

Impaired Tumor-Infiltrating T Cells in Patients with Chronic Obstructive Pulmonary Disease Impact Lung Cancer Response to PD-1 Blockade

Jérôme Biton, Hanane Ouakrim, Agnès Dechartres, Marco Alifano, Audrey Mansuet-Lupo, Han Si, Rebecca Halpin, Todd Creasy, Claudie Bantsimba-Malanda, Jennifer Arrondeau, et al.

▶ To cite this version:

Jérôme Biton, Hanane Ouakrim, Agnès Dechartres, Marco Alifano, Audrey Mansuet-Lupo, et al.. Impaired Tumor-Infiltrating T Cells in Patients with Chronic Obstructive Pulmonary Disease Impact Lung Cancer Response to PD-1 Blockade. American Journal of Respiratory and Critical Care Medicine, 2018, 198 (7), pp.928–940. 10.1164/rccm.201706-1110OC . hal-03892153

HAL Id: hal-03892153 https://hal.sorbonne-universite.fr/hal-03892153v1

Submitted on 15 Mar 2023 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Impaired tumor-infiltrating T cells in patients with COPD impacts lung cancer response to PD-1 blockade

3

Jérôme Biton^{1,2,3}, Hanane Ouakrim^{1,2,3,4,†}, Agnès Dechartres^{8,9,†}, Marco Alifano^{2,5}, Audrey
Mansuet-Lupo^{1,2,3,4}, Han Si¹⁰, Rebecca Halpin¹⁰, Todd Creasy¹⁰, Claudie Bantsimba-Malanda^{1,2,3,10},
Jennifer Arrondeau^{2,6}, François Goldwasser^{2,6}, Pascaline Boudou-Rouquette^{2,6}, Ludovic Fournel^{2,5},
Nicolas Roche⁷, Pierre-Régis Burgel⁷, Jeremy Goc^{1,2,3,11}, Priyanka Devi-Marulkar^{1,2,3}, Claire
Germain^{1,2,3}, Marie-Caroline Dieu-Nosjean^{1,2,3}, Isabelle Cremer^{1,2,3}, Ronald Herbst¹⁰, Diane
Damotte^{1,2,3,4,*}.

10

¹Institut National de la Santé et de la Recherche Médicale (INSERM), UMRS 1138, Cordeliers 11 Research Center, Team Cancer, Immune Control and Escape, Paris, F-75006, France. ²Paris 12 Descartes-Paris 5 University, Paris, F-75006, France. ³Pierre et Marie Curie-Paris 6 University, 13 Paris, F-75005, France. ⁴Department of Pathology, ⁵Department of Thoracic Surgery, 14 ⁶Department of Medical Oncology and ⁷Department of Respiratory and Intensive Care Medicine, 15 Hôpital Cochin, Assistance Publique – Hôpitaux de Paris, Paris, F-75014, France. ⁸Department of 16 clinical epidemiology, Hôtel-Dieu, Assistance Publique – Hôpitaux de Paris, Paris, F-75004, 17 France. ⁹METHODS Team, Center of Research in Epidemiology and Statistics Sorbonne Paris 18 Cité (CRESS), UMR1153, INSERM, Paris, F-75004, France. ¹⁰Oncology Research, MedImmune, 19 LLC, Gaithersburg, Maryland, USA. ¹¹Current address: Joan and Sanford I. Weill Department of 20 Medicine, Division of Gastroenterology and Hepatology, Department of Microbiology and 21 Immunology and The Jill Robert's Institute for Research in Inflammatory Bowel Disease, Weill 22 Cornell Medicine, Cornell University, New York, NY 1002. 23

- 25 ***Corresponding author:** Diane Damotte.
- 26 INSERM UMRS 1138, Cordeliers Research Center, Team Cancer, Immune Control and Escape.
- 27 15 rue de l'école de Médecine, F-75006, Paris, France.
- 28 Phone: +33-1-44-27-90-86
- 29 Fax: +33-1-44-27-81-17
- 30 E-mail: <u>diane.damotte@aphp.fr</u>
- 31 [†]These authors contributed equally to this work.
- 32

Author contributions: D.D and R.H designed and supervised the study. J.B, H.O, J.G, H.K, P.D-M and C.G acquired immunohistochemical data. J.B acquired flow cytometry data. H.S, Reb.H and T.C acquired and analyzed WES experiments. J.B, H.O, C.B-M, C.G, R.H and D.D analyzed the data. J.B and A.D performed statistical analysis. H.O, A.M-L, M-A and D-D were responsible for clinical data. A.M-L and D.D were responsible for pathological data. J.B, R.H and D.D interpreted data. J.B, D.D and R.H wrote the manuscript. M.A, H.O, N.R, P-R.B, C.G, I.C and M-C.D-N revised the manuscript.

40

Footnotes: This work was supported by the Institut National de la Santé et de la Recherche Médicale (INSERM), Paris Descartes-Paris 5 University, Pierre et Marie Curie-Paris 6 University, the Cancer Research for Personalized Medicine (CARPEM), the LabEx Immunooncology, the Institut National du Cancer (2011-PLBIO-06-INSERM 6-1) and MedImmune.

45

46 **Running title:** COPD disrupts NSCLC tumor immune contexture.

47 Descriptor number: Lung diseases/9.26 Lung cancer: cellular and molecular aspects; Lung
48 diseases/9.09 COPD: General; Immunology and inflammation/7.15 Lymphocytes.

49 **Total word count for the body of the manuscript:** 4689 words.

50 At the Glance Commentary:

51 <u>Scientific knowledge on the subject</u>: The immune system is strongly involved in the 52 establishment of chronic inflammation in chronic obstructive pulmonary disease (COPD) and in 53 the control of tumor burden in lung cancer. However, despite the strong epidemiological link 54 between these two diseases, the impact of COPD associated chronic inflammation on the immune 55 contexture of lung cancer remains poorly defined.

What this study adds to the field: Here, we report that COPD disrupts the immune 56 microenvironment of non-small cell lung cancer (NSCLC), and we identify CD8 tumor 57 infiltrating lymphocytes (CD8 TILs) as the most affected population. Indeed, we observed higher 58 exhaustion of CD8 TILs, identified by PD-1/TIM-3 coexpression, in NSCLC patients with 59 coexisting moderate to severe COPD. In agreement, the prognostic value of intra-tumor CD8⁺ T 60 cells that has been found favorable in most cancer types and particularly in NSCLC, has no 61 impact on the survival of patients with coexisting COPD. Together, our data point out COPD 62 patients as a potential NSCLC patient population to treat with immune-checkpoint blockers. In 63 accordance with this hypothesis, data obtained in a cohort of 39 nivolumab treated patients might 64 suggest a higher efficacy of anti-PD-1 treatment in NSCLC patients with a coexisting COPD. 65

66

67 This article has an online data supplement, which is accessible from this issue's table of content68 online at www.atsjournals.org.

69 Abstract

Rationale: Patients with chronic obstructive pulmonary disease (COPD) have a higher prevalence of lung cancer. The chronic inflammation associated with COPD probably promotes the earliest stages of carcinogenesis. However, once tumors have progressed to malignancy, the impact of COPD on the tumor immune microenvironment remains poorly defined, and its effects on immune-checkpoint blockers' efficacy are still unknown.

75 **Objectives:** To study the impact of COPD on the immune contexture of non-small cell lung 76 cancer (NSCLC).

Methods: We performed in depth immune profiling of lung tumors by immunohistochemistry and we determined its impact on patients' survival (n=435). Tumor-infiltrating T lymphocyte (TILs) exhaustion by flow cytometry (n=50) was also investigated. The effectiveness of an anti-PD-1 treatment (nivolumab) was evaluated in 39 advanced-stage NSCLC patients. All data were analyzed according to patients' COPD status.

Measurments and Main Results: Remarkably, COPD severity is positively correlated with the 82 coexpression of PD-1/TIM-3 by CD8 T cells. In agreement, we observed a loss of CD8 T cell-83 associated favorable clinical outcome in COPD⁺ patients. Interestingly, a negative prognostic 84 value of PD-L1 expression by tumor cells was observed only in highly CD8 T cell-infiltrated 85 86 tumors of COPD⁺ patients. Finally, data obtained on 39 advanced-stage NSCLC patients treated by an anti-PD-1 antibody showed longer progression free survival in COPD⁺ patients, and also 87 that the association between the severity of smoking and the response to nivolumab was 88 preferentially observed in COPD⁺ patients. 89

90 Conclusions: COPD is associated with an increased sensitivity of CD8 TILs to immune escape
91 mechanisms developed by tumors, thus suggesting a higher sensitivity to PD-1 blockade in
92 patients with COPD.

93	Keywords: NSCLC, tumor immunology, CD8 TILs, anti-PD-1.
94	
95	Total word count for the abstract: 250 words
96	
97	
98	
99	
100	
101	
102	
103	
104	
105	
106	
107	
108	
109	
110	
111	
112	
113	
114	
115	
116	
3 4	

117 Introduction

Despite abundant evidence that the immune system plays a central role in controlling 118 tumor burden (1-3), it may also have a dark side linked to the maintenance of deleterious 119 inflammation. For instance, patients with inflammatory bowel disease (IBD) (4) or chronic 120 pancreatitis (5) showed increased risk of developing colorectal and pancreatic cancer, 121 respectively. Similarly, chronic obstructive pulmonary disease (COPD) is considered to be an 122 123 important risk factor for lung cancer (6, 7). This inflammatory condition is linked to a more pronounced destructive inflammation of the lung, compared with non-COPD smokers, 124 characterized by a strong release of TNF- α and CXCL-8 by epithelial cells and alveolar 125 126 macrophages leading to the recruitment of inflammatory monocytes and neutrophils (8). The presence of B cells in lymphoid follicles has been reported in the airways and parenchyma of 127 patients with COPD (9), illustrating the involvement of adaptive immunity in COPD 128 pathophysiology. This chronic inflammation may promote the earliest stages of carcinogenesis 129 (8) through an increased expression of genes involved in cell proliferation and survival, including 130 NF- κ B and STAT3, which are activated by cytokines such as IL-6 and TNF- α . 131

Once tumors have progressed to malignancy, COPD was shown to worsen the survival of 132 patients with early-stage non-small cell lung cancer (NSCLC) (10) and emphysema was shown to 133 134 be associated with increased lung cancer mortality (11). Mechanisms governing this prognostic impact, including the role of the immune system, are currently undefined. Although the tumor 135 immune contexture in NSCLC has been extensively characterized, the COPD status of patients 136 has not been taken into account. Nevertheless, a high density of CD8 tumor-infiltrating T 137 lymphocytes (CD8 TILs), together with a concomitant high density of DC-Lamp⁺ cells that 138 signals the presence of tertiary lymphoid structures (TLS) within tumor tissues, identified patients 139 with the best prognostic outcome (12). However, overexpression of inhibitory receptors by 140

tumor-infiltrating T cells, also called immune-checkpoints, can keep the immune system under 141 control (13). In NSCLC, their cumulative expression, including programmed cell death-1 (PD-1), 142 T-cell immunoglobulin and mucin domain-containing molecule-3 (TIM-3), cytotoxic T 143 lymphocyte-associated antigen-4 (CTLA-4) and lymphocyte activation gene-3 (LAG-3), has been 144 described as being a hallmark of dysfunctional T cells and tumor progression (14). Drugs 145 targeting immune-checkpoints, in particular the PD-1/PD-L1 pathway, can unleash anti-tumor 146 147 immunity and mediate durable cancer regression (15-17). Nevertheless, these new treatments are not efficient in all patients and identifying factors that predict clinical response to these therapies 148 remains a challenge. In melanoma, an association between high PD-L1 expression and clinical 149 150 response to pembrolizumab had been reported (18). However, patients with PD-L1-negative tumors may also achieve durable responses. In NSCLC, several efforts have also been made 151 recently to more accurately identify patients that would respond to checkpoint therapy. The focus 152 here has largely been on the identification of predictive markers for response to anti-PD-1, such 153 as tumor mutational burden (TMB) (19), PD-L1 expression by tumor cells (20, 21), and gene 154 155 signature reflecting adaptive immunity (22).

In this context, the present study investigated the potential impact of COPD on the immune microenvironment of NSCLC and, thus, on patients' outcome. Our work reveals that COPD severity is positively correlated with the level of CD8 TIL exhaustion. In agreement, we observed a complete loss of CD8 T cell-associated favorable clinical outcome in COPD⁺ patients. Finally, data obtained on 39 advanced-stage NSCLC patients treated by an anti-PD-1 might suggest a higher sensitivity to this treatment in patients with COPD.

- 162
- 163
- 164
 - 7 8

165 Methods

166 Patients

A retrospective consecutive cohort of 435 NSCLC untreated patients seen between June 167 2001 and December 2005 at the department of Thoracic Surgery of Hôtel-Dieu hospital (Paris, 168 France) was used to study by immunohistochemistry the immune composition of the tumor 169 microenvironment. A second cohort of fresh tumor samples, distant non-tumoral lung samples 170 171 and peripheral blood were obtained from 50 patients with untreated NSCLC who underwent surgery between March 2014 and December 2015. These samples were used to perform flow 172 cytometry experiments. A third cohort of 39 patients with advanced-stage NSCLC receiving an 173 anti-PD-1 antibody (nivolumab) was used to assess the effectiveness of this treatment according 174 to COPD status. Additional details are provided in the online supplements. 175

176

177 COPD assessment

The Global Initiative for Chronic Obstructive Lung Disease (GOLD) criteria were used to assess the presence of COPD and to evaluate the severity of airflow obstruction (23). Additional details are provided in the online supplements.

181

182 Immunohistochemistry

183 Serial section of paraffin-embedded NSCLC tumors were stained as previously described184 (24). Additional details are provided in the online supplements.

185

186 Methods for cell Quantification

187 Calopix software (Tribvn) was used to count CD66b⁺ and CD68⁺ cells in the whole tumor
 188 section; CD8 T cells were counted separately in the tumor nests and in tumor stroma. DC-Lamp⁺
 9 5

cells were counted manually in the whole tumor section. The proportion of PD-L1⁺ cells among tumor cells was determined manually by at least two independent observers (JB, HO or DD). The positivity threshold was fixed at \geq 1%. Additional details are provided in the online supplements (Table E1).

193

194 Flow cytometry

Multiple stainings on isolated mononuclear cells from Tumor, NT and Blood were performed using various antibodies (see Table E2 and Figure E1), as previously described (24) Additional details are provided in the online supplements.

198

199 Genomic DNA extraction and Illumina-based whole-exome sequencing.

Genomic DNA from tumors was isolated from formalin-fixed paraffin-embedded blocks using Maxwell® 16 FFPE Tissue LEV DNA Purification Kit (Promega), according to the manufacturer's instructions. DNA whole exome sequence (WES) data were sequenced on the Novaseq 6000 platform (25). Additional details are provided in the online supplements.

204

205 Statistics

Categorical data were compared using Chi-square tests or Fisher exact tests, as appropriate, while they were compared according to COPD stages using exact Cochran-Armitage trend test. For log-rank tests, the prognostic value of continuous variables was assessed by a quartile stratification. For Cox proportional-hazard models, immune cell densities were logtransformed. Multivariate analysis for OS was adjusted for age, gender, vascular emboli, smoking history and stratified on the tumor stage. In flow cytometry experiments, according to data distribution, a parametric test (ANOVA, student's t test) or a non-parametric test (Kruskal-Wallis, Mann-Whitney), with appropriate post-hoc comparisons, was used to compare quantitative variables across the different groups. Correlations between quantitative parameters were assessed by using the Spearman test. Additional details are provided in the online supplements. Results

236 COPD does not affect immune cell density in NSCLC tumor microenvironment.

We first investigated the impact of COPD on the composition of the tumor immune 237 microenvironment in a retrospective cohort of 435 NSCLC patients. Among them, 45% had 238 COPD, and in the COPD⁺ group, 29% had a COPD GOLD stage I (COPD⁺ I), 60% a COPD 239 GOLD stage II (COPD⁺II) and 11% a COPD GOLD stage III (COPD⁺III) (Table 1 and E3). The 240 mean age, the percentages of male and of smokers were higher for COPD⁺ than COPD⁻ patients 241 242 (Table 1). Coexisting COPD was associated with significant worse survival only for NSCLC stage I patients (Figure E2). Because of the small number of COPD⁺ III patients, the COPD⁺ II 243 and COPD⁺ III groups were merged for most of the subsequent analyses. COPD⁺ and COPD⁻ 244 patients did not differ in density of neutrophils (CD66b⁺ cells), macrophages (CD68⁺ cells), 245 mature DCs (DC-Lamp⁺ cells) and CD8 T cells in tumor nests (CD 8_{Tu}) and in stroma (CD 8_s), 246 regardless of GOLD stage (Figure E3). 247

248

Absence of immune cell prognostic value in NSCLC patients with moderate to severe COPD

To indirectly investigate whether the functionality of immune cells in the tumor 251 microenvironment could be modified by coexisting COPD, we first determined their impact on 252 253 patient survival according to COPD status and GOLD stage (Figure 1) (Table E4 and E5). In the whole retrospective cohort and in the COPD⁻ group, univariate Cox-regression analysis showed 254 that CD8_{Tu}, CD8s and DC-Lamp⁺ cell densities were all associated with favorable prognostic 255 256 value, while neutrophil and macrophage density had no impact on patient survival (Figure 1A and B). Strikingly, $CD8_{Tu}$ cell density did not affect patient survival in COPD⁺ patients (Figure 1C). 257 Furthermore, CD8_{Tu}, CD8_s and DC-Lamp⁺ cell densities were not significantly associated with 258 improved survival in COPD⁺ II-III patients (Figure 1D). 259

260 Then, patients were stratified by quartiles of $CD8_{Tu}$ cell density. In COPD⁻ patients, $CD8_{Tu}$ cell density was associated with longer overall survival (OS) as soon as the 2nd quartile was 261 reached (Figure 1E), whereas in COPD⁺ and COPD⁺ II-III patients the survival curves for all 262 quartiles merged together (Figure 1F and G). DC-Lamp⁺ cell and CD8_s cell densities were not 263 associated with significant prognostic value in COPD⁺ II-III patients (Figure E4). Multivariate 264 Cox-regression analysis adjusted for age, gender, vascular emboli, smoking history and stratified 265 266 on tumor stages, highlighted the absence of CD8 T cell prognostic value for the stroma and tumor nests in COPD⁺ patients (Figure 1H-J). Together, these data might suggest that the protective 267 impact of a high adaptive immune cell infiltration in NSCLC is altered in COPD⁺ patients and 268 identify $CD8_{Tu}$ cells as the most affected population. 269

- 270
- 271

272 TIL exhaustion, identified by PD-1/TIM-3 co-expression, is correlated with COPD severity

Based on above results, we investigated whether effector functions of TILs were altered 273 in COPD⁺ patients using a prospective cohort of 50 NSCLC patients (Figure 2 and E5) (Table E6 274 and E7). Regardless of COPD status, within the tumor tissue (Tumor), the proportion of CTLA-275 4⁺, LAG-3⁺, PD-1⁺ and TIM-3⁺ cells among CD4 (Figure E5A) and CD8 T cells (Figure 2A and 276 277 B) was consistently higher than in the other anatomical sites (blood and non-tumor distal lung tissue (NT)). A marked increased of CD4⁺ FoxP3⁺ regulatory T cells (Treg) frequency among 278 279 total CD4 T cells in Tumor was also observed (Figure E6A and B). Regarding cytokine secretion, the frequency of CD4 (Figure E5B) and CD8 T cells (Figure 2C and D) positive for Granzyme B, 280 TNF- α , IFN- γ and IL-17 was lower in Tumor than in NT. As shown by the correlation matrix 281 exposed on Figure 2E, among CD8 TILs, PD-1 and TIM-3 expression was strongly positively 282 correlated, as was the frequency of IFN- γ^+ and TNF- α^+ cells. Remarkably, CD8 TILs co-283

expressing PD-1 and TIM-3 were restricted to Tumor (Figure 2F), and for this cell subset only, there was a significant inverse correlation with both IFN- γ and TNF- α secretion (Figure 2G). Overall, similar results were observed regarding CD4 TILs (Figure E5C and D). However, less CD4 TILs coexpressed PD-1/TIM-3 (Figure E5D), and the relationship between cytokine secretion and PD-1/TIM-3 coexpression was weaker (Figure E5C and E).

Based on above results, we investigated the link between COPD and TIL exhaustion. In 289 290 COPD, airflow obstruction severity is inversely correlated with the Forced Expiratory Volume in 1 second expressed as a percentage of normal predicted values (FEV1% predicted) (see Method 291 292 section). Remarkably, FEV1% predicted was inversely correlated with the proportion of CD8 293 TILs expressing PD-1 and co-expressing PD-1/TIM-3 in COPD⁺ patients only (Figure 3A). In agreement, in COPD⁺ patients, FEV1% predicted was positively correlated with the proportion of 294 CD8 TILs secreting TNF- α and IFN- γ (Figure 3A). Regarding CD4 TILs, only IFN- γ was 295 positively correlated with FEV1% predicted (Figure E7A). Interestingly, frequency of TIM-3⁺, 296 PD-1⁺ and TIM-3⁺/PD-1⁺ cells among CD4 and CD8 TILs was higher in COPD⁺ II-III patients 297 than in COPD⁻ patients (Figure 3B) (Figure E7B). The proportion of Treg among CD4 TILs was 298 not different according to patients' COPD status (Figure E7C). Overall, these results demonstrate 299 that COPD severity is strongly correlated with TIL exhaustion, and that this association is more 300 301 pronounced for CD8 TILs.

302

303 Strong correlation between CD8_{Tu} cell density and CD8 TIL exhaustion: a phenomenon 304 exacerbated in COPD⁺ patients

An association between CD8 TIL exhaustion (PD-1⁺ cell frequency) and the immune composition of the tumor microenvironment (density of CD8⁺ T cells) was recently reported in colorectal cancer (26). We investigated this interrelation in our prospective cohort and then

studied the impact of COPD. First, CD8 TIL exhaustion (based on PD-1 and TIM-3 expression) 308 and cytokine secretion were only linked to CD8_{Tu} cell and CD8_s cell densities (Figure E8A). 309 Regarding CD4 TILs, none of the immune cell densities studied was associated with their 310 exhaustion, and only their cytokine secretion was slightly inversely correlated with CD8s cell 311 density (Figure E7D). Due to the strong association between CD8 TIL exhaustion and their 312 density in the tumor nests, we then focused our analysis on CD8 TILs. Importantly, CD8_{Tu} cell 313 314 density and CD8 TIL exhaustion were more strongly associated in COPD⁺ patients than in COPD⁻ patients (Figure 3C) and (Figure E8B and C). 315

To confirm these results, median $CD8_{Tu}$ cell density was used to separate patients into two groups according to a Low (Figure 3D) or a High (Figure 3E) $CD8_{Tu}$ cell density. In the $CD8_{Tu}^{Low}$ group, the level of CD8 TIL exhaustion did not differ according to COPD status (Figure 3D). In the $CD8_{Tu}^{High}$ group, the frequencies of CD8 TILs expressing TIM-3 and co-expressing PD-1/TIM-3 were significantly higher in COPD⁺ patients than in COPD⁻ patients (Figure 3E). Overall, CD8 TIL exhaustion was restricted to highly CD8 T cell infiltrated tumors and this phenomenon was exacerbated in COPD⁺ patients.

323

324

PD-L1 expression by malignant cells is associated with shorter survival only in CD8_{Tu}^{High} COPD⁺ patients

The strong impact of immunosuppression on tumor burden is based on TIL exhaustion, but also on concomitant mechanisms that malignant cells develop to avoid immune surveillance. The most-studied mechanism is probably PD-L1 expression by malignant cells (Figure E9A-D). No difference of PD-L1 expression by tumor cells was observed according to patients' COPD status and Gold stages (retrospective cohort) (Figure E9F). Consistent with previous studies, we

found that high CD8 T cell density is associated with high PD-L1 expression by tumor cells (Figure E9G) (27, 28). The frequency of tumor cells expressing PD-L1 was also higher, but to a lesser extent, in neutrophil^{High}, macrophage^{High} and DC-Lamp^{High} groups. Additionally, among these highly infiltrated groups, PD-L1 expression was similar between COPD⁻ and COPD⁺ patients (Figure E9G).

We then investigated whether coexisting COPD modified the prognostic value of PD-L1 337 338 expression by tumor cells. Whatever the group of patients considered, PD-L1 expression was not associated with significant prognostic value (Figure 4A and B). Since PD-L1 expression was 339 strongly linked to CD8 T cell density (Figure E9G), we then deciphered the prognostic value of 340 PD-L1 according to a High/Low CD8_{Tu} cell density, and also to patient COPD status. For 341 CD8_{Tu}^{Low} groups, PD-L1 expression was not associated with significant prognostic value in 342 COPD⁻ or in COPD⁺ patients (Figure 4C and D). Interestingly, for CD8_{Tu}^{High} groups, PD-L1 343 expression did not affect survival for COPD⁻ patients (Figure 4E), but was associated with a 344 reduced OS for COPD⁺ patients (Figure 4F). Moreover, in COPD⁻ patients, the prognostic value 345 of CD8_{Tu} and of CD8_s cell density was similar whether tumor cells expressed PD-L1 or not 346 (Figure 4G). Remarkably, for COPD⁺ patients, CD8_{Tu} and CD8_s cell densities were associated 347 with extended OS for those with PD-L1⁻ tumors, while these prognostic values were not observed 348 349 in the PD-L1 $^+$ groups (Figure 4H). Finally, these results were confirmed in subgroups of patients defined by a cut-off of PD-L1⁺ tumor cell frequency $\geq 5\%$ (Figure E10A and B) and $\geq 10\%$ 350 (Figure E10C and D). 351

352

Anti-PD-1 antibody (nivolumab) efficacy in advanced-stage NSCLC patients according to
 coexisting COPD and to smoke exposure

We investigated the impact of COPD on the effectiveness of an anti-PD-1 antibody from a 355 cohort of 39 patients with advanced-stage NSCLC receiving nivolumab (Table E8). The 356 percentage of smokers and the number of pack-years were higher for COPD⁺ than COPD⁻ 357 patients, while no significant differences were observed between the two groups of patients 358 regarding NSCLC subtypes, the duration of follow-up, the age and the gender (Table E8). At the 359 completion of the study, a significantly longer progression-free survival (PFS) (Figure 5A and B) 360 361 and a higher percentage of patients still alive (Table E8 and E9) (Figure 5C) were observed in the COPD⁺ group. However, we did not observe any impact of COPD severity, assessed using the 362 FEV1 % predicted, on nivolumab efficacy (Table E9). 363

364 In addition, it was previously shown that the efficacy of pembrolizumab, another anti-PD-1 antibody, was greater in patients with a smoking-associated mutational signature or with a 365 higher nonsynonymous mutation burden in tumor (19). Consequently, we investigated whether 366 the increased PFS seen with nivolumab in COPD⁺ patients was linked to the higher smoke 367 exposure observed in this group (Table E8). In the whole cohort, a smoke exposure >30 pack-368 years was associated with a better PFS and OS (Table E9), while in non-COPD patients, smoke 369 exposure >30 pack-years was not associated significantly with a better PFS or OS (Table E9) 370 (Figure 5D and F). Remarkably, in the COPD⁺ group, a smoke exposure >30 pack-years was 371 372 associated with a better PFS and also with a dramatic improvement of OS (Table E9) (Figure 5E and G). Regarding PFS, these results were confirmed when smoke exposure was assessed using 373 number of pack-years as a continuous variable (Table E9). 374

A strong relationship between tobacco smoke exposure and the number of somatic mutations was previously reported in NSCLC (29). Consequently, we tried to investigate whether the number of nonsynonymous mutations per megabase (TMB) differ according to patients'COPD status, by performing whole exome sequencing (WES) experiments. Among the

22 patients for whom enough DNA was available (Figure 5C), we did not observe any difference 379 regarding the TMB in COPD⁺ patients compared to non-COPD patients (Figure E11A). 380 Remarkably, the number of pack-years was significantly correlated with the TMB only in non-381 COPD patients (Figure E11C-E). Additionally, it has been shown that TP53 and/or KRAS-382 mutated tumors, two mutations strongly associated with tobacco smoke exposure (30, 31), had a 383 better response to PD-1 blockade (32). Among the 31 patients who had a molecular interrogation 384 385 of their tumor before starting anti-PD-1 treatment (Figure 5C), using next-generation sequencing (NGS), we did not detect a differential distribution of TP53 and/or KRAS-mutated tumors 386 according to patients' COPD status (Figure E11B). 387

388 Our preliminary data might suggest a differential link between smoking history and response to nivolumab, but also between TMB and smoking history, in COPD⁺ patients vs non-389 COPD patients. Consequently, we investigated whether COPD and tobacco had a synergistic 390 impact on CD8 TIL exhaustion and on immune cell prognostic value. First, in the prospective 391 cohort, the number of pack-years was positively correlated with the proportion of CD8 TILs co-392 expressing PD-1/TIM-3 in COPD⁺ patients only (Figure E12A). Moreover, a higher CD8 TILs 393 exhaustion was observed in COPD⁺ group compared with COPD⁻ group, in patients with a 394 number of pack-years > 60 (Figure E12B). Secondly, in our retrospective cohort of 435 NSCLC 395 396 patients, we investigated whether immune cell prognostic value (CD8Tu, CD8S and DC-Lamp⁺ cells) was impacted by a strong smoke exposure (>30 pack-years). In heavy smokers, immune 397 cell prognostic value was stronger in non-COPD patients (Figure E12C), than the one observed in 398 Figure 1B. Conversely, in COPD⁺ patients, the CD8_s cell prognostic value was not significant in 399 heavy smokers, and was completely absent for $CD8_{Tu}$ cells (HR:1.01, p=0.948) (Figure E12D). 400 Altogether, NSCLC patients with COPD, a group characterized by a complete loss of CD8 T cell-401

- 402 associated favorable clinical outcome in heavy smokers probably due to their marked exhaustion,
- 403 also showed a longer PFS after nivolumab treatment.

406 Discussion

Our main objective was to evaluate the potential impact of COPD on the immune 407 contexture of NSCLC. First, immune cell densities did not differ according to the COPD status of 408 the patients. Immune cell recruitment into the malignant lesion is probably driven by tumor cells 409 according to their immunogenicity linked to their mutational burden, thereby attenuating the 410 impact of coexisting COPD. Co-occurring genetic alterations in KRAS-mutant lung 411 412 adenocarcinoma, were associated with different tumor immune patterns, and could be a first argument supporting this hypothesis (33). However, our study showed higher TIL exhaustion in 413 the COPD⁺ II-III group, an impaired protective impact of immune cells in patients with COPD, 414 and identified CD8 TILs as the most affected population. 415

The characteristics of the NSCLC immune contexture linked to CD8 TIL exhaustion are 416 not completely defined, and the role of COPD in this phenomenon is not completely elucidated. 417 We identified that CD8 TIL exhaustion was restricted to highly CD8 T cell infiltrated tumors, 418 and these findings were exacerbated in COPD⁺ patients. Interestingly, PD-1 expression, which is 419 upregulated on T cells after TCR ligation (34), is also upregulated in activated T cells by IL-6 420 through STAT3-dependent mechanisms (35). Accordingly, high frequency of exhausted TILs 421 observed in COPD⁺ II-III patients could be driven in part by the increased amounts of IL-6 422 423 previously reported in the sputum of COPD⁺ patients (36). It is also conceivable that when a tumor forms near an emphysematous/inflammatory lesion, the surrounding inflammation, 424 including numerous cytokines (IL-6, TNF- α , IL-1 β) and chemokines (CXCL8, CXCL1) (37) 425 modify autocrine and paracrine interactions between malignant cells and infiltrating leucocytes. 426 Interestingly, ex vivo infections with influenza virus of lung resections showed an impaired 427 antiviral function of CD8⁺ T cells in COPD⁺ patients compared to non-COPD patients, through an 428 429 up-regulation of PD-1 expression. Moreover, if these PD-1 expressing CD8 T cells, coming from

the non-tumor distal lung tissue, are overrepresented in the tumor immune microenvironment of COPD⁺ patients, they might participate in the deviation of the anti-tumor immune response observed in patients with COPD (38). Nevertheless, orthogonal approaches, including gene expression analysis related to the immune response in cancer, are required to precisely identify the characteristics of the inflammation disrupting the tumor immune contexture of patients with NSCLC and COPD.

436 In agreement with other studies (17, 27, 39), we observed that PD-L1 expression by NSCLC tumor cells highlights the presence of an active tumor immune microenvironment in lung 437 cancer, independently of COPD. This is probably due to the fact that cancer cells may upregulate 438 PD-L1 expression in response to IFN- γ secretion by TILs (39). Studies of the prognostic value of 439 PD-L1 expression in patients with NSCLC has yielded inconsistent data (40-44). These 440 conflicting results could be due to the fact that the amount of CD8 TILs and patients' COPD 441 status had not been taken into account. In our study, the prognostic value of PD-L1 expression 442 was restricted to COPD⁺ patients belonging to the CD8_{Tu}^{High} group. It probably reflects the 443 effectiveness of mechanisms that cancer cells develop to avoid immune surveillance in a 444 subpopulation of patients characterized by a strong CD8 TIL exhaustion. In CD8_{Tu}^{Low}, the lack of 445 PD-L1 prognostic value was probably linked to the lowest impact of the PD-1/PD-L1 pathway on 446 447 a weakly active anti-tumor immune response. In this situation, PD-L1 expression is probably driven more through oncogenic pathways, including inactivation of STK11/LKB1 (45) or loss of 448 function of the tumor suppressor PTEN (46), and not associated to a strong PD-1 expression by 449 CD8 TILs. Moreover, immune cell prognostic value was completely absent for tumors from 450 COPD⁺ patients expressing PD-L1, probably because the level of TIL exhaustion was higher in 451 this group. Interestingly, in melanoma, preexisting CD8 TILs in tumor microenvironment were 452 453 required for tumor regression after treatment with pembrolizumab (27), and CD8 TILs

454 coexpressing PD-1/CTLA-4 had been proposed as a biomarker to predict response to anti-PD-1
455 (47). Altogether, for NSCLC patients with moderate to severe COPD, our results support an
456 increased predictive potential of PD-L1 expression by tumor cells for the response to checkpoint
457 inhibitors targeting the PD-1/PD-L1 pathway (48).

In accordance with this assumption, our preliminary data obtained in a cohort of 39 458 nivolumab treated NSCLC patients showed a longer PFS in patients with coexisting COPD (19). 459 460 Nevertheless, we did not identify any impact of COPD severity on the response to anti-PD-1. It is conceivable that we had not enough patients to observe this kind of effect. Moreover, other 461 462 confounding factors not explored in our work could explain the lack of association between COPD severity and the response to nivolumab, including among others, CD8 TIL density and 463 PD-L1 expression by tumor cells. Remarkably, at the time of publication, the study from Mark et 464 al, also observed that the presence of COPD was associated with longer progression free survival 465 interval in patients treated with anti-PD-1 or anti-PD-L1 (49). However, a smoking-associated 466 mutational signature had previously been suggested to signal a better response to immunotherapy. 467 Since, tobacco and COPD may be confounding factors in the response to nivolumab, we tried to 468 investigate the interplay between these two factors, and if possible their respective impact. Our 469 study suggests that the impact of tobacco on the response to nivolumab would be mainly 470 471 observed in COPD⁺ patients. In agreement with this assumption, when anti-PD1/PD-L1 impact was evaluated only in former smokers, Mark et al still observed a longer PFS and OS in COPD⁺ 472 patients (49). We also showed a stronger impact on patients' survival of the immune 473 microenvironment in heavy smokers without COPD. In this situation, tumors are probably more 474 immunogenic and the presence of a strong specific adaptive anti-tumor immune response even 475 more important for patients' survival. Interestingly, in COPD⁺ patients, the number of pack-years 476 was positively correlated with the level of CD8 TIL exhaustion, and the impact of CD8_{Tu} cell 477

density on patients' survival was completely absent in heavy smokers. Altogether, the higher
nivolumab efficacy observed in COPD⁺ patients, probably reflects the effectiveness of PD-1
blockers to unleash anti-tumor CD8 T cell response in a subpopulation of patients characterized
by strong CD8 TIL exhaustion.

Finally, we tried to investigated whether the longer PFS observed in nivolumab COPD⁺ 482 treated patients was linked to a higher TMB induced by a stronger tobacco smoke exposure. 483 484 Unfortunately, we were able to perform such work only on 22 anti-PD-1 treated patients. However, our preliminary data are not in favor of a higher TMB in COPD⁺ patients. Moreover, 485 the frequency of KRAS and TP53 mutations, two mutations strongly linked to tobacco smoke 486 exposure (30, 31) that have been suggested as being associated with longer PFS in anti-PD-1 487 treated patients (32) were not enriched in COPD⁺ patients. In agreement with these results, our 488 previous work did not detect a higher frequency of TP53 or KRAS mutations in COPD⁺ patients, 489 again characterized by a stronger smoke exposure, in a cohort of 282 lung adenocarcinomas (31). 490 Beyond the scope of the present work, studies using larger cohorts of patients are mandatory to 491 precisely determine whether the longer PFS after PD-1 blockade observed in COPD⁺ patients is 492 mostly linked or not to their higher tobacco smoke exposure. Such studies will also allow to 493 determine whether tobacco smoke exposure differentially impacts CD8 TIL exhaustion, TMB 494 495 and response to PD-1 blockers, in COPD⁺ patients vs non-COPD patients.

However, there were some limitations to our study. First, our cohort of anti-PD-1 treated patients is restricted to 39 patients, mainly because we needed a follow up of at least one year, combined to fully characterized respiratory functions. Secondly, the use of three cohorts inevitably increased the number of comparisons and the number of false discovery rate, but we tried to reduce this risk using the most appropriate statistical methodology and by applying appropriate adjustments for multiple comparisons. The impact of histological subtypes on our

502 findings was not fully analyzed, even it does not seem to impact our main results (Data not 503 shown). In fact, we did not address this point to avoid an increase of multiple comparisons. 504 Another limitation, inherent to this kind of work, is related to tissue heterogeneity, slide thickness 505 and surface area covered. However, one strength of our study is that in immunochemistry 506 experiments, we worked on whole tumor sections and not on tumor microarrays (TMA), with 507 senior pathologists, using automates for staining, scanning and counting. All these precautions 508 were intended to increase the accuracy and reproducibility of our data.

By deciphering the immune network of NSCLC, we pinpointed that the use of 509 immunological biomarkers to evaluate patient prognosis (50) and to predict the response to 510 therapy should definitively take into account the coexistence of COPD. This chronic 511 inflammatory disease of the lung is not fully understood and its impact on tumor immune 512 microenvironment functions and on response to immune checkpoint inhibitors will deserve 513 additional experiments. We recommend that clinical trials should investigate whether CD8 TIL 514 density and smoke exposure, together with coexisting COPD may identified the best responders 515 to therapies targeting immune-checkpoints. Overall, our study emphasizes the need to consider 516 the impact of coexisting chronic inflammation on the tumor immune microenvironment in other 517 cancer types. In the era of precision medicine, such studies should extend the clinical success of 518 519 immunotherapies in cancer.

520

Acknowledgments: We thank Patricia Bonjour, Béatrice Marmey (Department of Pathology,
Hôpital Cochin, Paris), Sarah Leseurre (Department of Thoracic Surgery, Hôpital Cochin, Paris),
Nathalie Jupiter and Samantha Knockaert (UMRS 1138, Cordeliers Research Center, Team 13,
Paris) for technical assistance.

- 525
- 39 40

526		
527		
528		
529		
530		
531		
532		
533		
534		
535		
536		
537		
538		
539		
540		
541		
542		
543		
544		
545		
546		
547		
548		
549		
550		
551	Re	ferences
552		
553	1.	Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pagès C, Tosolini M,
554		Camus M, Berger A, Wind P, Zinzindohoué F, Bruneval P, Cugnenc P-H, Trajanoski Z,

555		Fridman W-H, Pagès F. Type, density, and location of immune cells within human colorectal
556		tumors predict clinical outcome. Science 2006;313:1960–1964.
557	2.	Azimi F, Scolyer RA, Rumcheva P, Moncrieff M, Murali R, McCarthy SW, Saw RP,
558		Thompson JF. Tumor-infiltrating lymphocyte grade is an independent predictor of sentinel
559		lymph node status and survival in patients with cutaneous melanoma. J Clin Oncol
560		2012;30:2678–2683.
561	3.	Sharma P, Shen Y, Wen S, Yamada S, Jungbluth AA, Gnjatic S, Bajorin DF, Reuter VE,
562		Herr H, Old LJ, Sato E. CD8 tumor-infiltrating lymphocytes are predictive of survival in
563		muscle-invasive urothelial carcinoma. Proc Natl Acad Sci USA 2007;104:3967-3972.
564	4.	Kim ER, Chang DK. Colorectal cancer in inflammatory bowel disease: the risk,
565		pathogenesis, prevention and diagnosis. World J Gastroenterol 2014;20:9872-9881.
566	5.	Pinho AV, Chantrill L, Rooman I. Chronic pancreatitis: A path to pancreatic cancer. Cancer
567		Letters 2014;345:203–209.
568	6.	de Torres JP, Marín JM, Casanova C, Cote C, Carrizo S, Cordoba-Lanus E, Baz-Dávila R,
569		Zulueta JJ, Aguirre-Jaime A, Saetta M, Cosio MG, Celli BR. Lung cancer in patients with
570		chronic obstructive pulmonary disease incidence and predicting factors. Am J Respir Crit
571		<i>Care Med</i> 2011;184:913–919.
572	7.	Young RP, Hopkins RJ, Christmas T, Black PN, Metcalf P, Gamble GD. COPD prevalence
573		is increased in lung cancer, independent of age, sex and smoking history. Eur Respir J
574		2009;34:380–386.
575	8.	Houghton AM. Mechanistic links between COPD and lung cancer. Nat Rev Cancer
576		2013;13:233–245.
577	9.	Cosio MG, Saetta M, Agusti A. Immunologic aspects of chronic obstructive pulmonary
578		disease. N Engl J Med 2009;360:2445–2454.

580	survival of patients with early-stage non-small cell lung cancer undergoing surgical resection
581	<i>Chest</i> 2014;145:346–353.
582	11. Turner MC, Chen Y, Krewski D, Calle EE, Thun MJ. Chronic obstructive pulmonary diseas
583	is associated with lung cancer mortality in a prospective study of never smokers. Am J Resp
584	<i>Crit Care Med</i> 2007;176:285–290.
585	12. Goc J, Germain C, Vo-Bourgais TKD, Lupo A, Klein C, Knockaert S, de Chaisemartin L,
586	Ouakrim H, Becht E, Alifano M, Validire P, Remark R, Hammond SA, Cremer I, Damotte
587	Fridman W-H, Sautès-Fridman C, Dieu-Nosjean M-C. Dendritic cells in tumor-associated
588	tertiary lymphoid structures signal a Th1 cytotoxic immune contexture and license the
589	positive prognostic value of infiltrating CD8+ T cells. Cancer Res 2014;74:705–715.
590	13. Fourcade J, Sun Z, Benallaoua M, Guillaume P, Luescher IF, Sander C, Kirkwood JM,
591	Kuchroo V, Zarour HM. Upregulation of Tim-3 and PD-1 expression is associated with
592	tumor antigen-specific CD8+ T cell dysfunction in melanoma patients. J Exp Med
593	2010;207:2175–2186.
594	14. Thommen DS, Schreiner J, Müller P, Herzig P, Roller A, Belousov A, Umana P, Pisa P,
595	Klein C, Bacac M, Fischer OS, Moersig W, Savic Prince S, Levitsky V, Karanikas V,
596	Lardinois D, Zippelius A. Progression of Lung Cancer Is Associated with Increased
597	Dysfunction of T Cells Defined by Coexpression of Multiple Inhibitory Receptors. Cancer
598	Immunol Res 2015;3:1344–1355.
599	15. Brahmer JR, Tykodi SS, Chow LQM, Hwu W-J, Topalian SL, Hwu P, Drake CG, Camacho
600	LH, Kauh J, Odunsi K, Pitot HC, Hamid O, Bhatia S, Martins R, Eaton K, Chen S, Salay
601	TM, Alaparthy S, Grosso JF, Korman AJ, Parker SM, Agrawal S, Goldberg SM, Pardoll D

10. Zhai R, Yu X, Shafer A, Wain JC, Christiani DC. The impact of coexisting COPD on

579

Gupta A, Wigginton JM. Safety and activity of anti-PD-L1 antibody in patients with
advanced cancer. N Engl J Med 2012;366:2455–2465.
16. Eggermont AMM, Chiarion-Sileni V, Grob J-J, Dummer R, Wolchok JD, Schmidt H, Hamid
O, Robert C, Ascierto PA, Richards JM, Lebbé C, Ferraresi V, Smylie M, Weber JS, Maio
M, Konto C, Hoos A, de Pril V, Gurunath RK, de Schaetzen G, Suciu S, Testori A. Adjuvant
ipilimumab versus placebo after complete resection of high-risk stage III melanoma (EORTC
18071): a randomised, double-blind, phase 3 trial. Lancet Oncol 2015;16:522–530.
17. Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, Powderly JD,
Carvajal RD, Sosman JA, Atkins MB, Leming PD, Spigel DR, Antonia SJ, Horn L, Drake
CG, Pardoll DM, Chen L, Sharfman WH, Anders RA, Taube JM, McMiller TL, Xu H,
Korman AJ, Jure-Kunkel M, Agrawal S, McDonald D, Kollia GD, Gupta A, Wigginton JM,
et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. N Engl J Med
2012;366:2443–2454.
18. Daud AI, Wolchok JD, Robert C, Hwu W-J, Weber JS, Ribas A, Hodi FS, Joshua AM,
Kefford R, Hersey P, Joseph R, Gangadhar TC, Dronca R, Patnaik A, Zarour H, Roach C,
Toland G, Lunceford JK, Li XN, Emancipator K, Dolled-Filhart M, Kang SP, Ebbinghaus S,
Hamid O. Programmed Death-Ligand 1 Expression and Response to the Anti-Programmed
Death 1 Antibody Pembrolizumab in Melanoma. J Clin Oncol 2016;34:4102–4109.
19. Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, Lee W, Yuan J,
Wong P, Ho TS, Miller ML, Rekhtman N, Moreira AL, Ibrahim F, Bruggeman C, Gasmi B,
Zappasodi R, Maeda Y, Sander C, Garon EB, Merghoub T, Wolchok JD, Schumacher TN,
Chan TA. Cancer immunology. Mutational landscape determines sensitivity to PD-1
blockade in non-small cell lung cancer. Science 2015;348:124–128.

625	20.	Borghaei H, Paz-Ares L, Horn L, Spigel DR, Steins M, Ready NE, Chow LQ, Vokes EE,
626		Felip E, Holgado E, Barlesi F, Kohlhäufl M, Arrieta O, Burgio MA, Fayette J, Lena H,
627		Poddubskaya E, Gerber DE, Gettinger SN, Rudin CM, Rizvi N, Crinò L, Blumenschein GR,
628		Antonia SJ, Dorange C, Harbison CT, Graf Finckenstein F, Brahmer JR. Nivolumab versus
629		Docetaxel in Advanced Nonsquamous Non-Small-Cell Lung Cancer. N Engl J Med
630		2015;373:1627–1639.
631	21.	Reck M, Rodríguez-Abreu D, Robinson AG, Hui R, Csőszi T, Fülöp A, Gottfried M, Peled
632		N, Tafreshi A, Cuffe S, O'Brien M, Rao S, Hotta K, Leiby MA, Lubiniecki GM, Shentu Y,
633		Rangwala R, Brahmer JR, KEYNOTE-024 Investigators. Pembrolizumab versus
634		Chemotherapy for PD-L1-Positive Non-Small-Cell Lung Cancer. N Engl J Med
635		2016;375:1823–1833.
636	22.	Prat A, Navarro A, Paré L, Reguart N, Galván P, Pascual T, Martínez A, Nuciforo P,
637		Comerma L, Alos L, Pardo N, Cedrés S, Fan C, Parker JS, Gaba L, Victoria I, Viñolas N,
638		Vivancos A, Arance A, Felip E. Immune-Related Gene Expression Profiling After PD-1
639		Blockade in Non-Small Cell Lung Carcinoma, Head and Neck Squamous Cell Carcinoma,
640		and Melanoma. Cancer Res 2017;77:3540–3550.
641	23.	Rabe KF, Hurd S, Anzueto A, Barnes PJ, Buist SA, Calverley P, Fukuchi Y, Jenkins C,
642		Rodriguez-Roisin R, van Weel C, Zielinski J, Global Initiative for Chronic Obstructive Lung
643		Disease. Global strategy for the diagnosis, management, and prevention of chronic
644		obstructive pulmonary disease: GOLD executive summary. Am J Respir Crit Care Med
645		2007;176:532–555.
646	24.	de Chaisemartin L, Goc J, Damotte D, Validire P, Magdeleinat P, Alifano M, Cremer I,
647		Fridman W-H, Sautès-Fridman C, Dieu-Nosjean M-C. Characterization of chemokines and

- adhesion molecules associated with T cell presence in tertiary lymphoid structures in human
 lung cancer. *Cancer Res* 2011;71:6391–6399.
- 650 25. Guo G, Sun X, Chen C, Wu S, Huang P, Li Z, Dean M, Huang Y, Jia W, Zhou Q, Tang A,

451 Yang Z, Li X, Song P, Zhao X, Ye R, Zhang S, Lin Z, Qi M, Wan S, Xie L, Fan F, Nickerson

- 652 ML, Zou X, Hu X, Xing L, Lv Z, Mei H, Gao S, et al. Whole-genome and whole-exome
- sequencing of bladder cancer identifies frequent alterations in genes involved in sister
- chromatid cohesion and segregation. *Nat Genet* 2013;45:1459–1463.
- 655 26. Mlecnik B, Bindea G, Angell HK, Maby P, Angelova M, Tougeron D, Church SE,
- Lafontaine L, Fischer M, Fredriksen T, Sasso M, Bilocq AM, Kirilovsky A, Obenauf AC,
- Hamieh M, Berger A, Bruneval P, Tuech J-J, Sabourin J-C, Le Pessot F, Mauillon J, Rafii A,
- Laurent-Puig P, Speicher MR, Trajanoski Z, Michel P, Sesboüe R, Frebourg T, Pagès F, et al.
- 659 Integrative Analyses of Colorectal Cancer Show Immunoscore Is a Stronger Predictor of

660 Patient Survival Than Microsatellite Instability. *Immunity* 2016;44:698–711.

- 661 27. Tumeh PC, Harview CL, Yearley JH, Shintaku IP, Taylor EJM, Robert L, Chmielowski B,
- 662 Spasic M, Henry G, Ciobanu V, West AN, Carmona M, Kivork C, Seja E, Cherry G,
- 663 Gutierrez AJ, Grogan TR, Mateus C, Tomasic G, Glaspy JA, Emerson RO, Robins H, Pierce
- 664 RH, Elashoff DA, Robert C, Ribas A. PD-1 blockade induces responses by inhibiting
- adaptive immune resistance. *Nature* 2014;515:568–571.
- 28. Thompson ED, Zahurak M, Murphy A, Cornish T, Cuka N, Abdelfatah E, Yang S, Duncan
- 667 M, Ahuja N, Taube JM, Anders RA, Kelly RJ. Patterns of PD-L1 expression and CD8 T cell
- 668 infiltration in gastric adenocarcinomas and associated immune stroma. Gut
- 669 2016;doi:10.1136/gutjnl-2015-310839.
- 670 29. Govindan R, Ding L, Griffith M, Subramanian J, Dees ND, Kanchi KL, Maher CA, Fulton R,
- 671 Fulton L, Wallis J, Chen K, Walker J, McDonald S, Bose R, Ornitz D, Xiong D, You M,
- 51 52

- Dooling DJ, Watson M, Mardis ER, Wilson RK. Genomic landscape of non-small cell lung
 cancer in smokers and never-smokers. *Cell* 2012;150:1121–1134.
- 30. Le Calvez F, Mukeria A, Hunt JD, Kelm O, Hung RJ, Tanière P, Brennan P, Boffetta P,
- 2675 Zaridze DG, Hainaut P. TP53 and KRAS mutation load and types in lung cancers in relation
- to tobacco smoke: distinct patterns in never, former, and current smokers. *Cancer Res*

677 2005;65:5076–5083.

- 678 31. Mansuet-Lupo A, Alifano M, Pécuchet N, Biton J, Becht E, Goc J, Germain C, Ouakrim H,
- 679 Régnard J-F, Cremer I, Laurent-Puig P, Dieu-Nosjean M-C, Blons H, Damotte D.
- 680 Intratumoral Immune Cell Densities Are Associated with Lung Adenocarcinoma Gene
- 681 Alterations. *Am J Respir Crit Care Med* 2016;194:1403–1412.
- 682 32. Dong Z-Y, Zhong W-Z, Zhang X-C, Su J, Xie Z, Liu S-Y, Tu H-Y, Chen H-J, Sun Y-L,
- 683 Zhou Q, Yang J-J, Yang X-N, Lin J-X, Yan H-H, Zhai H-R, Yan L-X, Liao R-Q, Wu S-P,
- 684 Wu Y-L. Potential Predictive Value of TP53 and KRAS Mutation Status for Response to PD-
- 1 Blockade Immunotherapy in Lung Adenocarcinoma. *Clin Cancer Res* 2017;23:3012–3024.
- 686 33. Skoulidis F, Byers LA, Diao L, Papadimitrakopoulou VA, Tong P, Izzo J, Behrens C, Kadara
- 687 H, Parra ER, Canales JR, Zhang J, Giri U, Gudikote J, Cortez MA, Yang C, Fan Y, Peyton
- 688 M, Girard L, Coombes KR, Toniatti C, Heffernan TP, Choi M, Frampton GM, Miller V,
- 689 Weinstein JN, Herbst RS, Wong K-K, Zhang J, Sharma P, et al. Co-occurring genomic
- alterations define major subsets of KRAS-mutant lung adenocarcinoma with distinct biology,
- 691 immune profiles, and therapeutic vulnerabilities. *Cancer Discov* 2015;5:860–877.
- 692 34. Agata Y, Kawasaki A, Nishimura H, Ishida Y, Tsubata T, Yagita H, Honjo T. Expression of
- the PD-1 antigen on the surface of stimulated mouse T and B lymphocytes. *Int Immunol*
- 694 1996;8:765–772.

695	35.	Austin JW, Lu P, Majumder P, Ahmed R, Boss JM. STAT3, STAT4, NFATc1, and CTCF
696		regulate PD-1 through multiple novel regulatory regions in murine T cells. J Immunol
697		2014;192:4876–4886.
698	36.	Bhowmik A, Seemungal TA, Sapsford RJ, Wedzicha JA. Relation of sputum inflammatory
699		markers to symptoms and lung function changes in COPD exacerbations. Thorax
700		2000;55:114–120.
701	37.	Brusselle GG, Joos GF, Bracke KR. New insights into the immunology of chronic
702		obstructive pulmonary disease. Lancet 2011;378:1015-1026.
703	38.	McKendry RT, Spalluto CM, Burke H, Nicholas B, Cellura D, Al-Shamkhani A, Staples KJ,
704		Wilkinson TMA. Dysregulation of Antiviral Function of CD8(+) T Cells in the Chronic
705		Obstructive Pulmonary Disease Lung. Role of the PD-1-PD-L1 Axis. Am J Respir Crit Care
706		Med 2016;193:642–651.
707	39.	Taube JM, Anders RA, Young GD, Xu H, Sharma R, McMiller TL, Chen S, Klein AP,
708		Pardoll DM, Topalian SL, Chen L. Colocalization of inflammatory response with B7-h1
709		expression in human melanocytic lesions supports an adaptive resistance mechanism of
710		immune escape. Sci Transl Med 2012;4:127ra37.
711	40.	Schmidt LH, Kümmel A, Görlich D, Mohr M, Bröckling S, Mikesch JH, Grünewald I, Marra
712		A, Schultheis AM, Wardelmann E, Müller-Tidow C, Spieker T, Schliemann C, Berdel WE,
713		Wiewrodt R, Hartmann W. PD-1 and PD-L1 Expression in NSCLC Indicate a Favorable
714		Prognosis in Defined Subgroups. PLoS ONE 2015;10:e0136023.
715	41.	Chen Y, Mu C-Y, Huang J-A. Clinical significance of programmed death-1 ligand-1
716		expression in patients with non-small cell lung cancer: a 5-year-follow-up study. Tumori
717		2012;98:751–755.

718	42.	Cooper WA, Tran T, Vilain RE, Madore J, Selinger CI, Kohonen-Corish M, Yip P, Yu B,
719		O'Toole SA, McCaughan BC, Yearley JH, Horvath LG, Kao S, Boyer M, Scolyer RA. PD-
720		L1 expression is a favorable prognostic factor in early stage non-small cell carcinoma. Lung
721		<i>Cancer</i> 2015;89:181–188.
722	43.	Velcheti V, Schalper KA, Carvajal DE, Anagnostou VK, Syrigos KN, Sznol M, Herbst RS,
723		Gettinger SN, Chen L, Rimm DL. Programmed death ligand-1 expression in non-small cell
724		lung cancer. Lab Invest 2014;94:107–116.
725	44.	Mu C-Y, Huang J-A, Chen Y, Chen C, Zhang X-G. High expression of PD-L1 in lung cancer
726		may contribute to poor prognosis and tumor cells immune escape through suppressing tumor
727		infiltrating dendritic cells maturation. Med Oncol 2011;28:682-688.
728	45.	Xu C, Fillmore CM, Koyama S, Wu H, Zhao Y, Chen Z, Herter-Sprie GS, Akbay EA,
729		Tchaicha JH, Altabef A, Reibel JB, Walton Z, Ji H, Watanabe H, Jänne PA, Castrillon DH,
730		Rustgi AK, Bass AJ, Freeman GJ, Padera RF, Dranoff G, Hammerman PS, Kim CF, Wong
731		K-K. Loss of Lkb1 and Pten leads to lung squamous cell carcinoma with elevated PD-L1
732		expression. Cancer Cell 2014;25:590-604.
733	46.	Parsa AT, Waldron JS, Panner A, Crane CA, Parney IF, Barry JJ, Cachola KE, Murray JC,
734		Tihan T, Jensen MC, Mischel PS, Stokoe D, Pieper RO. Loss of tumor suppressor PTEN
735		function increases B7-H1 expression and immunoresistance in glioma. Nat Med 2007;13:84-
736		88.
737	47.	Daud AI, Loo K, Pauli ML, Sanchez-Rodriguez R, Sandoval PM, Taravati K, Tsai K, Nosrati
738		A, Nardo L, Alvarado MD, Algazi AP, Pampaloni MH, Lobach IV, Hwang J, Pierce RH,
739		Gratz IK, Krummel MF, Rosenblum MD. Tumor immune profiling predicts response to anti-
740		PD-1 therapy in human melanoma. J Clin Invest 2016;126:3447–3452.

741	48. Borghaei H, Paz-Ares L, Horn L, Spigel DR, Steins M, Ready NE, Chow LQ, Vokes EE,
742	Felip E, Holgado E, Barlesi F, Kohlhäufl M, Arrieta O, Burgio MA, Fayette J, Lena H,
743	Poddubskaya E, Gerber DE, Gettinger SN, Rudin CM, Rizvi N, Crinò L, Blumenschein GR,
744	Antonia SJ, Dorange C, Harbison CT, Graf Finckenstein F, Brahmer JR. Nivolumab versus
745	Docetaxel in Advanced Nonsquamous Non-Small-Cell Lung Cancer. N Engl J Med
746	2015;373:1627–1639.
747	49. Mark NM, Kargl J, Busch SE, Yang GHY, Metz HE, Zhang H, Hubbard JJ, Pipavath SNJ,
748	Madtes DK, Houghton AM. COPD Alters Immune Cell Composition and Immune
749	Checkpoint Inhibitor Efficacy in NSCLC. Am J Respir Crit Care Med
750	2017;doi:10.1164/rccm.201704-0795OC.
751	50. Galon J, Mlecnik B, Bindea G, Angell HK, Berger A, Lagorce C, Lugli A, Zlobec I,
752	Hartmann A, Bifulco C, Nagtegaal ID, Palmqvist R, Masucci GV, Botti G, Tatangelo F,
753	Delrio P, Maio M, Laghi L, Grizzi F, Asslaber M, D'Arrigo C, Vidal-Vanaclocha F,
754	Zavadova E, Chouchane L, Ohashi PS, Hafezi-Bakhtiari S, Wouters BG, Roehrl M, Nguyen
755	L, et al. Towards the introduction of the "Immunoscore" in the classification of malignant
756	tumours. J Pathol 2014;232:199–209.
757	

758 Table 1

		<u>COPD</u> -		<u>COPD</u> ⁺		
Characteristic	S	Number	(%)	Number	(%)	P-value
Subjects		238	(55)	197	(45)	
Gender Male/Female		165/73	(69/31)	174/23	(88/12)	<0.0001*
Age (years) Mean ± SD		61 =	± 12	64 :	± 9	0.01
Smokers		182	(78)	173	(91)	0.0003*
Smoking histo Pack-years ± S	ry SD	36 =	± 24	49 ±	= 24	<0.0001
Histological Subtypes	ADC SSC Others	173 53 12	(73) (22) (5)	136 50 11	(69) (25) (6)	0.70*
pTNM stages	I II III-IV	102 58 78	(43) (24) (33)	94 59 44	(48) (30) (22)	0.05*
Vascular Emboli	Yes No ND	125 98 15	(53) (41) (6)	118 65 14	(60) (33) (7)	0.21*
Pleural Invasion	Yes No ND	124 110 4	(52) (46) (2)	110 83 4	(56) (42) (2)	0.70#
Lobectomy Pneumonector	ny	210 28	(88) (12)	171 26	(87) (13)	0.65*
% FEV1/FVC Mean ± SD		79	± 5	60 :	± 8	<0.0001
FEV1 (% of a predicted value) Mean ± SD		90	± 17	71 -	± 17	<0.0001

759

760 Table 1. Baseline characteristics of 435 patients with NSCLC (retrospective cohort).

All variables were evaluated among the 435 patients with NSCLC. Quantitative data were compared between COPD⁻ and COPD⁺ patients by Student's t test. Categorical data were compared by chi-square test (*) or Fisher exact test (#), as appropriate. *P* values <0.05 were considered statistically significant and appear in bold. Abbreviations: ADC, adenocarcinoma; SSC, squamous cell carcinoma; ND, Not determined.

766

61

62

- **Figures and figure legends:** 767
- Figure 1 768



Figure 1. Prognostic value of immune cell densities in NSCLC according to COPD 770 status (retrospective cohort). (A-D) Forest plots of univariate Cox-regression analysis 771 showing the impact of neutrophil (n=435), macrophage (n=435), CD8_{Tu} (n=427), CD8s 772 (n=435) and DC-Lamp⁺ cell (n=435) density on overall survival (OS) in the whole cohort (all 773 patients) (A), COPD⁻ (B), COPD⁺ (C) and COPD⁺ II-III patients (D). (E-G) The quartile 774 method was used to stratify patients into four groups by density of CD8_{Tu} cells, from the 775 lowest (1st quartile, black curves) to the highest density of CD8_{Tu} cells (4th quartile, red 776 curves). Kaplan-Meier curves of OS according to $CD8_{Tu}$ cell density in $COPD^{-}(E)$, $COPD^{+}$ 777 64 65

778	(F) and $COPD^+$ II-III patients (G). The horizontal dashed lines represent the median
779	survivals. (H-J) Forest plots of multivariate Cox-regression analysis showing the impact of
780	$CD8_{Tu}$, $CD8s$ and $DC-Lamp^+$ cell density on OS adjusted for age, gender, vascular emboli,
781	smoking history and stratified on the stage of the tumor, in $\text{COPD}^-(\text{H})$, $\text{COPD}^+(\text{I})$ and
782	COPD ⁺ II-III patients (J). P values <0.05 were considered statistically significant and appear
783	in bold.
784	
785	
786	
787	
788	
789	
790	
791	
792	
793	
794	
795	
796	
797	
798	
799	
800	
801	
802	
67 68 69	33



Figure 2. Characterization of CD8 tumor-infiltrating T lymphocytes (CD8 TILs) in 805 **NSCLC** (prospective cohort). 806

(A) Histograms showing, for one representative patient, the frequency of CTLA-4⁺, LAG-3⁺, 807 PD-1⁺ and TIM-3⁺ cells among CD8 T cells in Tumor, NT and blood. (B) Frequency of 808 CTLA-4⁺, LAG-3⁺, PD-1⁺ and TIM-3⁺ cells among CD8 T cells in Tumor, NT and blood. (C) 809 Histograms showing, for one representative patient, the frequency of Granzyme B⁺, TNF- α^+ , 810 IFN- γ^+ and IL-17⁺ cells among CD8 T cells in Tumor, NT and blood. (**D**) Frequency of 811 Granzyme B⁺, TNF- α^+ , IFN- γ^+ and IL-17⁺ cells among CD8 T cells in Tumor, NT and blood. 812 (E) Spearman-correlation matrix of parameters studied by flow cytometry. Each colored 813 square illustrates the correlation between two parameters. Red color illustrates a strong 814 70 71

815	positive correlation, and green color illustrates a strong negative correlation. (F) Frequencies
816	of PD-1 ⁺ TIM-3 ⁺ , PD-1 ⁺ TIM-3 ⁻ , PD-1 ⁻ TIM-3 ⁺ , PD-1 ⁻ TIM-3 ⁻ cells among CD8 T cells in
817	Tumor (black circle, n=50), NT (grey circle, n=47) and blood (white circle, n=41). (G)
818	Correlation between the frequencies of PD-1 ⁺ TIM-3 ⁺ , PD-1 ⁻ TIM-3 ⁺ , PD-1 ⁺ TIM-3 ⁻ and PD-
819	1 ⁻ TIM-3 ⁻ cells among CD8 TILs and frequencies of Granzyme B ⁺ , IL-17 ⁺ , TNF- α^+ and IFN-
820	$\gamma^{\scriptscriptstyle +}$ cells among CD8 TILs. In (B), (D) and (F) data are expressed as mean and an ANOVA or
821	a Kruskal-Wallis test followed by an appropriate correction was applied based on Shapiro
822	normality test. In (E) and (G) a spearman test was applied. $*p < 0.05$, $**p < 0.01$ and
823	*** <i>p</i> <0.001.
824	
825	
826	
827	
828	
829	
830	
831	
832	
833	
834	
835	
836	
837	
838	
839	
73 74 75	35



Figure 3. CD8 TIL exhaustion according to the COPD status of the patients
(prospective cohort).

(A-E) CD8 TIL characterization in COPD⁻ (n=30), COPD⁺ I (n=8) and COPD⁺ II-III (n=12) 844 845 patients. (A) Graphical representation of Spearman correlations between the FEV1% predicted and the frequencies of PD-1⁺, TIM-3⁺, PD-1⁺ TIM-3⁺, Granzyme B⁺, IFN- γ^+ and 846 TNF- α^+ cells among CD8 TILs, in COPD⁻ patients and COPD⁺ patients. Red and blue colors 847 indicate positive and negative correlations, respectively; the lighter is the color, the less 848 significant is the corresponding correlation. The filled fraction of the circle in each of the pie 849 charts corresponds to the absolute value of the associated Spearman correlation coefficient. 850 (B) Frequencies of PD-1⁺, TIM-3⁺ and PD-1⁺ TIM-3⁺ cells among CD8 TILs. (C) Graphical 851 representation of Spearman correlations in COPD⁻ and COPD⁺ patients, between CD8_{Tu} cell 852 76

77

853	densities and flow cytometry data, including the frequencies of PD-1 ⁺ , of TIM-3 ⁺ , of PD-1 ⁺
854	TIM-3 ⁺ , Granzyme B ⁺ , IFN- γ^+ and TNF- α^+ cells among CD8 TILs. Graphical representation
855	is the same than in Figure 3A. (D-E) The median $CD8_{Tu}$ cell density was used to stratify
856	patients into $CD8_{Tu}^{Low}(\mathbf{D})$ or $CD8_{Tu}^{High}(\mathbf{E})$ groups. (D-E) Histograms and radar plots showing
857	the frequencies of PD-1 ⁺ , TIM-3 ⁺ , PD-1 ⁺ TIM-3 ⁺ , IFN- γ^+ and TNF- α^+ cells among CD8 TILs
858	according to COPD status in $CD8_{Tu}^{Low}$ group and $CD8_{Tu}^{High}$ group. In (A) and (C) a Spearman
859	Test was performed. In (B) data are expressed as mean \pm SEM and a parametric test
860	(ANOVA with post-hoc Bonferroni correction) or a nonparametric test (Kruskal-Wallis test
861	followed by a post-hoc Dunn's test) was applied based on Shapiro normality test. In (D-E)
862	data are expressed as mean, and a parametric test (Student't test) or a nonparametric test
863	(Mann Whitney test) was applied based on Shapiro normality test. $p<0.05$, $p<0.01$ and
864	*** <i>p</i> <0.001.
865	
866	
867	
868	
869	
870	
871	
872	
873	
874	
875	
876	
877	
878 79	
80	37





Kaplan-Meier curves of OS in NSCLC patients according to PD-L1 expression by tumor cells, in COPD⁻ patients (n=234) (**A**) and in COPD⁺ patients (n=192) (**B**). (**C**) and (**D**) Kaplan-Meier curves of OS in the $CD8_{Tu}^{Low}$ group according to PD-L1 expression by tumor 82

886	cells in COPD ⁻ (n=111) (C) and COPD ⁺ patients (n=101) (D). (E) and (F) Kaplan-Me	eier
887	curves of OS in the $CD8_{Tu}^{High}$ group according to PD-L1 expression by tumor cells in COI	PD-
888	(n=123) (E) and COPD ⁺ patients (n=91) (F). (G) and (H) Forest plots of univariate Co	ox-
889	regression analyses showing the impact of $CD8_{Tu}$ cell, $CD8s$ cell and DC -Lamp ⁺ cell dense	sity
890	on the OS according to PD-L1 expression by tumor cells, in COPD ⁻ patients (G) and	in
891	COPD ⁺ patients (H). P values <0.05 were considered statistically significant and appear	in in
892	bold.	
893		
894		
895		
896		
897		
898		
899		
900		
901		
902		
903		
904		
905		
906		
907		
908		
909		
910		
85		
86 87		39



Figure 5. Nivolumab efficacy in advanced-stage NSCLC patients according to a
coexisting COPD and to smoke exposure. (A-B) Kaplan-Meier curves of progression-free

915 survival (PFS) and overall survival (OS) in COPD⁻ (n=20) and COPD⁺ patients (n=19). Progression-free survival was defined as the time from the start date of nivolumab treatment 916 to the date of the first documented event of tumor progression. (C) Characteristics of the 917 response to nivolumab treatment according to patients' COPD status and to smoking 918 exposure. TMB, Tumor mutation burden expressed as number of nonsynonymous mutations 919 per megabase and determined using whole exome sequencing experiments; Mu, Mutated-920 tumor; WT, Wild type-Tumor, NA; Not available. (D-G) Kaplan-Meier curves of 921 progression-free survival (D-E) and of overall survival (F-G) in COPD⁻ (n=20) (D,F) and 922 923 $COPD^+$ (n=19) (E,G) patients according to smoking exposure. Patients were stratified into two groups, Smoker^{High} (>30 pack-years) and Smoker^{Low} (≤30 pack-years). Tick marks 924 925 indicate censoring events. *P* values <0.05 were considered statistically significant and appear 926 in bold.