in our system. Thus, the opposite response of the LCDB<sub>Sp</sub> to the average trophic level and the Shannon diversity is consistent with the fact that the dissimilarity pointed out by the LCBD pertains to both rich and degraded waterholes. Species and food-web beta diversity bring complementary understanding LCBD<sub>Sp</sub> and LCBD<sub>FW</sub> are only partly related to the same indices of community structure. Indeed, LCBD<sub>Sp</sub> increases with food-web average trophic level while LCBD<sub>FW</sub> decreases with connectance. This is also true for the SCBD values: while species contribution to food-web beta diversity increases with species degree and trophic level, species contribution to species-level beta diversity is lower for more connected species. These differences arise from the fact that by definition species degree matters to the calculation of food-web beta diversity, inedible basal species being necessarily excluded. In addition, our measure of species beta diversity includes species abundance, which is not the case for our measure of food-web beta diversity. Consequently, while the set of waterholes identified as unique by both indices are characterized by simple planktonic food-webs, waterhole uniqueness can also be defined by 

the opposite values of LCBD<sub>Sp</sub> and LCBD<sub>FW</sub> of some waterholes. Waterholes that do not

 contribute more than expected to beta diversity for one value may have a high value for the other ones. That is for instance the case for Nengasha and Jambile artificial in July, and Dopi in September and November, which have both low LCBD<sub>Sp</sub> and high LCBD<sub>FW</sub>. Although their planktonic communities are composed of common, evenly distributed phytoplankton species, they miss highly connected top consumer species that have pivotal role in the network such as E. gracilis and M. varicans. Conversely, some waterholes, such as Livingi in February, Ballaballa, Nyamandlovu and Tshebetshebe in April and Marist in November, have high LCBD<sub>Sp</sub> and low LCBD<sub>FW</sub>. These waterholes are characterized by the dominance of a few phytoplankton species in high abundance but which are poorly connected to other species in the food web. These species include cyanobacteria, which are excluded for the computation of the LCBD<sub>FW</sub> and SCBD<sub>FW</sub> because they have not involved in trophic interactions, and poorly edible phytoplankton species. In summary, the combination of the two indices allows to identify different sets of waterholes, which are likely related to very different functioning. Waterholes with high LCBD<sub>Sp</sub> and low LCBD<sub>FW</sub> can be characterized by either planktonic food-webs that are dominated by inedible or poorly connected phytoplankton species or by food-webs with relatively high species abundance at higher trophic levels (e.g. Makwa in March). In the former case, the primary production is diverted to dead-end or does not support efficiently a rich food-web whereas it is the contrary in the latter case. Meanwhile, waterholes with high LCBD<sub>FW</sub> are generally characterized by the lack of common and highly generalist top consumers, which might play an important functional role in these ecosystems. In conclusion, our analysis reveals that the beta diversity of plankton communities in the waterholes of a semi-arid savanna is mainly driven by strong temporal variations of species composition and food-web structure during the dry season. Meanwhile, the spatial variation of these communities does not appear related with water management (i.e. pumping) contrary to our initial hypothesis. This study highlights the importance of considering the spatio-temporal

dynamics of communities, and of combining approaches based on species composition and trophic interactions. To predict the effects of global change stressors on food-web structure and dynamics, we need simplified and intuitive indices related to community health (Heleno et al. 2020). The species and food-web indices are complementary as they help to understand why waterholes are dissimilar in space and time, bridging further the taxonomical and

functional perspectives needed to the study of beta diversity.

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**Table 1**: Effects of sampling month, pump presence and location on LCBD values. The table reports the results for the complete models.

LCBD values	Explanatory variables	df	Chisq (for LCBD <sub>Sp</sub> ) or F (for LCBD <sub>FW</sub> )	P value
LCBD <sub>Sp</sub>	Month	6	22.80	0.0008
	Pump presence	1	0.018	0.89
	Location	2	5.167	0.07
LCBD <sub>FW</sub>	Month	6	2.955	0.0097
	Pump presence	1	0.03	0.8628
	Location	2	0.471	0.6256

**Table 2**: Effects of diversity and food web indices on LCBD and SCBD values. The table reports the results for the best models.

LCBD and SCBD values	Explanatory variables	df	Estimates	Chisq (for LCBD) or F (for SCBD)	P value
LCBD <sub>Sp</sub>	Average trophic level	1	4.587e-03	34.113	<0.0001
	Shannon diversity	1	-2.290e-03	97.793	<0.0001
LCBD <sub>FW</sub>	Species richness	1	-3.020e-05	15.152	<0.0001
	Connectance	1	-1.234e-03	7.2067	0.0072
SCBD <sub>Sp</sub>	Species occurrence frequency	1	2.917e-02	115.99	<0.0001
	Species degree in metaweb	1	-9.339e-05	5.4712	0.0212
SCBD <sub>FW</sub>	Species occurrence frequency	1	3.413e-02	188.822	<0.0001
	Species degree in metaweb	1	1.611e-04	55.633	<0.0001
	Trophic level in metaweb	1	4.590e-03	19.492	<0.0001

#### Figure legends

Figure 1. Map of the study site and food-web examples. (A) Map with Hwange National Park, the forestry and the communal areas and the waterhole locations. B and C identify the waterholes Nengasha and Ganda. (B) and (C) Representation of the food-webs at the different dates (February to December) in Nengasha and Ganda respectively.

#### Figure 2. Species (a) and food-web (b) based LCBD over all waterholes and dates.

The two color scales are proportional to the LCBD. The while stars denote significantly dissimilar waterholes. The dates are February (Feb), March (Mar), April (Apr), July (Jul), September (Sep), November (Nov) and December (Dec). The waterholes are separated in the natural (left) and articifial (right) waterholes. The abbreviations are by order of appearance Ballaballa (Bal), Jambile Natural (JamNat), Mabuyamabema (Mab), Jwapi, Magoli (Mag), Mahule (Mahu), Mambanje (Mamb), Marist (Mar), Mazibomvu (Maz), Nengasha (Nenga), Ngweshla Natural (NgweNat), Ngwenya, Sedina (Sed), Silewu (Sil), Caterpillar (Cat), Dom, Dopi (Dop), Foster (Fos), Ganda (Gan), Guvalala (Guv), Jambile Artificial (JamArt), Kennedy 1 (Ken1), Kennedy 2 (Ken2), Livingi (Liv), Makwa (Mak), Ngweshla Artificial (NgweArt), Nyamandlovu (Nyam), Safari lodge (Saf), Tchabema (Tchab) and Tshebetshebe (Tsheb).

### Figure 3. Species and food-web (A) LCDB and (B) SCBD over all waterholes and dates.

In both panels, black diamonds, grey triangles and inverted grey triangles identify respectively waterholes or species – groups that have highest values for both indices, foodweb based index and species based index. The plain line represents the correlation line. (A) Particular pans are identified by their abbreviation (see Figure 2) and first letter of the sampling month. (B) Species abbreviations are *Afrocyclops gibsoni* (AG), *Anabaena* sp.

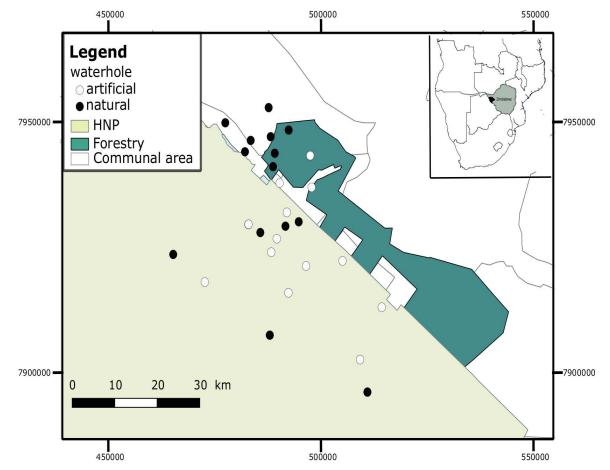
(Ansp), bacteria (Bact), Brachionus calyciflorus (BC), Camphylodiscus sp. (CAsp), Chroococcus sp. (CHsp), Cymbella sp. (CyMsp), Cyclotella sp. (Cysp), Daphnia longispina (DLO), Daphnia pulex (DP), Diatoma sp. (Dsp), Diaphanosoma tropicum (DT), Eudiaptomus gracilis (EG), Euglena sp. (Esp), Filinia pejleri (FP), Microcystis aeruginosa (MA), Macrocyclops albidus (MAL) Melosira sp. (MELsp) microphytoplankton (MicroPhyto), Megacyclops viridis (MV), Oscillatoria sp. (Ossp), Phacus curvicauda (PHC), particulate and dissolved organic matter (PomDom), protists (Prot), Pediastrum simplex (PS), Pinnularia sp. (Psp), Polyarthra vulgaris (PV), Spyrogira sp (SPYsp), Tetraedron assymetricum (TA), nauplii (TN), T. limnetica (TL) and T. regulare (TR).

Figure 4. Food-web (grey square) and species (yellow triangle) based LCBD as a function of (a.) Shannon diversity, (b.) food web average trophic, (c.) species richness and (d.) connectance with significant regression lines (see text).

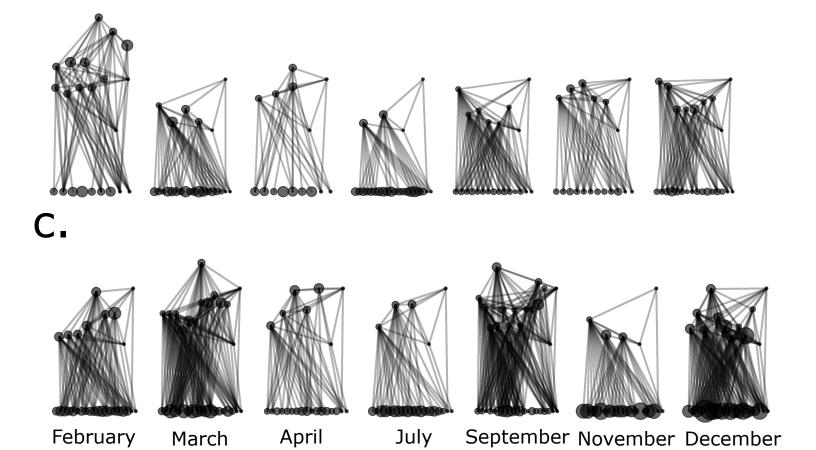
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Figure 5. Food-web (grey square) and species (yellow triangle) based SCBD as a function of (a.) species frequency of occurrence, (b.) species degree in metaweb and (c.) species trophic level in metaweb with significant regression lines (see text).

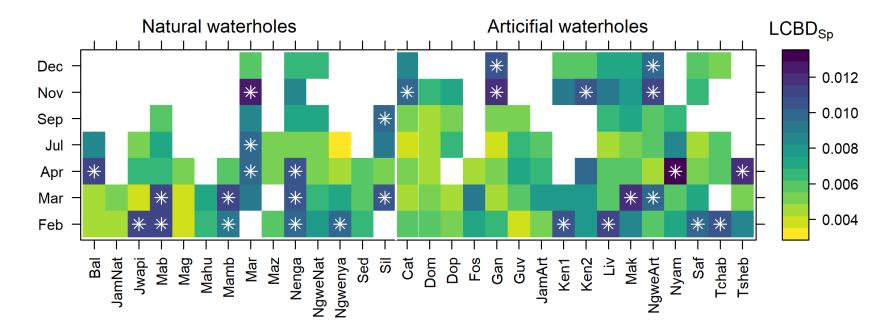
a.



b.



a.



# b.

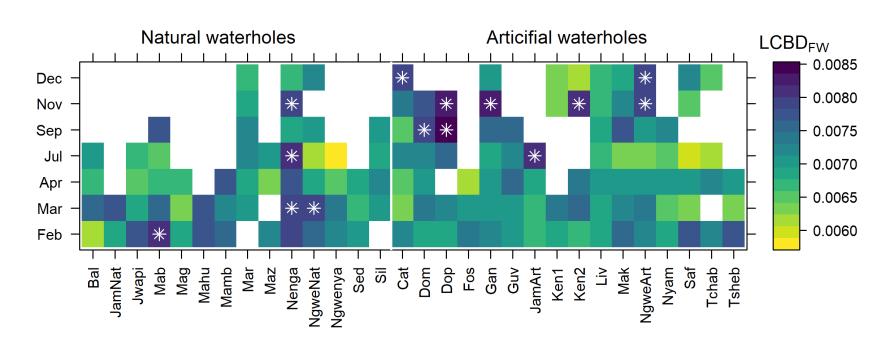
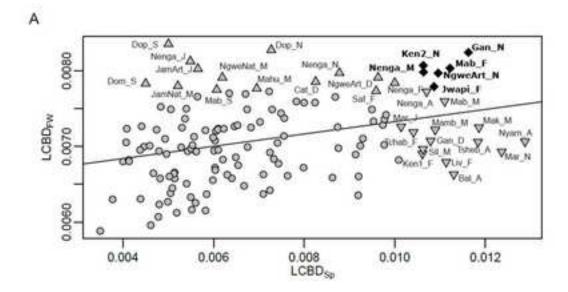
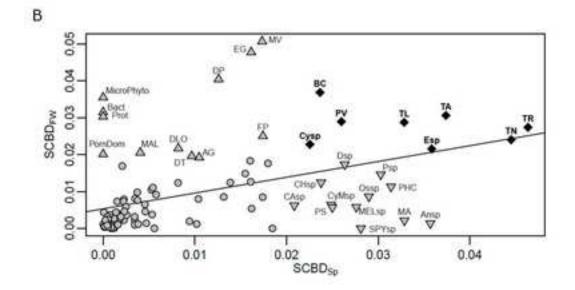
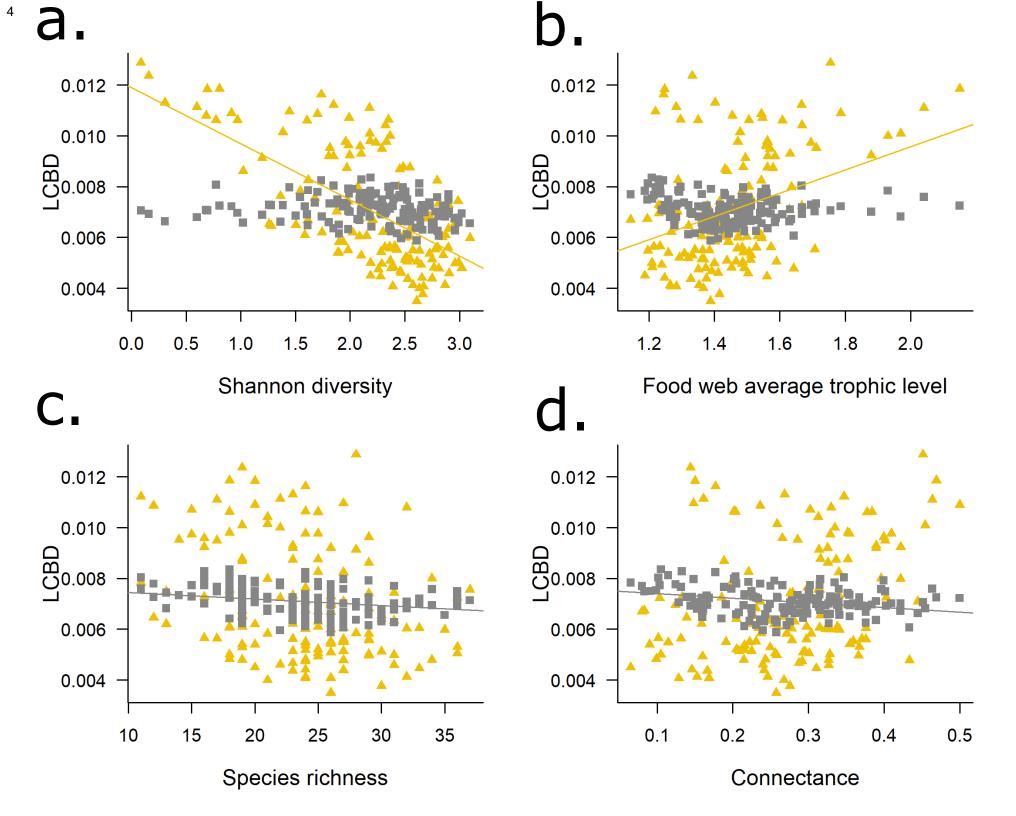
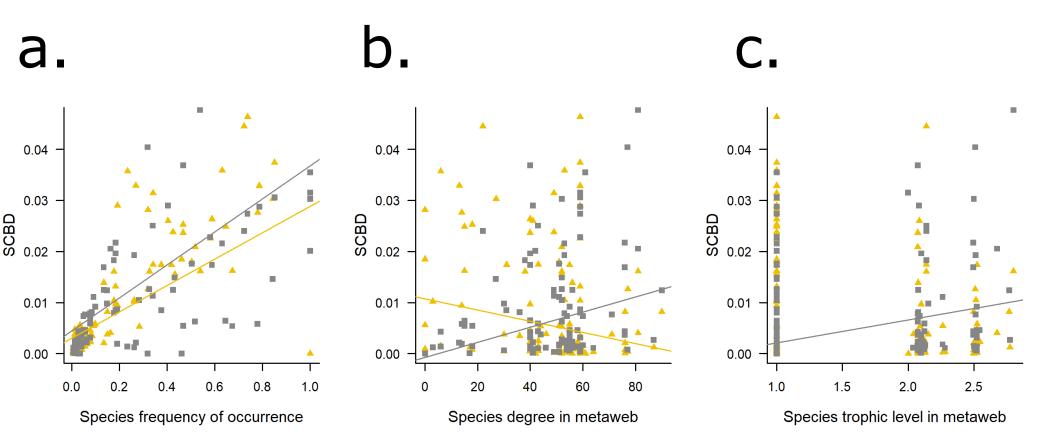


Figure 3









## Appendix 1

Table S1: Spatial autocorrelation of LCBD $_{\rm Sp}$  and LCBD $_{\rm FW}$  by sampling month, as measured by Moran's I.

Month	LCE	$\mathrm{BD}_{\mathrm{Sp}}$	LCB	$D_{FW}$
Month	Moran's I	P value	Moran's I	P value
February	-0.003	0.64	0.086	0.08
March	-0.004	0.65	-0.018	0.79
April	-0.03	0.89	-0.029	0.88
July	-0.11	0.51	0.069	0.20
September	0.082	0.25	-0.099	0.88
November	-0.094	0.97	-0.17	0.46
December	-0.18	0.55	0.06	0.35

Figure S1: Schematic diagram representing the procedure to build the interaction-by-sampling unit matrix from the meta food-web and the species composition of sampling units. The interaction-by-sampling unit matrix is then used to compute dissimilarities among sampling units and LCBD values, as done originally on the species-by-sampling unit matrix (Legendre and De Caceres 2013).

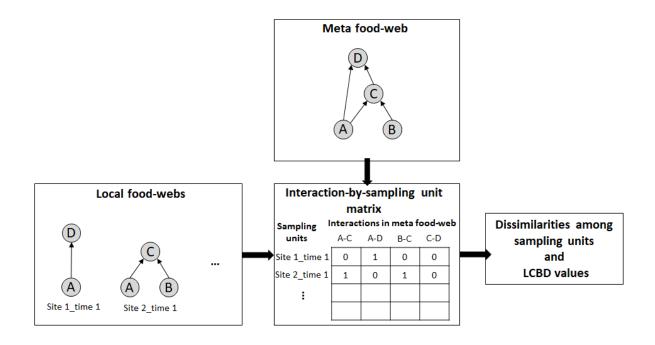
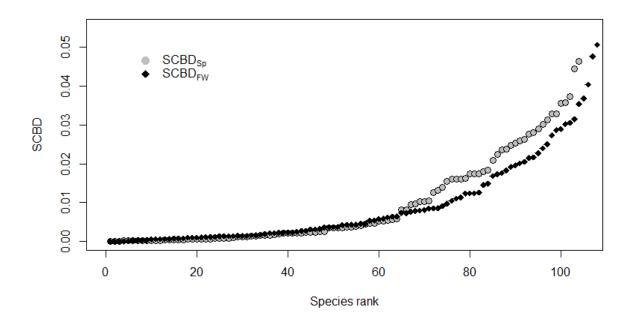
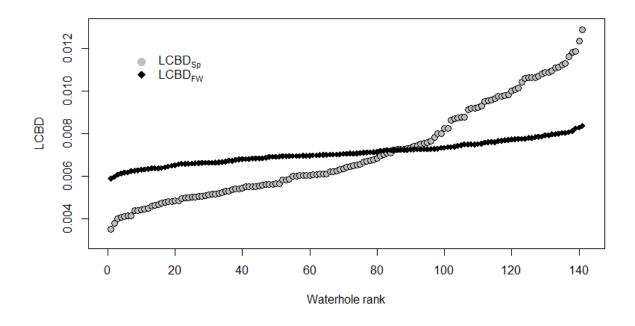


Figure S2. Species and food-web based SCBD and LCBD as a function of species (A) and waterhole (B) ranks.

A.



B.



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Actinastrum hantzchii Afrocyclops gibsoni MicroPhyto Anabaena
     Ankistrodesmus spiralis Ankistrodesmus spirilliformis
sp.
     Arthrodesmus sp. Bacteria Bosmina brehmi
                                                   Brachionus ungularis
     Brachionus calyciflorus
                              Brachionus caudatus
                                                          Brachionus
dimidiatus Brachionus dolabratus Brachionus falcatus
                                                          Brachionus
havanaensis Brachionus patulus
                                  Brachionus plicatilis Brachionus
quadridentatus Brachionus urceusCamphylodiscus sp.
                                                          Cephalodella
qibba Ceratium sp.
                     Ceriodaphnia cornuta Ceriodaphnia laticaudata
     Chyrodidae non-limnetica
                                  Ceriodaphnia paradoxa Ceriodaphnia
                            Chroococcus sp. Closterium sp.
reticulata Chlorella sp.
                                       Cyclops vicinus Cyclotella sp.
     Cosmarium sp.
                     Cracticula sp.
     Cymbella sp.
                      Dactylococcopsis sp. Daphnia gessneri Daphnia
laevis
           Daphnia longispina Daphnia monacha Daphnia pulex
     Desmidium sp.
                      Diaphanosoma dentatum Dinobryon setularia
     Diaphanosoma tropicum Diatoma sp. Diaphanosoma sarsi
     Eudiaptomus gracilis Eudorina elegans Epactophanes richardii
     Euglena sp. Filinia longiseta Filinia pejleri Gleocystis sp.
     Gomphosphaeria sp.
                            Hyalotheka sp. Keratella lenzi Keratella
           Keratella tecta Keratella tropicaKeratella vulga Lecane
bulla Lecane leontina Lecane luna Macrocyclops albidus Metaboeckella
          Megacyclops viridis
                                 Melosira sp.
                                                  Merismopedia sp.
dilatata
     Maetacyclops minutus Micractinium sp. Micrasterias sp.
     Microcystis aeruginosa Nauplii
                                       Oscillatoria
                                                         Paracyclops
           Paracyclops chiltoni
                                  Paracyclops fimbriatus Paracyclops
poppei
           Pediastrum simplex
                                  Phacus curvicauda Pinnularia sp.
      Polyarthra vulgaris
                             PomDom
                                        Protists
                                                    Scapholebris kingi
     Scenedesmus quadricauda
                                                    Staurastarum
                                  Spyrogira sp.
brachiatum Sididae non-limnetica Staurastaram paradoxum Staurastram
polymorphum Staurastrum tetracerum Synedra sp. Tetraedron assymetricum
     Tetraedron limnetica Tetraedron regulare
                                                    Trichotria tetractis
     Thermocyclops oblongata
                                  Trichotria elongata
                                                         Trichocerca
           Trichocerca similis
                                  Trichotria tetractis
pusilla
                                                          Tropocyclops
           Tropocyclops tenellis Volvox sp.
prasinus
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Ceriodaph	nia co		0	0	0	0	0	0	0	0	0
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Chlorel		0 1	1 1	0 1	0 1	0 1	0 1	0 1	0 1	1 0	1 1
1 0	1 1	1	1	1	1	0	0	0	0	0	0
1		0	0	1	1	1	1	1	0	1	1
1		0	1	0	1	0	1	1	0	0	0
1		1	1	1	1	1	1	1	1	1	0
0	1	0	0	0	1	0	1	1	1	1	0
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0	0	0	0	0	0	1	1	1	1	1	1
1		O	U	9	J	_	_	Τ.	_	_	_
_	O										

Chroococcus sp.  1	0 0 1 0	1 1 1 0	0 1 1	0 1 1	0 1 0 1	0 1 0 1	0 1 0 1	0 1 0 0	1 0 0	1 1 0 1
1 0	0	0	0	1	0	0	1	0	0	0
1 1 0 1	1 0	1 0	1 0	0	0	0 1	0 1	1 1	0 1	0
0 0	1	0	0	1	0	0	0	1	0	0
0 0	0	0	0	0	1	1	1	1	1	1
1 0			_				_			
Closterium sp. 0 0	0 0	0	0	0	0	0	0	0 0	0 0	0
0 0	0	0	0	0	0	0	0	0	0	0
1 0	0	0	0	0	0	0	0	0	0	0
0 0	0	0	0	0	0	0	0	0	0	0
0 0	0 0	0	0	0	0	0	1	0	1 0	0
0 0	0	0	0	0	0	0	0	0	0	0
0 0	0	0	0	0	0	0	0	0	0	0
0 0			_				_			
Coelastrum sp.	0 0	1 1	0 1	0	0	0	0	0 1	1 0	1
1 1 0 1	1	1	1	1 1	1 0	1 0	1 0	0	0	1 0
1 0	0	0	1	1	1	1	1	0	1	1
1 0	0	1	0	1	0	1	1	0	0	0
1 1	0	1	1	0	0	0	1	1	1	0
0 1 0 0	0 1	0	0	0 1	0	1 0	1 0	1 1	1 0	0
0 0	0	0	0	0	1	1	1	0	0	1
1 0										
Cosmarium sp.	0	0	0	0	0	0	0	0	0	0
0 0 0 1	0 1	0 1	0 1	0 1	0	0 0	0	0	0	0
1 0	0	0	1	1	1	1	1	0	1	1
1 0	0	0	0	0	0	0	0	0	0	0
0 0	0	0	0	0	0	0	1	0	1	0
0 0	0 0	0	0 0	0 1	0	0 0	0 0	0	0	0
0 0	0	0	0	0	0	0	0	0	0	0
0 0										
Cracticula sp.	0	1	0	0	0	0	0	0	1	1
1 1 0 1	0 1	1 1	1 1	1 1	1 0	1 0	1 0	1 0	0 0	1 0
1 0	0	0	1	1	1	1	1	0	1	1
1 0	0	1	0	1	0	1	1	0	0	0
1 1	1	1	1	0	0	0	1	1	1	0
0 1 0 0	0 1	0	0 0	0 1	0 0	1 0	1 0	1 1	1 0	0
0 0	0	0	0	0	1	1	1	0	0	1
1 0										
Cyclops vicinus	0	0	0	0	0	0	0	0	0	0
0 0	0 0	0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0
0 0	0	0	0	0	0	0	0	0	0	0

0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0
0 Cyclotella 0 1 1 1 0 0	0 a sp. 1 1 0 0 1 1 0 0 0	0 1 1 0 0 1 0	1 1 1 0 1 1 0 0	0 1 1 1 0 1 0 0	0 1 1 1 1 1 1 1	0 1 0 1 0 1 0 0	0 1 0 1 1 1 1 0	0 1 0 1 1 1 1 0	0 1 0 0 0 1 1 1	1 0 0 1 0 1 1 0	1 1 0 1 0 0 0 0
Eudiaptom	0		0 0 1 0 0 0 0	0 0 1 0 0 0 0	0 0 1 1 0 0 0	0 0 1 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 1 0	0 0 0 0 0 0	1 0 0 0 0 0 1 0
Dactylocol 0 1 0 1 0 0 0 0 0 0 1 1 1 1 1 1 1 1 1			0 1 1 0 0 1 0 1	1 1 0 1 1 0 0	0 1 1 1 0 1 0 0	0 1 1 1 1 1 1 1 0	0 1 0 1 0 1 0 0	0 1 0 1 0 1 1 0	0 1 0 1 0 1 1 0	0 1 0 0 0 1 1 0	1 0 0 1 0 1 1 0
Daphnia go 0 0 1 0 0 0 0 0 0 0 0 0 0 0 0 0		-	0 0 0 0 1 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 1 0 0	0 0 0 0 0 0 0	0 0 0 0 0 1 0 0	0 0 0 0 0 0 0
Daphnia 10 0 0 1 0 0 0 0		0 0 0 0 0 0	0 0 0 0 1 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0 1 0	0 0 0 0 0 0	0 0 0 0 0 0 1 0	0 0 0 0 0 0

0	0	0	0	0	0	0	0	0	0	0	0
0 Danhnia	0	no	0	0	0	0	0	0	0	0	0
Daphnia O	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	1	0	0	0	0	0	0	0	0	0	0
0	0	0	0	1	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	1	0	1
0	0	0	0	0	0	0	0	0	0	0	0
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0	0	0	0	0	0	0	0	0	0	0	0
0	0	0									
Daphnia	monacha	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
1	0	0	0	0	0	0	0	0	0	0	0
0	0	0	1	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	1	0	1	0
0	0	0	0	0	0	0	0	0	0	0	0
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0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
Daphnia	_	0	0	0	0	0	0	0	0	0	0
0	0 0	0 0	0	0	0	0	0	0	0	0	0
1	0	0	0	0	0	0	0	0	0	0	0
0	0	0	1	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	1	0	1	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0										
Desmidiu	m sp.	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
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0	0	0	0	0	0	0	0	0	0	0	0
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Diaphano 0	soma der 0	o 0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	1	0	0	0	0	0	0	0	0	0	0
0	0	0	0	1	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	1	0	1
0	0	0	0	0	0	0	0	0	0	0	0
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0	0	0	0	0	0	0	0	0	0	0	0
0	0	0									
Diaphano	soma sar	rsi	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0

0 0 0 0 0	0 1 0 0 0 0	0 0 0 0 0	0 0 0 0 0 0	0 0 1 0 0 0	0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0	0 0 0 0 0	0 0 0 1 0 0	0 0 0 0 0	0 0 0 1 0 0
0 Diaphano	0 osoma tro	0 nicum	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	1	0	0	0	0	0	0	0	0	0	0
0	0	0	0	1	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	1	0	1
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0 1	0	0	0	0	0	0	0	1	1
Diatoma 1	sp. 0	1	0 1	0 1	0 1	0 1	0 1	0 1	0	1 1	1 0
0	0	0	0	0	0	0	0	0	0	0	1
0	0	0	1	1	1	1	1	0	0	0	0
0	0	1	0	1	0	0	0	0	0	0	1
1	1	1	1	0	0	0	1	1	1	0	0
1	0	0	0	0	0	1	1	1	1	0	0
0	1	0	0	1	0	0	0	1	0	0	0
0	0	0	0	0	1	1	0	0	0	1	1
	_			_							
	on setula		0	1	0	0	0	0	0	0	1
1	1	1	1	1	1	1	1	1	1	1	0
1	0 1	1 0	1 0	1 0	1 0	1 0	0 0	0	0	0	0
1	1	0	0	1	0	1	0	1	1	0	1 0
0	1	1	1	1	1	1	1	1	1	1	1
0	0	1	0	0	0	1	0	1	1	1	1
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0	0	0	0	0	0	0	1	1	1	1	1
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	omus grac		0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
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0	0	0	0	0	0	0	0	0	0	0	0
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Eudorina	a elegans	0	0	0	0	0	0	0	0	0	0
1	1	0	1	1	1	1	1	1	1	0	1
0	1	1	1	1	1	0	0	0	0	0	0
1	0	0	0	1	1	1	1	1	0	0	1
0	0 1	0	1 1	0 1	1	0	0	0 1	0	0 1	0
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0	0	0	0	0	0 1	0	0	0	0 1	0	0
0	0	0	0	0	0	0	0	0	0	0	0
	hanes ric	hardi	i 0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
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0	0	0	0	0	1	1	1	1	1	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0 0	0 0	0 0	0	0 0	0 0	0 0	0
0	0	0	0	0	0	1	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	Ü	Ŭ	Ü	Ü	Ŭ	Ŭ	Ŭ	Ŭ	Ŭ
Euglena	sp. 0	1	0	0	0	0	0	0	1	0	1
1	0	1	1	1	1	1	1	1	0	1	0
1	1	1	1	1	0	0	0	0	0	0	1
0	0	0	1	1	1	1	1	0	1	1	1
0	0	1	0	1	0	0	0	0	0	0	1
1	1	1	1	1	1	1	1	1	1	0	0
1	0	0	0	0	0	1	1	1	1	0	0
0	1	0	0	1	0	0	0	1	0	0	0
0	0	0	0	0	1	1	0	1	1	1	1
0 Filinia	longiset	· a ()	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
1	0	0	0	0	0	0	0	0	0	0	0
0	0	0	1	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	1	0	1	0
0	0	0	0	0	0	0	0	0	0	0	0
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0	0	0	0	0	0	0	0	0	0	0	0
1	0	0	0	0	0	0	0	0	0	0	0
0	0	0	1	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	1	1	1	0
0	1	0	0	0	0	0	1	1	1	1	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	1	0	0	0	0	1
1	0	0	1	0	0	0	0	0	0	1	1
Gleocys <sup>*</sup>	tis sp. 1	0 1	1 1	0 1	0 1	0 1	0 1	0 1	0 1	1 0	1 1
0	1	1	1	1	1	0	0	0	0	0	0
1	0	0	0	1	1	1	1	1	0	1	1
1	0	0	1	0	1	0	1	1	0	0	0
1	1	1	1	1	1	1	1	1	1	1	0
0	1	0	0	0	1	0	1	1	1	1	0
0	0	1	0	0	1	0	0	0	1	0	0
0	0	0	0	0	0	1	1	1	1	1	1
1	0										

Gomphosphaeria s	SP. 1 1 0 0 0	0 0 1 0 0 1	1 1 1 0 1 1	0 1 1 1 0 1	0 1 1 1 1 0	0 1 0 1 0 0	0 1 0 1 0 0	0 1 0 1 1 1	0 1 0 0 0 1	1 0 0 1 0 1
0 0 0 0 1 1	0 0	1 0	0	0	1	0	0	0	0	0
Hyalotheka sp.	0	0	0	0	0	0	0	0	0	0
0 0	0	0	0	0	0	0	0	0	0	0
0 0	0 0	0	0	0	0	0	0	0 0	0 0	0
0 0	0	0	0	0	0	0	0	0	0	0
0 0	0	0	0	0	0	0	0	0	0	0
0 0	0	0	0	0	0	0	0	0	0	0
0 0	0 0	0	0	0	0	0	0	0	0 0	0
0 0	U	U	U	U	U	U	U	U	U	U
Keratella lenzi	0	1	0	0	0	0	0	0	1	0
0 0	0	0	0	0	0	0	0	0	0	0
0 0	1	1	0	0	0	0	0	0	0	0
1 0 0	0 0	0 1	1 0	1 0	1 0	1 0	1 0	0	0	0
0 0	0	0	0	0	0	0	1	1	1	0
0 1	0	0	0	0	0	1	1	1	1	0
0 0	0	0	0	1	0	0	0	0	0	0
0 0 1 0	0	0	0	0	1	0	0	0	0	1
Keratella quadra	nta	0	1	0	0	0	0	0	0	1
0 0	0	0	0	0	0	0	0	0	0	0
0 0	0	1	1	0	0	0	0	0	0	0
0 1	0	0	0	1	1	1	1	1	0	0
0 0	0 0	0	1 0	0	0 0	0	0	0 1	0 1	0 1
0 0	1	0	0	0	0	0	1	1	1	1
0 0	0	0	0	0	1	0	0	0	0	0
0 0	0	0	0	0	0	1	0	0	0	0
1 1	0	1	0	0	0	0	0	0	1	0
Keratella tecta 0 0	0 0	1 0	0	0 0	0	0	0	0	1 0	0
0 0	1	1	0	0	0	0	0	0	0	0
1 0	0	0	1	1	1	1	1	0	0	0
0 0	0	1	0	0	0	0	0	0	0	0
0 0	0	0	0	0	0	0	1 1	1 1	1 1	0
0 1 0 0	0 0	0	0	0 1	0	1 0	0	0	0	0
0 0	0	0	0	0	1	0	0	0	0	1
1 0			-			-				
Keratella tropio		1	0	0	0	0	0	0	1	0
0 0	0	0	0	0	0	0	0	0	0	0
0 0 1 0	1 0	1 0	0 1	0 1	0 1	0 1	0 1	0	0	0
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0 0 0 0	0 0 1 0	0 0 0 0	1 0 0 0 0	0 0 0 0	0 0 0 1 0	0 0 0 0	0 0 1 0	0 1 1 0	0 1 1 0 0	0 1 1 0 0	0 0 0 0
1 Keratella 0 0 1 0 0 0 0	0 vulga 0 0 0 0 0 1 0	0 0 1 0 0 0 0	1 0 1 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 0 0 0	0 0 0 1 0 0 1 0	0 0 0 1 0 1 1 0	0 0 0 0 0 1 1 0	1 0 0 0 0 1 1 0	0 0 0 0 0 0 0
1 Lecane bu 0 0 1 0 0 0	0 0 0 0 0 1 0	0 0 1 0 0 0 0	1 0 1 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 0 1	0 0 0 1 0 1 1 0	0 0 0 0 0 1 1 0	1 0 0 0 0 1 1 0	0 0 0 0 0 0 0
1 Lecane le 0 0 1 0 0 0	0 0 0 0 0 1 0	0 0 1 0 0 0 0	1 0 1 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 0 0 1	0 0 0 1 0 0 0 0	0 0 0 1 0 0 1 0	0 0 0 1 0 1 1 0	0 0 0 0 0 1 1 0	1 0 0 0 0 1 1 0	0 0 0 0 0 0 0 0
1 Lecane lu 0 0 0 0 0 1 0	0 na 0 0 1 0 0 0 0 0 0 0 0 0	1 0 1 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 0 0 1	0 0 0 1 0 0 0 0	0 0 0 1 0 0 1 0	0 0 0 1 0 1 1 0	0 0 0 0 0 1 1 0	1 0 0 0 0 0 1 1 0	0 0 0 0 0 0 0 0	0 0 1 0 0 0 0 0
0 Macrocycl 0 0 0 0 0 0	ops alk 0 0 1 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 1 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 1 0

0 0	0 0	0	0	0	0	0	0	0	0
Megacyclops vir	idis (		0	0	0	0	0	0	0
0 0	0 0		0	0	0	0	0	0 0	0
0 1	0 0		0	0	0	0	0	0	0
0 0	0 0		0	0	0	0	0	0	0
0 0	0 0		0	0	0	0	0	0	1
0 0	0 0		0	0	0	0	0	0	0
0 0	0 0	_	0	0	0	0	0	0	0
0 0	0 0	0	0	0	0	0	0	0	0
0 0	0								
Metaboeckella d	ilatata (	0	0	0	0	0	0	0	0
0 0	0 0	0	0	0	0	0	0	0	0
0 0	0 0	0	0	0	0	0	0	0	0
0 1	0 0	0	0	0	0	0	0	0	0
0 0	0 0		0	0	0	0	0	0	0
0 0	0 0	_	0	0	0	0	0	0	1
0 0	0 0		0	0	0	0	0	0	0
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Melosira sp.	0 0		0	0	0	0	0	0	0
0 0	0 0		0	0	0	0	0	0	0
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0 0	0 0		1	1	1	1	0	1	1
1 0	0 0	_	0	0	0	0	0	0	0
0 0	0 0	_	0	0	0	0	0	0	0
0 0	0 0	_	0	0	0	0	0	0	0
0 0	0 (	_	1	0	0	0	0	0	0
0 0 0 0	0 (	0	0	0	0	0	0	0	0
Merismopedia sp	. 0 0	) 0	0	0	0	0	0	1	1
Merismopedia sp	0 1		1	1	1	1	1	0	1
0 1	1 1		1	0	0	0	0	0	0
1 0	0 0		1	1	1	1	0	0	0
0 0	0 1		1	0	0	1	0	0	0
1 0	0 0		0	0	0	1	0	1	0
0 0	0 0		0	0	0	0	0	0	0
0 0	0 0		1	0	0	0	0	0	0
0 0	0 0		0	0	0	0	0	0	0
0 0									
Maetacyclops min	nutus (	0	0	0	0	0	0	0	0
0 0	0 0	0	0	0	0	0	0	0	0
0 0	0 0	0	0	0	0	0	0	0	0
0 1	0 0		0	0	0	0	0	0	0
0 0	0 0		0	0	0	0	0	0	0
0 0	0 0		0	0	0	0	0	0	1
0 0	0 0		0	0	0	0	0	0	0
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0 0	0								
Micractinium sp	. 0 1		0	0	0	0	0	1	1
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0 1 1 1 0 0	1 0 0 1 1 0	1 0 0 1 0 1	1 0 1 1 0 0	1 0 1 0 0	1 1 0 0 1	0 1 0 0 0 0	0 1 1 0 1 0	0 1 1 1 1 0	0 0 0 1 1 1	0 1 0 1 1 0 0	0 1 0 0 0 0
1 Micraster: 0 0 0 0 0 0 0	0 ias sp. 0 0 0 0 0 0 0 0 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0
Microcyst: 1 0 0 0 0 0 0 0 0		ginosa 0 0 0 0 0 1 0 0 0 0	0 0 0 0 0 0 0 0 1 0 0 0 0 0 0 0 0 0 0 0	0 0 0 0 0 1 0 0	0 0 0 1 0 1 0 0	0 0 0 1 0 0 0 1	0 0 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 0 0 0 0 0	0 0 0 0 0 0
Nauplii 0 0 0 0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 1 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 1 0 0	0 0 0 0 0 0	0 0 0 0 0 0 1 0 0	0 0 0 0 0 0	0 0 1 0 0 0 0
Oscillato: 1 0 1 1 1 0 0 0 0 0 1	1 0 0 1 1 0	0 1 1 0 0 1 0 1	1 1 0 1 1 0 0	0 1 1 1 0 1 0 0	0 1 1 1 1 1 1 1 0	0 1 0 1 0 1 0 0	0 1 0 1 1 1 1 0	0 1 0 1 1 1 1 0	0 1 0 0 0 1 1 1	1 0 0 1 0 1 1 0	1 0 1 0 0 0 0
Paracyclop 0 0 0 0	0 ps affi 0 0 1 0	nis 0 0 0 0	0 0 0 0 0	0 0 0 0 1	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0 0

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Paracyclo	ps chi	ltoni	0	0	0	0	0	0	0	0	0
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0	0	0	0	0	0	0	0	0	0	0	0
0	1	0	0	0	0	0	0	0	0	0	0
0	0	0	0	1	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	1
0	0	0	0	0	0	0	0	0	0	0	0
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0	1	0	0	0	0	0	0	0	0	0	0
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Paracyclo	ps pop	pei	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	1	0	0	0	0	0	0	0	0	0	0
0	0	0	0	1	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	1
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0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	1
Pediastru	_		0	0	0	0	0	0	0	0	1
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Phacus cu	rvicau	ıda 0	1	0	0	0	0	0	0	1	0
1	1	0	1	1	1	1	1	1	1	0	1
0	1	1	1	1	1	0	0	0	0	0	0
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0	1	0	0	0	0	0	1	1	1	1	0
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Pinnularia 1 0 1 0 0 0 0 0 0 0 0	sp. 1 1 0 0 0 0 0 0 0 0	0 0 1 0 0 0 0 0	0 1 1 0 1 0 0 0	0 1 1 1 0 0 0	0 1 1 1 1 0 0	0 1 0 1 0 0 0	0 1 0 1 0 0 0	0 1 0 1 0 1 0 0	0 1 0 0 0 0 0 0	1 0 0 0 0 1 0 0	0 1 0 0 0 0 0
Polyarthra	vulg 0 0 1 0 0 0 0 0	aris 0 0 0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 0 0 1	0 0 0 1 0 0 0	0 0 0 1 0 0 0	0 0 0 1 0 1 0 0	0 0 0 0 0 0 0	0 0 0 0 0 1 0 0
PomDom  1  1 0 0 1 0 0 0 0 0 0 0 0	0 1 1 0 0 1 0 0	0 1 1 0 0 1 0 0	0 1 1 0 1 0 0 0	0 1 1 1 1 1 0 1	0 1 0 1 0 1 0 0	0 1 0 1 1 1 0 0	0 1 0 1 1 0 0 0	1 0 0 0 0 0 0 1	1 0 0 1 0 0 0 0	1 0 1 0 0 0 0	1 0 0 1 1 0 0 0
Protists  1 1 0 0 1 0 0 0 0 0 1 0 0 0 0	0 1 1 0 0 0 0 0	1 1 0 1 0 0 0 0	0 1 1 1 0 0 0	0 1 1 1 1 1 1 1 0	0 1 0 1 0 1 0 0	0 1 0 1 1 1 1 0	0 1 0 1 1 1 1 0	0 1 0 0 0 1 1 0	1 0 0 1 0 1 1 0	1 0 1 0 0 0 0	1 0 1 1 0 0 0 0
Scapholebr 0 0 0 0 0 0 0 0 0	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	ngi 0 0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 1 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 0 1 0 0	0 0 0 0 0 0 0	0 0 0 0 0 1 0 0
Scenedesmu 1 0			1 1 1 0	0 0 1 0	1 1 1 0	0 1 1	0 1 1	0 1 0 1	0 1 0 1	0 1 0 1	0 1 0 0

1 0 1 1	1 0 0 0	1 1 0 0	0 1 1 0	0 1 0 1	1 1 0 0	0 1 0 0	1 0 0 1	0 0 0 0	1 0 1 0	1 1 1 0	0 1 1 1
0 Spyrogin 0 0	0 0	1 0 0 0	0 0 0	0 0 0	0 0 0	0 0 0	0 0 0	0 0 0	0 0 0	0 0 0	0 0 0
0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0
0 0 Staurast	0	0 rachiat	0 cum	0	0	0	0	0	0	0	0
1 0 0 1	1 1 0 1	1 0 1 1	1 1 0 0	0 1 0 0	1 1 0 1	1 1 1 0	1 1 1	1 0 1 0	1 0 1 1	1 0 1 1	1 0 0 0
0 1 1	0 0	1 0 0	1 1 0	1 0 1	1 0 0	1 0 0	0 0 1	0 0 0	0 1 0	1 1 0	1 1 1
0 0 Sididae 0	0 1 non-lim 0	0 1 nnetica 0	0 0 a 0 0	0 0	0 0	0 0	0 0	1 0 0	1 0 0	0 0	0 0
0 0 0	0 1 0	0 0 0	0 0 0	0 0 1	0 0 0						
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0 Staurast 1	1	1	1	1 1	0	0	0 1	0 1	0 1	0 1	1 0
1 0 1 0	0 1 1 1	1 0 0 1	1 0 0 1	1 0 1 1	1 1 0 1	1 1 1 1	0 1 0 1	0 1 1 1	0 1 1 1	0 0 0 1	0 1 0 1
0 0 0 1	0 0 0 1	1 0 0 0	0 1 0	0 0 0	0 0 0	1 1 0	0 0 1	1 0 1	1 0 1	1 1 1	1 0 1
Staurast 1 0	eram pol 1 1	ymorph 1 0	1 1	0 1 1	1 1 1	0 1 1	0 1 1	0 1 0	0 1 0	0 1 0	0 1 0
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1	0	0	0	1	0	0	1	0	0	0	1

0	0	0	0	0	0	0	0	1	1	1	1
1	. 1	1	0	-	0	0	0	0	0	0	-
	trum tet			1	0	0	0	0	0	0	1
1 1	1 0	1 1	1 1	1 1	1 1	1 1	1 0	1 0	1 0	1 0	0
0	1	0	0	0	1	1	1	1	1	0	1
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0	0	0	1	0	0	1	0	0	0	1	0
0	0	0	0	0	0	0	1	1	1	1	1
1	1	0	O	O	O	O	_	_	_	_	_
Synedra		0	0	0	0	0	0	0	1	0	0
0	0	0	0	0	0	0	0	0	0	0	0
1	1	1	1	1	0	0	0	0	0	0	0
0	0	0	1	1	1	1	1	0	1	1	1
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	1	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0											
Tetraed:	ron assy	metric	um	0	1	0	0	0	0	0	0
1	1	1	1	1	1	1	1	1	1	1	1
0	1	0	1	1	1	1	1	0	0	0	0
0	0	1	0	0	0	1	1	1	1	1	0
1	1	1	0	0	1	0	1	0	1	1	0
0	0	1	1	1	1	1	1	1	1	1	1
1	0	0	1	0	0	0	1	0	1	1	1
1	0	0	0	1	0	0	1	0	0	0	1
0	0	0	0	0	0	0	0	1	1	1	1
1	1	1	0	-	0	0	0	0	0	0	-
Tetraed:		etica	0	1	0	0	0	0	0	0	1
1	1	1	1	1	1	1	1	1	1	1	0
1	0	1	1	1	1	1	0	0	0	0	0
0 1	1 1	0	0	0 1	1	1 1	1	1 1	1 1	0	1
0	1	1	1	1	1	1	1	1	1	1	1
0	0	1	0	0	0	1	0	1	1	1	1
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0	0	0	0	0	0	0	1	1	1	1	1
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Tetraed:		lare	0	1	0	0	0	0	0	0	1
1	1	1	1	1	1	1	1	1	1	1	0
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0	0	1	0	0	0	1	0	1	1	1	1
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0	0	0	0	0	0	0	1	1	1	1	1
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Trichot			0	1	0	0	0	0	0	0	1
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Thermo	1 cyclo )	1 ps ob 0	0 longat 0	:a 0	0	0	0	0	0	0	0	0
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Tricho				0	0	0	0	0	0	0	0	0
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	)	0	0	0	0	0	0	0	0	0	0	0
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Tricho	cerca )	simi O	lis O	0	1	0	0	0 0	0 0	0	0	0 0
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0	0	1	0	0	0	0	0	1	1	1	1
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Tropocyclo			0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	1	0	0	0	0	0	0	0	0	0	0
0	0	0	0	1	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	1	0	1
0	0	0	0	0	0	0	0	0	0	0	0
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0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	1	0	0	^	0	0	^	0
Tropocyclo		enellis O	0	1 0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	1	0	0	0	1	1	1	1	1	0	0
0	0	0	0	1	0	0	0	0	0	0	0
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0	0	1	0	0	0	0	0	1	1	1	1
0	0	0	0	0	0	1	0	0	0	0	0
0	0	0	0	0	0	0	1	0	0	0	0
1	1	0	Ü	Ü	Ü	Ü	_	Ü	Ü	Ü	Ü
Volvox sp.	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
1	0	0	1	1	0	0	0	0	0	0	0
0	0	0	1	1	1	1	1	0	1	1	1
0	0	0	0	0	0	0	0	0	0	0	0
1	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	0	0	0	0	0	0	0	0
0	0	0	0	1	0	0	0	1	0	0	0
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