



**HAL**  
open science

# Use of Viral Load as a Surrogate Marker in Clinical Studies of Cytomegalovirus in Solid Organ Transplantation: A Systematic Review and Meta-analysis

Yoichiro Natori, Ali Alghamdi, Mahmood Tazari, Veronica Miller, Shahid Husain, Takashi Komatsu, Paul Griffiths, Per Ljungman, Ani Orchanian-Cheff, Deepali Kumar, et al.

► **To cite this version:**

Yoichiro Natori, Ali Alghamdi, Mahmood Tazari, Veronica Miller, Shahid Husain, et al.. Use of Viral Load as a Surrogate Marker in Clinical Studies of Cytomegalovirus in Solid Organ Transplantation: A Systematic Review and Meta-analysis. *Clinical Infectious Diseases*, 2018, 66 (4), pp.617-631. 10.1093/cid/cix793 . hal-03942946

**HAL Id: hal-03942946**

**<https://hal.sorbonne-universite.fr/hal-03942946>**

Submitted on 6 Feb 2023

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Copyright

IMMUNOCOMPROMISED HOSTS: David R. Snydman, Section Editor

# Use of Viral Load as a Surrogate Marker in Clinical Studies of Cytomegalovirus in Solid Organ Transplantation: A Systematic Review and Meta-analysis

Yoichiro Natori,<sup>1</sup> Ali Alghamdi,<sup>1,2</sup> Mahmood Tazari,<sup>1</sup> Veronica Miller,<sup>3</sup> Shahid Husain,<sup>1</sup> Takashi Komatsu,<sup>4</sup> Paul Griffiths,<sup>5</sup> Per Ljungman,<sup>6</sup> Ani Orchanian-Cheff,<sup>7</sup> Deepali Kumar,<sup>1,a</sup> and Atul Humar<sup>1,a</sup>; for the CMV Consensus Forum<sup>b</sup>

<sup>1</sup>Multi-Organ Transplant Program, University Health Network, Toronto, Ontario, Canada; <sup>2</sup>King Faisal Specialist Hospital & Research Center, Riyadh, Saudi Arabia; <sup>3</sup>Forum for Collaborative Research, University of California, Berkeley; <sup>4</sup>Division of Antiviral Products, Center for Drug Evaluation and Research, Food and Drug Administration, Silver Spring, Maryland; <sup>5</sup>Institute for Immunity and Transplantation, University College London Medical School, United Kingdom; <sup>6</sup>Division of Hematology, Department of Medicine Huddinge, Karolinska Institutet, Stockholm, Sweden; and <sup>7</sup>Library and Information Services, University Health Network, Toronto, Ontario, Canada

Symptomatic cytomegalovirus (CMV) disease has been the standard endpoint for clinical trials in organ transplant recipients. Viral load may be a more relevant endpoint due to low frequency of disease. We performed a meta-analysis and systematic review of the literature. We found several lines of evidence to support the validity of viral load as an appropriate surrogate end-point, including the following: (1) viral loads in CMV disease are significantly greater than in asymptomatic viremia (odds ratio, 9.3 95% confidence interval, 4.6–19.3); (2) kinetics of viral replication are strongly associated with progression to disease; (3) pooled incidence of CMV viremia and disease is significantly lower during prophylaxis compared with the full patient follow-up period (viremia incidence: 3.2% vs 34.3%;  $P < .001$ ) (disease incidence: 1.1% vs 13.0%;  $P < .001$ ); (4) treatment of viremia prevented disease; and (5) viral load decline correlated with symptom resolution. Based on the analysis, we conclude that CMV load is an appropriate surrogate endpoint for CMV trials in organ transplant recipients.

**Keywords.** cytomegalovirus (CMV) viremia; CMV disease; clinical trials; preemptive therapy; prophylaxis.

Cytomegalovirus (CMV) is one of the most common opportunistic infections after solid organ transplantation (SOT) and can produce a spectrum of illness categorized as either viral syndrome or tissue-invasive disease [1]. Viral syndrome typically presents with fever, fatigue, and cytopenias; published guidelines exist for appropriate definitions of CMV disease within particular organ groups for use in clinical trials [2]. The natural history of CMV infection after SOT is complex. A recipient lacking CMV immunoglobulin G antibodies (seronegative) before transplant may be infected from a seropositive donor to cause primary infection (D+/R–). The highest risk of CMV disease occurs after primary infection (D+/R–), followed by either reinfection or reactivation, which are less likely to cause disease [3]. There may be a trend toward more viremia in the D+/R+ group versus the D–/R+ group [1].

There are several laboratory methods to detect CMV, but most centers currently use quantitative viral load testing of whole blood or plasma, which most commonly detects CMV DNA using either a commercially available or in-house assay.

Cytomegalovirus prevention strategies include either universal antiviral prophylaxis typically with (val)ganciclovir, or a preemptive strategy that involves regular monitoring of viral load with initiation of antiviral therapy after detection above a certain threshold in order to prevent CMV disease [4–8]. The choice of prevention strategy depends on patient risk factors, including serostatus and type of transplant [1].

In the past, large randomized trials, designed primarily to demonstrate efficacy of antiviral strategies and to obtain regulatory approval for prophylaxis and/or treatment indications, have used symptomatic CMV disease as the primary endpoint. However, more recently this primary endpoint has been questioned for a number of reasons. In current clinical practice, rates of CMV disease are often low, in part due to prolonged prophylaxis, but also due to early detection of viremia and initiation of antiviral therapy before definitive symptoms attributable to CMV are evident [9]. In addition, currently, the most common form of CMV disease after SOT is viral syndrome. Although definitions for viral syndrome exist, it is clear that it represents a spectrum of illness that shares many overlapping features with other infectious and noninfectious etiologies. At a recent forum of content experts, industry, and regulatory advisors, including the US Food and Drug Administration and European Medicines Agency (CMV Forum, a project of the Forum for Collaborative Research University of California, Berkeley), the issue about the potential use of viral load as a surrogate marker in trials of CMV prevention or treatment arose as a major question pertaining to

Received 7 June 2017; editorial decision 22 August 2017; accepted 1 September 2017; published online September 5, 2017.

<sup>a</sup>D. K. and A. H. contributed equally to this work.

<sup>b</sup>Forum members listed in the appendix.

Correspondence: A. Humar, University Health Network, 585 University Ave, 11-PMB-175, Toronto, ON M5G 2N2, Canada (atul.humar@uhn.ca).

Clinical Infectious Diseases® 2018;66(4):617–31

© The Author(s) 2017. Published by Oxford University Press for the Infectious Diseases Society of America. All rights reserved. For permissions, e-mail: journals.permissions@oup.com. DOI: 10.1093/cid/cix793

development of new clinical trials. The CMV Forum delineated a process by which the utility of CMV load as a potential accepted surrogate endpoint in clinical trials could be more systematically evaluated. The results of that process and systematic review are compiled and presented here as they relate to SOT.

## METHODS

### Search Methods

A comprehensive search strategy was developed to identify published English-language literature on “cytomegalovirus,” “solid organ transplantation,” and “viral load.” The search strategy was developed by a medical librarian using a combination of database-specific subject headings and text words. Additional keywords were mined from sample articles and generated through input from subject specialists on the team. The search strategy was then customized for each database. No limits for date were applied. Animal-only studies were excluded where applicable, and no study-type filters were applied. Books and conference materials were excluded from Embase results. To ensure sensitivity, the initial strategy in MEDLINE was tested against 7 seminal articles and modified accordingly. The following databases were searched from inception to the date of the search (15 December 2016): Ovid MEDLINE, Ovid MEDLINE In-Process & Other Non-Indexed Citations, Ovid Embase, Cochrane Database of Systematic Reviews, and Cochrane Central Register of Controlled Clinical Trials. A supplementary search was conducted in PubMed for non-MEDLINE records. See the Supplementary Appendix for all search strategies. Two authors (Y. N. and A. A.) independently assessed all studies for risk of bias.

### Inclusion and Exclusion Criteria

We included cohort studies or randomized controlled trials of cytomegalovirus, solid organ transplantation, and viral load where the total cohort was >20 cases. We excluded animal studies, those of primarily other pathogens, studies with no use of quantitative polymerase chain reaction (PCR; eg, antigenemia, mRNA, qualitative PCR), review articles /letters without any new data, those not dealing with a solid organ transplant population, and those that did not document blood viral load. From included studies, we collected the following variables: number of subjects, organ transplant types, CMV load, incidence of CMV viremia and disease, CMV prevention strategy (prophylaxis or preemptive), and follow-up period.

### Data Analysis

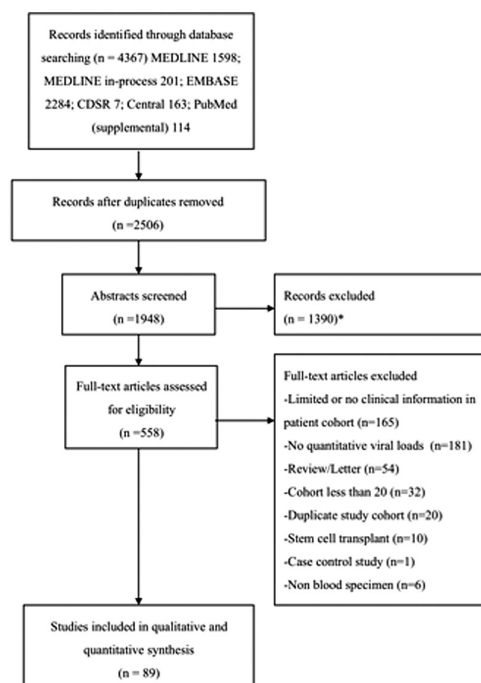
Where relevant, meta-analysis was performed by calculating the mean from the median and quartiles [10] and then standardizing the mean differences (SMD) between 2 groups. The log odds ratios were estimated from SMD [11] and then meta-analysis [12] done using SAS version 9.4 (SAS Institute, Cary, NC) using DerSimonian and Laird’s model. Heterogeneity among

studies was assessed with  $I^2$  values, which show the variation among studies that is not due to chance. The sensitivity analysis was repeated with fixed and random models with extracted outliers. Publication bias was examined by Egger test and graphed using a funnel plot [13]. Forest plots were based on the log odds ratios and confidence intervals with a value of 1 as the reference. Where relevant, pooled incidence rates were calculated based on the relative risk ratios (event/total) and their confidence interval [14]. We used the exact method of Clopper and Pearson in confidence interval estimation. The pooled values are based on a fixed-effect model for study on CMV disease during prophylaxis and a random-effects model for the other studies. The fixed or random model, for each combined study, was based on assessment of between-study heterogeneity.

## RESULTS

### Description of Studies

Our strategy resulted in 2506 potential studies. Of these, 1948 were excluded based on review of titles and abstracts because they did not meet eligibility criteria. These included studies that were not in human subjects or not in solid organ transplant recipients. A total of 558 studies underwent full text review, of which 469 were excluded for reasons outlined in Figure 1. This left a total of 89 studies for inclusion in the systematic review, of which a subset of studies were included in each of the meta-analyses performed.

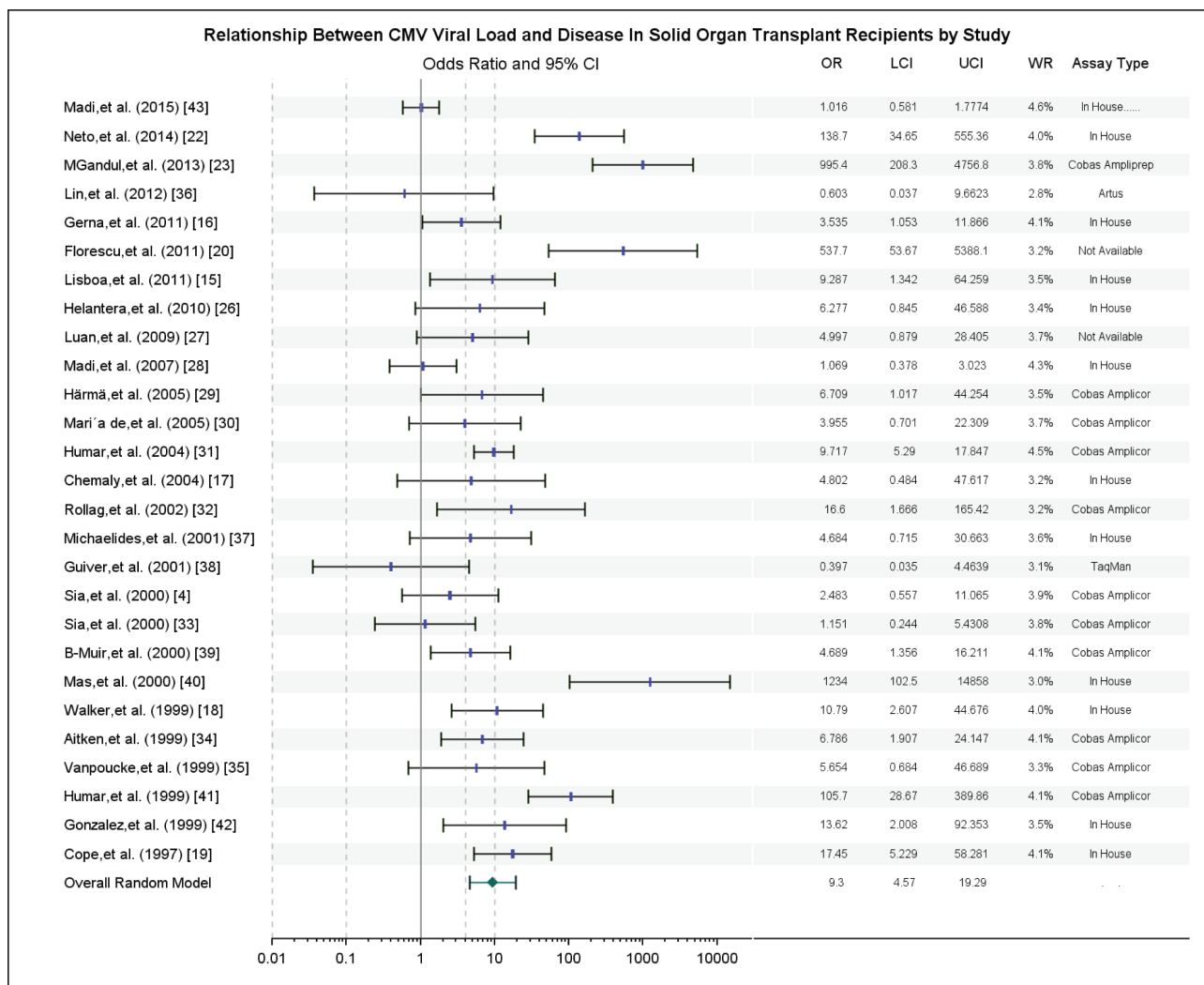


**Figure 1.** Study selection flow. \*The majority of these records were excluded because studies were qualitative, study cohort was <20 subjects, or the studies were primarily of viruses other than cytomegalovirus. Abbreviation: CDSR, Cochrane Database for Systematic Reviews.

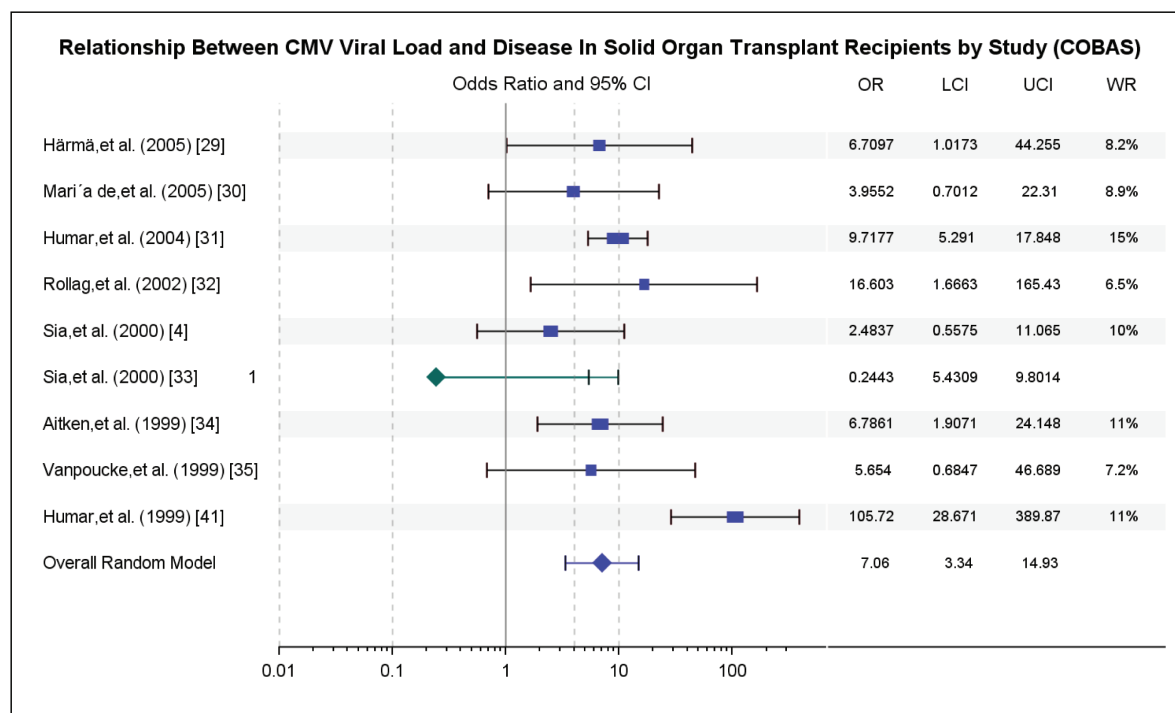
### Cytomegalovirus Load and Disease Prediction

We first assessed whether viral load was predictive of symptomatic CMV disease. Specifically we ascertained whether differences in viral load were present in patients with asymptomatic viremia versus symptomatic CMV disease (including both viral syndrome and tissue-invasive disease). We used any detectable viral load as the cutoff for positivity when assessing a study. A total of 30 studies that had addressed this question were identified [4, 15–43]. Of these 30 studies, 9 studies were natural history studies [17–19, 32, 34, 38–41]. Ten studies were conducted with COBAS Amplicor viral load assay, whereas the remaining studies used alternative PCR-based assays [4, 26, 29–35, 39, 41]. In a meta-analysis, we included all studies that directly compared these 2 groups (asymptomatic viremia vs CMV disease) (Figure 2A). As a second evaluation, we also conducted a meta-analysis

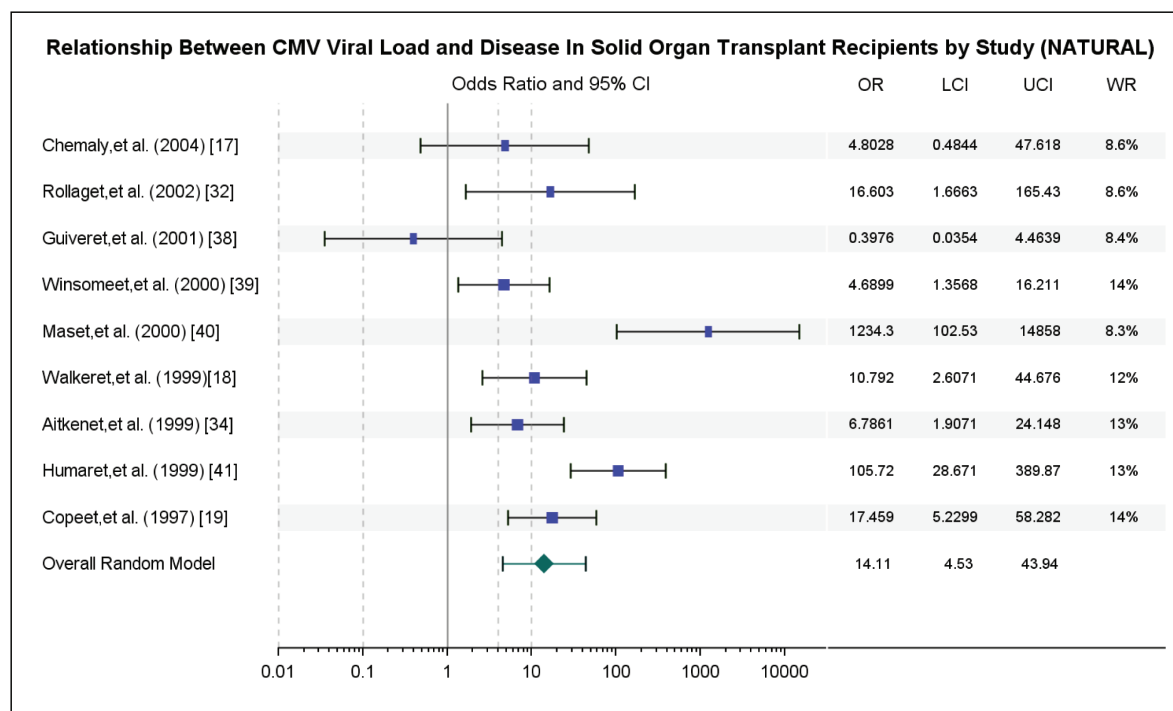
with only studies that used the COBAS Amplicor assay because this was the most common assay used (Figure 2B). Finally, we conducted a third meta-analysis that included only natural history studies (defined as studies in which no prophylaxis or preemptive therapy for asymptomatic CMV viremia is given) (Figure 2C). In all 3 analyses, symptomatic (vs asymptomatic) patients had a substantially and significantly greater viral load (Table 1). In the combined analysis, the mean fold difference in viral load for symptomatic versus asymptomatic patients was 14.9 (95% confidence interval [CI], 6.7–32.5; random-effects model). Based on a sensitivity analysis, there was publication bias for all included studies ( $P < .001$ ). After we removed 3 outliers, we again conducted a meta-analysis, which revealed that mean viral load in CMV disease remained significantly higher than in asymptomatic viremia (odds ratio [OR], 9.3; 95% CI, 4.6–19.3;



**Figure 2.** Relationship between cytomegalovirus (CMV) viral load and CMV disease in organ transplant recipients by study. Odds ratio and 95% confidence intervals are shown. A, All studies excluding outliers are shown (n = 27). Diamond symbol indicates the result of the DerSimonian-Laird random-effects model. Viral load assay type used in the study is also indicated.



**Figure 2.** B, Relationship between CMV load and disease in studies using the COBAS AmpliCor Viral Load Assay (n = 10).



**Figure 2.** C, Relationship between CMV viral load and disease in natural history studies (n = 9). Abbreviations: CI, confidence interval; LCI, lower confidence interval; OR, odds ratio; UCI, upper confidence interval; WR, weight (random).

random-effects model) without publication bias ( $P = .10$ ) (Figure 3). For studies using only the COBAS AmpliCor PCR assay, the fold difference was 7.0 (95% CI, 3.3 to 14.9; random-effects model). Finally for natural history studies only, the fold difference was 14.1 (95% CI, 4.5 to 43.9; random-effects model).

#### Viral Load Kinetics and the Risk of Cytomegalovirus Disease

We evaluated studies that assessed how viral load kinetics (change in viral load over time) influenced the likelihood of CMV disease. The hypothesis was that if viral load is an appropriate surrogate, viral kinetics will influence the likelihood of

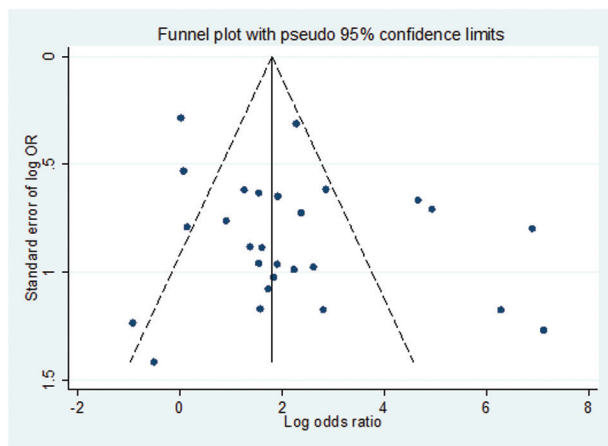
**Table 1. Summary of Odds Ratios of Asymptomatic Versus Symptomatic Cytomegalovirus Disease in Organ Transplant Recipients**

Study type	Number of studies	Model	Pooled OR (95% CI), <i>P</i> value	Heterogeneity (I <sup>2</sup> )	Studies excluded (references)
All included studies	27	REM	9.3 (4.6–19.3), <i>P</i> < .001	85.8	Elfadawy et al (2013), <sup>21</sup> Lisboa et al (2012), <sup>15</sup> Gala-Lopez et al (2011) <sup>24</sup>
Studies using COBAS Amplicor Viral Load Assay	10	REM	7.1 (3.3–14.9), <i>P</i> < .001	65.0	Helantera et al (2010) <sup>26</sup>
Natural history studies	9	REM	14.1 (4.5–43.9), <i>P</i> < .001	77.4	None

Abbreviations: CI, confidence interval; OR, odds ratio; REM, random-effects model.

disease development. Five studies were identified that specifically examined the change in viral load versus the risk of developing CMV disease. The first of the 5 studies showed that the rate of increase in CMV load between the last PCR-negative and first PCR-positive sample was significantly faster in patients with CMV disease ( $\log_{10}$  0.33 versus  $\log_{10}$  0.19 genomes/mL daily; *P* < .001) [44]. In multivariate-regression analyses, both initial CMV load and rate of viral load increase were independent risk factors for CMV disease [44].

A second study showed that the rate of increase in viral replication was strongly associated with progression to CMV disease [45]. A third study in lung transplant recipients, demonstrated that 1 –  $\log_{10}$  increases in CMV DNA levels at any time point during the first 6 months after transplant predicted CMV pneumonitis (sensitivity, 67%; specificity, 93%; *P* < .01) [37]. Another study demonstrated a 5- to 10-fold increase in the CMV DNA titers just prior to disease development [46]. Finally, in a study of kidney transplant recipients, an increase in viral load of  $\log_{10}$  0.7 copies per week also distinguished between CMV disease and asymptomatic seropositive recipients with high sensitivity (100%) and specificity (95%) [47]. In summary, all 5 studies suggested that a more rapid viral load increase correlated with the development of CMV disease.



**Figure 3.** Funnel plot of studies for relationship between cytomegalovirus load and disease (all studies excluding outliers, *n* = 27). Each dot represents a log odds ratio of each study. Black line represents calculated mean log odds ratio with all studies after excluding 3 outliers. Abbreviation: OR, odds ratio.

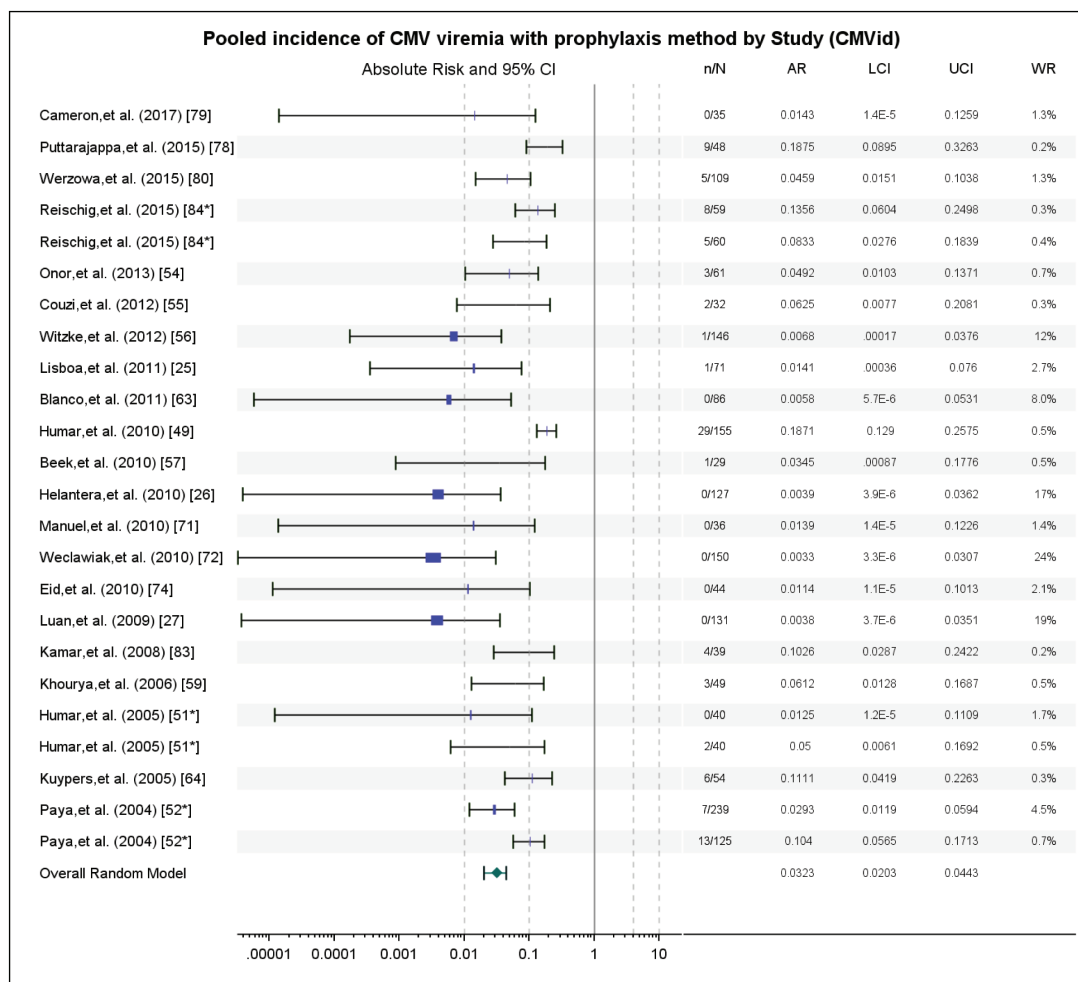
### Viral Load During and After Antiviral Prophylaxis

We hypothesized that, if viral load was an appropriate surrogate marker, then rates of viremia and disease may be low while a patient was on appropriate prophylaxis and significantly higher after discontinuation of prophylaxis. We identified 44 prophylaxis studies [20, 25–27, 35–37, 48–84]. Of these, 27 studies were identified as prophylaxis studies, whereas 17 studies were identified as prophylaxis versus preemptive therapy studies. The pooled incidence of CMV viremia during prophylaxis was determined for 24 studies for which sufficient data were available. The incidence of CMV viremia during prophylaxis varied from 0% to 18.6%. In the random-effect model, the pooled incidence of CMV viremia while on prophylaxis was 3.2% (95% CI, 2.0%–4.4%) (Figure 4). This contrasted with the incidence of viremia during the entire follow-up period (this includes during and after discontinuation of prophylaxis), which was 5.8% to 73% of patients. After excluding 1 outlier [65], we determined a pooled incidence for viremia of 34.3% (95% CI, 30.0%–38.6%; *P* < .001 vs. pooled incidence during prophylaxis) (Figure 5) (Table 2).

We next analyzed the incidence of CMV disease during prophylaxis based on data from 22 studies. The incidence of CMV disease during prophylaxis varied from 0% to 22.0%. Studies showed good homogeneity (*P* = .10), so a fixed-effect model was used. The pooled incidence was 0.8% (95% CI, 0.4%–1.3%) (Figure 6). In contrast, during the entire follow-up period, CMV disease incidence ranged 0%–36.8% in 41 studies. After excluding 1 outlier [65], we showed that the pooled incidence of disease was 13.0% (95% CI, 10.5%–15.5%; random-effects model; *P* < .001 vs pooled incidence during prophylaxis) (Figure 7). Therefore the incidence of viremia and disease during prophylaxis was overall low, with the majority of viremia and disease occurring after discontinuation of prophylaxis (Table 2).

### Therapy of Asymptomatic Viremia and Disease Prevention

We hypothesized that if viral load were an appropriate surrogate marker, then therapy of asymptomatic viremia should prevent progression to CMV disease. This is in fact the basis of preemptive strategies for CMV disease prevention. We identified 32 studies [23, 54–59, 62, 68, 70, 72, 76, 77, 80–83, 85–100] addressing this, of which 17 studies were prophylaxis versus preemptive therapy studies. The incidence of



**Figure 4.** Pooled incidence of cytomegalovirus viremia during antiviral prophylaxis showing the absolute risk and 95% confidence intervals for each study ( $n = 24$ ). Abbreviations: AR, absolute risk; CI, confidence interval; CMV, cytomegalovirus; LCI, lower confidence interval; N, total number of patients in the study; n, patients with the outcome of interest (cytomegalovirus viremia); UCI, upper confidence interval; WR, weight (random). \*Different cohort from same reference.

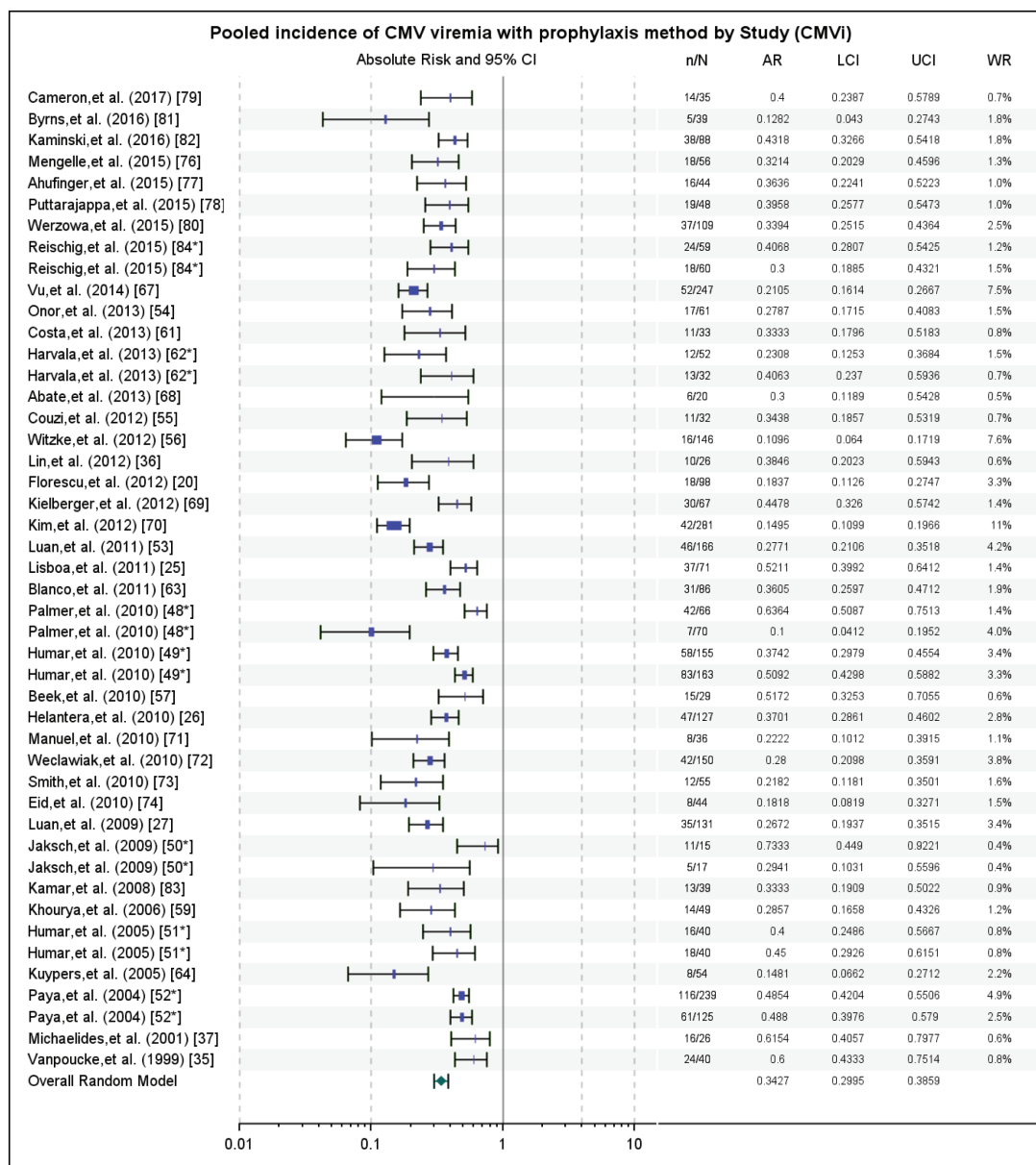
CMV viremia with a preemptive strategy varied from 5.3% (CMV-seropositive liver transplant recipient) to 91.7% (CMV D+/R– kidney, liver, and heart transplant recipients) with a pooled incidence of CMV viremia of 48.9% (95% CI, 39.6%–58.1%) (Figure 8). The incidence of CMV disease varied from 0% to 26.3% with a pooled incidence of 6.9% (95% CI, 5.2%–8.5%) (Figure 9). The pooled disease incidence was significantly lower than the pooled viremia incidence ( $P < .001$ ) (Table 2).

#### Viral Load Response Versus Symptom Resolution

We hypothesized that, if viral load is a valid surrogate marker, then clinical response in patients with CMV disease should mirror virologic response. Three studies were identified that provided relevant data [6, 101, 102]. Two studies were from the VICTOR cohort, a large randomized trial comparing intravenous ganciclovir versus oral valganciclovir for the treatment of CMV disease [101, 102]. Both studies demonstrated

a correlation between viral load decline and symptom resolution for both CMV tissue-invasive disease and viral syndrome using either the COBAS Amplicor [101] assay or the Roche Taqman international unit–based assay [102]. For example, of 267 patients, 251 had CMV disease resolution by day 49 of treatment. Patients with pretreatment CMV DNA of  $<18\ 200$  IU/mL had a faster time to disease resolution ( $P = .001$ ), and patients with CMV load suppression at days 7, 14, and 21 had faster times to clinical disease resolution ( $P = .005$ ,  $<.001$ , and  $<.001$ , respectively) [102]. One additional study assessed 26 patients with biopsy-proven gastrointestinal CMV disease [6]. The median time to negative viral load (22.5 days) was similar to the median reported time to negative follow-up endoscopy (27.0 days).

Therefore, despite a small number of studies, viral load decline during the treatment phase seems to correlate well with symptom resolution, and specifically, achieving viral load negativity correlates strongly with symptom response.



**Figure 5.** Pooled incidence of cytomegalovirus viremia for the entire follow-up period in studies using antiviral prophylaxis. Abbreviations: AR, absolute risk; CI, confidence interval; CMV, cytomegalovirus; LCI, lower confidence interval; N, total number of patients in the study; n, patients with the outcome of interest (cytomegalovirus viremia); UCI, upper confidence interval; WR, weight (random). \*Different cohorts from same reference.

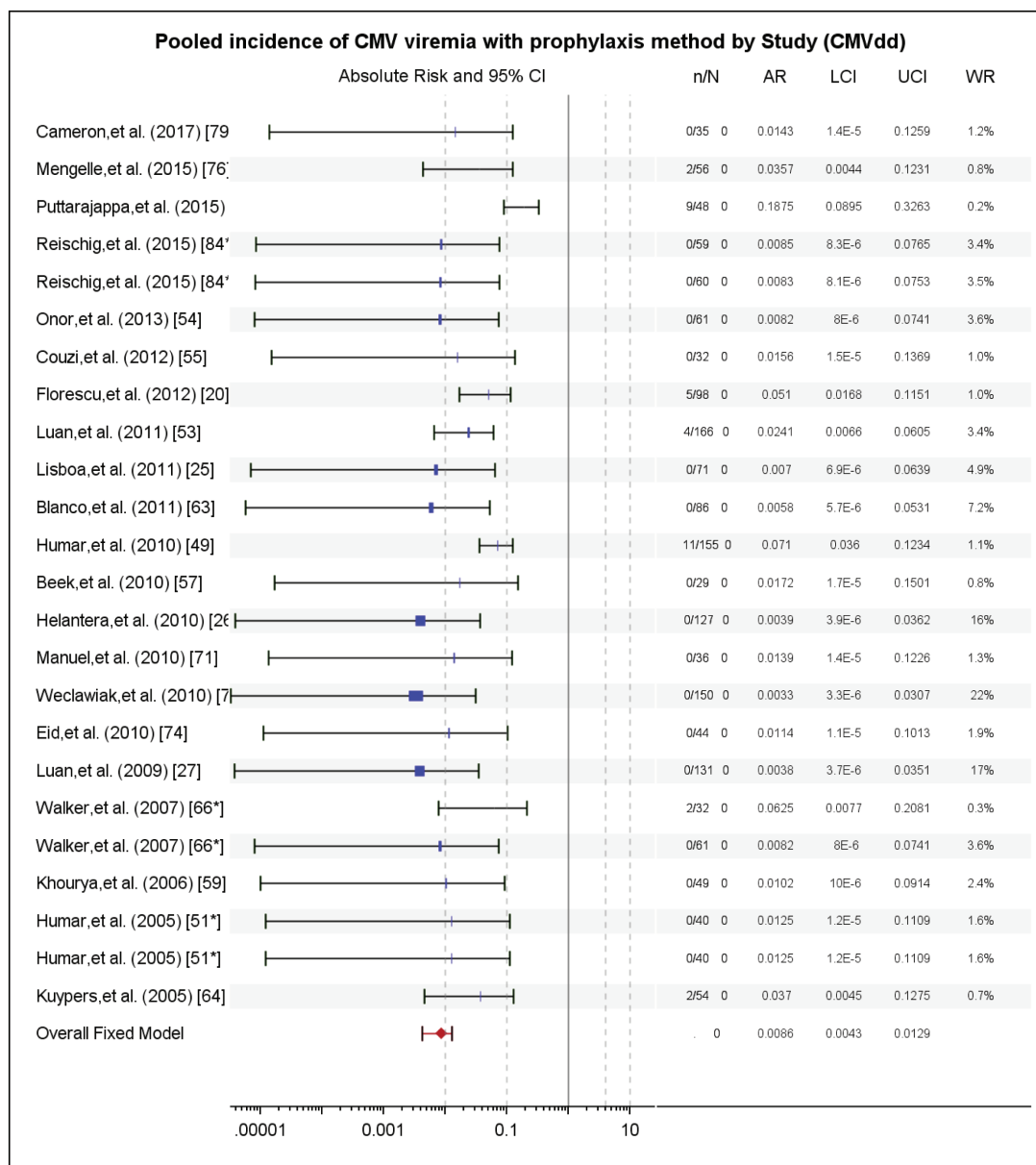
**Table 2. Summary of Pooled Incidences of Cytomegalovirus Infection Depending on Prevention Method and Followup Period**

Prevention method	Follow-up period <sup>a</sup>	Outcome	No. of studies	Model	Pooled incidence (%) (95% CI)	Heterogeneity ( $I^2$ ), $P$ value
Prophylaxis	During prophylaxis	CMV viremia	24	REM	3.2 (2.0–4.4)	75.5, $P < .001$
	During prophylaxis	CMV disease	24	FEM	0.8 (0.4–1.3)	28.1, $P = .10$
	Entire study period	CMV viremia	46	REM	34.3 (30.0–38.6)	88.5, $P < .001$
Preemptive	Entire study period	CMV viremia	33	REM	48.9 (39.6–58.1)	98.2, $P < .001$
	Entire study period	CMV disease	27	REM	6.9 (5.2–8.5)	78.6, $P < .001$

Abbreviations: CI, confidence interval; CMV, cytomegalovirus; FEM, fixed-effects model; REM, random-effects model.

<sup>a</sup>Entire study period includes complete study follow-up, including during and after prophylaxis.



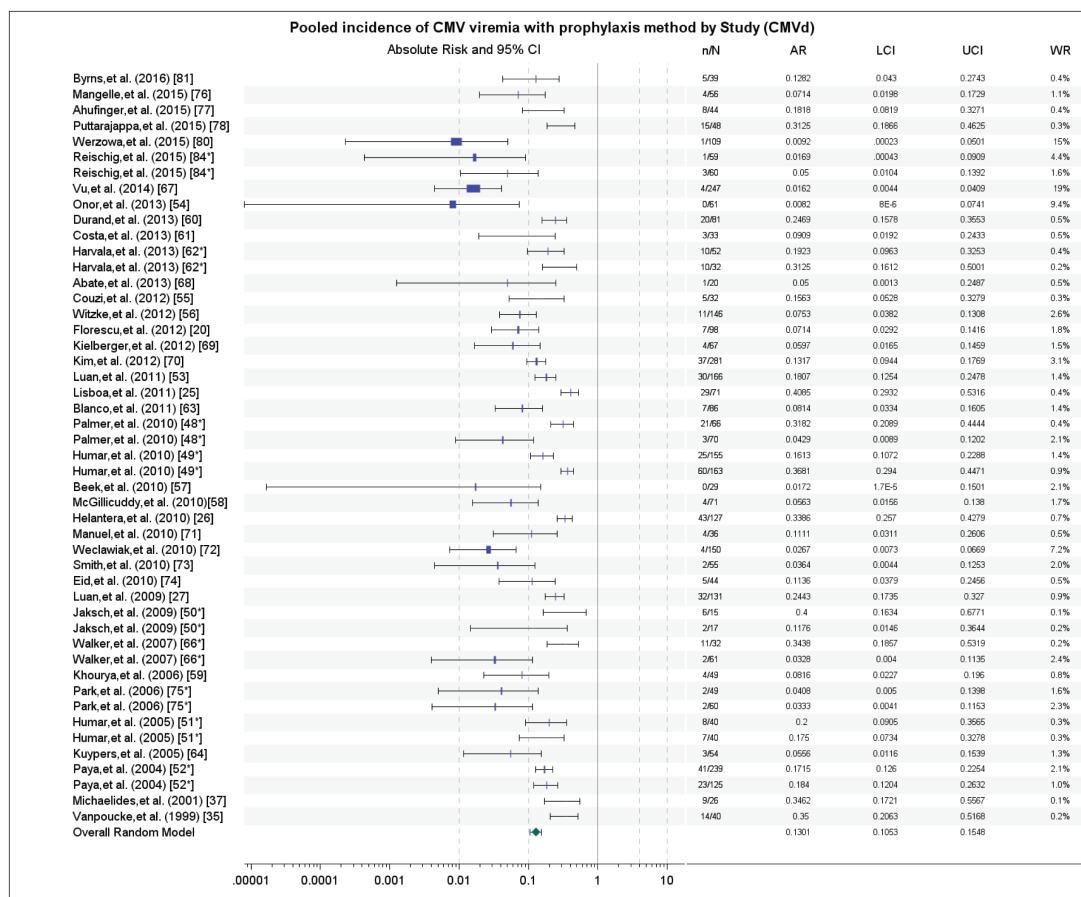


**Figure 6.** Pooled incidence of cytomegalovirus disease during antiviral prophylaxis. Abbreviations: AR, absolute risk; LCI, lower confidence interval; N, total number of patients in the study; n, patients with the outcome of interest (cytomegalovirus disease); UCI, upper confidence interval; WR, weight (random). \*Different cohorts from same reference.

## DISCUSSION

We conducted a systematic review and meta-analysis of data related to quantitative viral load testing for CMV and its potential utility as a surrogate marker in clinical trials of CMV after SOT. Viremia is considered a marker of viral lytic cycle activity, which is important for CMV disease development. To better define this, we assessed specific aspects related to different trial designs and indications of antivirals (prophylaxis, preemptive, and treatment). Five predefined and separate areas were examined to arrive at an overall understanding of the potential of viral load testing as a surrogate marker. First, we assessed whether viral load was higher in patients with asymptomatic

viremia versus symptomatic CMV disease. We found the fold difference in viral load was much greater in disease versus asymptomatic viremia, (9.3, 7.0, 14.1 with all included studies, trials, studies using only the COBAS Amplicor PCR assay, and natural history studies, respectively). We then assessed whether the rate in change of viral load was predictive of CMV disease. The number of studies was smaller, but each demonstrated a consistent effect relating viral load kinetics to likelihood of disease development. In the third section, we reviewed viremia and disease during prophylaxis versus the entire period of follow-up. The pooled analysis demonstrated low rates of viremia and disease during periods of antiviral prophylaxis compared

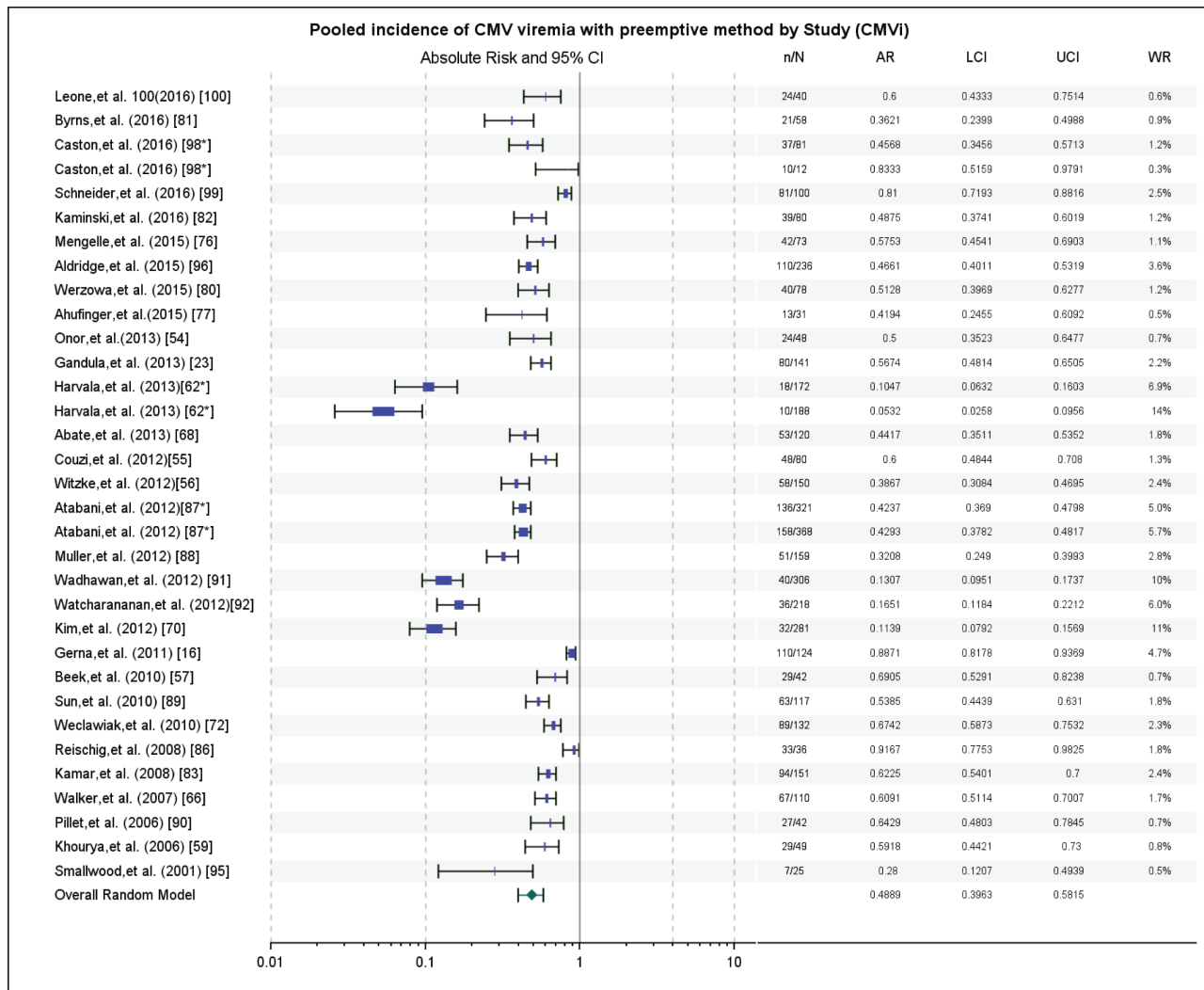


**Figure 7.** Pooled incidence of cytomegalovirus disease for the entire follow-up period in studies using antiviral prophylaxis. Abbreviations: AR, absolute risk; LCI, lower confidence interval; N, total number of patients in the study; n, patients with the outcome of interest (cytomegalovirus disease); UCI, upper confidence interval; WR, weight (random). \*Different cohorts from same reference.

with a significantly higher pooled incidence of viremia and disease in the total follow-up period of the trial. We next reviewed the pooled data on treatment of asymptomatic viremia and the development of CMV disease (preemptive strategy). We observed that the pooled incidence of asymptomatic viremia was 48.9% and the pooled incidence of disease was 6.9%, suggesting that treatment of asymptomatic viremia results in a low incidence of subsequent disease. Finally, we assessed treatment studies, which linked symptom resolution to virologic response. The number of studies was small but strongly indicated that clearance of viremia correlated with symptom resolution.

Although these data in isolation do not provide definitive proof for the utility of viral load as a surrogate endpoint, when taken together, they provide compelling rationale that this may be a reasonable primary outcome in clinical trials. Given the low incidence of CMV disease in most modern clinical settings, coupled with difficulties in the definitive clinical diagnosis of CMV disease after SOT (especially viral syndrome), CMV load may in fact be a preferable and more robust endpoint.

This systematic review and meta-analysis has several limitations for each of the areas analyzed. In all cases, we combined data across studies to estimate effect sizes or associations. However, studies by their nature often include different transplant types, different immunosuppression drugs (eg, different induction agents or rejection episodes), and other factors that may result in heterogeneity of populations. For example, the studies by Lin et al and Guiver et al were less supportive of the link between CMV load and disease likely due to relatively small sample sizes. Analysis-specific limitations were also present. For example, in the analysis of viral load in asymptomatic versus symptomatic patients, viral load assays (including in-house assays), disease definitions, and different prophylactic regimens all likely influence the outcome. To better control for this, we also assessed just studies that used the COBAS Amplicor assay (as the most common viral load assay) and natural history studies. We also did not differentiate viral loads between CMV syndrome and end-organ disease because a very small number of studies addressed these differences. This analysis is also difficult to interpret because some patients labeled



**Figure 8.** Pooled incidence of cytomegalovirus viremia in studies using preemptive method. Abbreviations: AR, absolute risk; LCI, lower confidence interval; N, total number of patients in the study; n, patients with the outcome of interest (cytomegalovirus disease); UCI, upper confidence interval; WR, weight (random). \*Different cohorts from same reference.

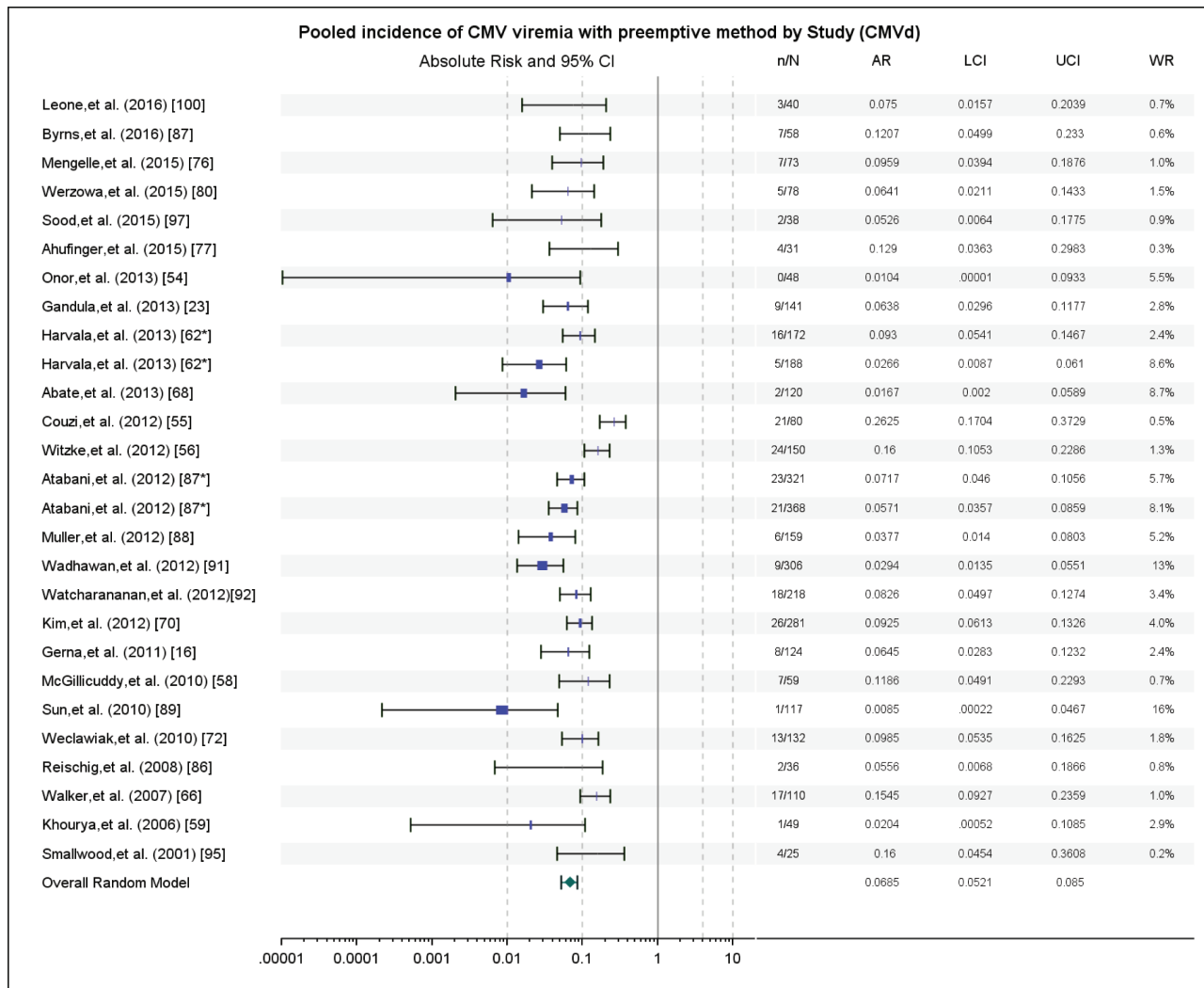
as syndrome may have had end-organ disease but did not have a biopsy to prove it. Finally, there are limited pediatric data, and the results presented here may not be generalizable to this population.

Although CMV viremia can be associated with end-organ disease, there are exceptions, including occasional patients who develop CMV disease (especially gastrointestinal disease) with an undetectable viral load [6]. The pathogenesis of end-organ disease in the absence of viremia is uncertain but may reflect local organ-restricted CMV reactivation, something that may behave differently in primary infection versus reactivation or reinfection. Similarly, assessment of viral load in specific organs or tissues such as bronchoalveolar lavage fluid is not included in this analysis. Future clinical trials of candidate CMV drugs or vaccines should therefore attempt to differentiate among primary infection,

reinfection, and reactivation and consider that the validity of CMV viremia as a surrogate marker may differ in these different types of infection.

This study was not designed to address the question of optimal viral thresholds, and as noted in the CMV Consensus Guidelines, there are insufficient data for this [1]. The majority of studies analyzed in this review used viral load in copies per milliliter rather than the international standard; however, the use of the international standard is to assist in comparison across laboratories [103], whereas the purpose of this analysis was to assess the potential for surrogacy of viral load regardless of how it is measured.

The findings in this study are consistent with published data in other areas. For example, in human immunodeficiency virus patients, CMV viremia has been shown to be an independent predictor of mortality [104, 105]. In addition,



**Figure 9.** Pooled incidence of cytomegalovirus disease in studies using preemptive method. Abbreviations: AR, absolute risk; LCI, lower confidence interval; N, total number of patients in the study; n, patients with the outcome of interest (cytomegalovirus disease); UCI, upper confidence interval; WR, weight (random). \*Different cohorts from same reference.

for other viruses, including hepatitis C virus, hepatitis B virus, and human immunodeficiency virus, viral load is routinely used as a surrogate endpoint in clinical trials. For example, recent trials of direct-acting antivirals for HCV routinely use virologic clearance as their primary endpoint [106, 107].

In conclusion, we performed a systematic review of viral load testing in different types of CMV-related trials in solid organ transplant recipients (prophylaxis, preemptive, and treatment). Based on this systematic review, we conclude that viral load likely predicts clinical endpoints in CMV trials in solid organ transplant recipients and may have some logistic and practical benefits over clinical endpoints. Overall, use of viral load as a surrogate endpoint in clinical trials may help speed up development of new antiviral agents and new diagnostics such as cell-mediated immunity assays.

### Supplementary Data

Supplementary materials are available at *Clinical Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

### Notes

**Acknowledgments.** Y. N., A. A., and A. O.-C. performed the literature search. T. M., Y. N., A. H., and D. K. performed the data analysis. All authors were responsible for the study design, data interpretation, and writing.

**Financial support.** The study was funded by the Forum for Collaborative Research, University of California–Berkeley. The CMV Forum provided a stipend for a postdoctoral fellow and funding for a consensus meeting to discuss the results of this study.

**Potential conflicts of interest.** D. K. has received clinical trials grants from Roche, Qiagen, and Oxford Immunotec, speaker honoraria from Astellas, and consulting fees from Qiagen. A. H. has received clinical trials grants from Roche, Qiagen, Astellas, and Chimerix. V. M. has received unrestricted grants from Abbott Laboratories, Astellas, Chimerix,

Microbiotix, Merck, Shire. P. L. has received clinical trials grants from Merck and Astellas and advisory board honoraria from AiCuris. All other authors declare no potential conflicts. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

## References

- Kotton CN, Kumar D, Caliendo AM, et al; Transplantation Society International CMV Consensus Group. Updated international consensus guidelines on the management of cytomegalovirus in solid-organ transplantation. *Transplantation* **2013**; 96:333–60.
- Ljungman P, Boeckh M, Hirsch HH, et al; Disease Definitions Working Group of the Cytomegalovirus Drug Development Forum. Definitions of cytomegalovirus infection and disease in transplant patients for use in clinical trials. *Clin Infect Dis* **2017**; 64:87–91.
- Grundy JE, Lui SF, Super M, et al. Symptomatic cytomegalovirus infection in seropositive kidney recipients: reinfection with donor virus rather than reactivation of recipient virus. *Lancet* **1988**; 2:132–5.
- Sia IG, Wilson JA, Groettum CM, Espy MJ, Smith TF, Paya CV. Cytomegalovirus (CMV) DNA load predicts relapsing CMV infection after solid organ transplantation. *J Infect Dis* **2000**; 181:717–20.
- Shanahan A, Malani PN, Kaul DR. Relapsing cytomegalovirus infection in solid organ transplant recipients. *Transpl Infect Dis* **2009**; 11:513–8.
- Eid AJ, Arthurs SK, Deziel PJ, Wilhelm MP, Razonable RR. Clinical predictors of relapse after treatment of primary gastrointestinal cytomegalovirus disease in solid organ transplant recipients. *Am J Transplant* **2010**; 10:157–61.
- Mumtaz K, Faisal N, Husain S, Morillo A, Renner EL, Shah PS. Universal prophylaxis or preemptive strategy for cytomegalovirus disease after liver transplantation: a systematic review and meta-analysis. *Am J Transplant* **2015**; 15:472–81.
- Zuk DM, Humar A, Weinkauff JG, Lien DC, Nador RG, Kumar D. An international survey of cytomegalovirus management practices in lung transplantation. *Transplantation* **2010**; 90:672–6.
- Manuel O, Kralidis G, Mueller NJ, et al; Swiss Transplant Cohort Study. Impact of antiviral preventive strategies on the incidence and outcomes of cytomegalovirus disease in solid organ transplant recipients. *Am J Transplant* **2013**; 13:2402–10.
- Hozo SP, Djulbegovic B, Hozo I. Estimating the mean and variance from the median, range, and the size of a sample. *BMC Med Res Methodol* **2005**; 5:13.
- da Costa BR, Rutjes AW, Johnston BC, et al. Methods to convert continuous outcomes into odds ratios of treatment response and numbers needed to treat: meta-epidemiological study. *Int J Epidemiol* **2012**; 41:1445–59.
- Hedges LV, Pigott TD. The power of statistical tests in meta-analysis. *Psychol Methods* **2001**; 6:203–17.
- Egger M, Davey Smith G, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ* **1997**; 315:629–34.
- Newcombe RG. Two-sided confidence intervals for the single proportion: comparison of seven methods. *Stat Med* **1998**; 17:857–72.
- Lisboa LF, Kumar D, Wilson LE, Humar A. Clinical utility of cytomegalovirus cell-mediated immunity in transplant recipients with cytomegalovirus viremia. *Transplantation* **2012**; 93:195–200.
- Gerna G, Lilleri D, Chiesa A, et al. Virologic and immunologic monitoring of cytomegalovirus to guide preemptive therapy in solid-organ transplantation. *Am J Transplant* **2011**; 11:2463–71.
- Chemaly RF, Yen-Lieberman B, Castilla EA, et al. Correlation between viral loads of cytomegalovirus in blood and bronchoalveolar lavage specimens from lung transplant recipients determined by histology and immunohistochemistry. *J Clin Microbiol* **2004**; 42:2168–72.
- Hassan-Walker AF, Kidd IM, Sabin C, Sweny P, Griffiths PD, Emery VC. Quantity of human cytomegalovirus (CMV) DNAemia as a risk factor for CMV disease in renal allograft recipients: relationship with donor/recipient CMV serostatus, receipt of augmented methylprednisolone and antithymocyte globulin (ATG). *J Med Virol* **1999**; 58:182–7.
- Cope AV, Sabin C, Burroughs A, Rolles K, Griffiths PD, Emery VC. Interrelationships among quantity of human cytomegalovirus (HCMV) DNA in blood, donor-recipient serostatus, and administration of methylprednisolone as risk factors for HCMV disease following liver transplantation. *J Infect Dis* **1997**; 176:1484–90.
- Florescu DF, Langnas AN, Grant W, et al. Incidence, risk factors, and outcomes associated with cytomegalovirus disease in small bowel transplant recipients. *Pediatr Transplant* **2012**; 16:294–301.
- Elfadawy N, Flechner SM, Liu X, et al. CMV viremia is associated with a decreased incidence of BKV reactivation after kidney and kidney-pancreas transplantation. *Transplantation* **2013**; 96:1097–103.
- David-Neto E, Triboni AH, Paula FJ, et al. A double-blinded, prospective study to define antigenemia and quantitative real-time polymerase chain reaction cutoffs to start preemptive therapy in low-risk, seropositive, renal transplanted recipients. *Transplantation* **2014**; 98:1077–81.
- Martin-Gandul C, Pérez-Romero P, Sánchez M, et al; Spanish Network for Research in Infectious Diseases. Determination, validation and standardization of a CMV DNA cut-off value in plasma for preemptive treatment of CMV infection in solid organ transplant recipients at lower risk for CMV infection. *J Clin Virol* **2013**; 56:13–8.
- Gala-Lopez BL, Senior PA, Koh A, et al. Late cytomegalovirus transmission and impact of T-depletion in clinical islet transplantation. *Am J Transplant* **2011**; 11:2708–14.
- Lisboa LF, Preiksaitis JK, Humar A, Kumar D. Clinical utility of molecular surveillance for cytomegalovirus after antiviral prophylaxis in high-risk solid organ transplant recipients. *Transplantation* **2011**; 92:1063–8.
- Helanterä I, Kyllönen L, Lautenschlager I, Salmela K, Koskinen P. Primary CMV infections are common in kidney transplant recipients after 6 months valganciclovir prophylaxis. *Am J Transplant* **2010**; 10:2026–32.
- Luan FL, Stuckey LJ, Park JM, Kaul D, Cibrik D, Ojo A. Six-month prophylaxis is cost effective in transplant patients at high risk for cytomegalovirus infection. *J Am Soc Nephrol* **2009**; 20:2449–58.
- Madi N, Al-Nakib W, Mustafa AS, Saeed T, Pacsa A, Nampoory MR. Detection and monitoring of cytomegalovirus infection in renal transplant patients by quantitative real-time PCR. *Med Princ Pract* **2007**; 16:268–73.
- Härmä M, Loginov R, Piipainen H, Halme L, Höckerstedt K, Lautenschlager I. HHV-6-DNAemia related to CMV-DNAemia after liver transplantation. *Transplant Proc* **2005**; 37:1230–2.
- de Oña M, Melón S, Gallarraga MC, et al. Comparison of cytomegalovirus pp-65 antigenemia assay and plasma DNA correlation with the clinical outcome in transplant recipients. *Transpl Int* **2005**; 18:43–6.
- Humar A, Paya C, Pescovitz MD, et al. Clinical utility of cytomegalovirus viral load testing for predicting CMV disease in D+/R- solid organ transplant recipients. *Am J Transplant* **2004**; 4:644–9.
- Rollag H, Sagedal S, Kristiansen KI, et al. Cytomegalovirus DNA concentration in plasma predicts development of cytomegalovirus disease in kidney transplant recipients. *Clin Microbiol Infect* **2002**; 8:431–4.
- Sia IG, Wilson JA, Espy MJ, Paya CV, Smith TF. Evaluation of the COBAS AMPLICOR CMV MONITOR test for detection of viral DNA in specimens taken from patients after liver transplantation. *J Clin Microbiol* **2000**; 38:600–6.
- Aitken C, Barrett-Muir W, Millar C, et al. Use of molecular assays in diagnosis and monitoring of cytomegalovirus disease following renal transplantation. *J Clin Microbiol* **1999**; 37:2804–7.
- Vanpoucke H, Van Vlem B, Vanholder R, Van Renterghem L. Significance of qualitative polymerase chain reaction combined with quantitation of viral load in the diagnosis and follow-up of cytomegalovirus infection after solid-organ transplantation. *Intervirology* **1999**; 42:398–404.
- Lin A, Worley S, Brubaker J, et al. Assessment of cytomegalovirus hybrid preventative strategy in pediatric heart transplant patients. *J Pediatric Infect Dis Soc* **2012**; 1:278–83.
- Michaelides A, Liolios L, Glare EM, et al. Increased human cytomegalovirus (HCMV) DNA load in peripheral blood leukocytes after lung transplantation correlates with HCMV pneumonitis. *Transplantation* **2001**; 72:141–7.
- Guiver M, Fox AJ, Mutton K, Mogulkoc N, Egan J. Evaluation of CMV viral load using TaqMan CMV quantitative PCR and comparison with CMV antigenemia in heart and lung transplant recipients. *Transplantation* **2001**; 71:1609–15.
- Barrett-Muir W, Breuer J, Millar C, et al. CMV viral load measurements in whole blood and plasma—which is best following renal transplantation? *Transplantation* **2000**; 70:1116–9.
- Mas V, Alvarez T, Albano S, et al. Utility of cytomegalovirus viral load in renal transplant patients in Argentina. *Transplantation* **1999**; 67:1050–5.
- Humar A, Gregson D, Caliendo AM, et al. Clinical utility of quantitative cytomegalovirus viral load determination for predicting cytomegalovirus disease in liver transplant recipients. *Transplantation* **1999**; 68:1305–11.
- Ferreira-Gonzalez A, Fisher RA, Weymouth LA, et al. Clinical utility of a quantitative polymerase chain reaction for diagnosis of cytomegalovirus disease in solid organ transplant patients. *Transplantation* **1999**; 68:991–6.
- Madi N, Al-Qaser M, Edan R, Al-Nakib W. Clinical utility of viral load in the management of cytomegalovirus infection in solid organ transplant patients in Kuwait. *Transplant Proc* **2015**; 47:1802–7.
- Emery VC, Sabin CA, Cope AV, Gor D, Hassan-Walker AF, Griffiths PD. Application of viral-load kinetics to identify patients who develop cytomegalovirus disease after transplantation. *Lancet* **2000**; 355:2032–6.
- Razonable RR, van Crujnsen H, Brown RA, et al. Dynamics of cytomegalovirus replication during preemptive therapy with oral ganciclovir. *J Infect Dis* **2003**; 187:1801–8.

46. Weinberg A, Hodges TN, Li S, Cai G, Zamora MR. Comparison of PCR, antigenemia assay, and rapid blood culture for detection and prevention of cytomegalovirus disease after lung transplantation. *J Clin Microbiol* **2000**; 38:768–72.
47. Tong CY, Cuevas LE, Williams H, Bakran A. Prediction and diagnosis of cytomegalovirus disease in renal transplant recipients using qualitative and quantitative polymerase chain reaction. *Transplantation* **2000**; 69:985–91.
48. Palmer SM, Limaye AP, Banks M, et al. Extended valganciclovir prophylaxis to prevent cytomegalovirus after lung transplantation: a randomized, controlled trial. *Ann Intern Med* **2010**; 152:761–9.
49. Humar A, Lebranchu Y, Vincenti F, et al. The efficacy and safety of 200 days valganciclovir cytomegalovirus prophylaxis in high-risk kidney transplant recipients. *Am J Transplant* **2010**; 10:1228–37.
50. Jaksch P, Zweytick B, Kerschner H, et al. Cytomegalovirus prevention in high-risk lung transplant recipients: comparison of 3- vs 12-month valganciclovir therapy. *J Heart Lung Transplant* **2009**; 28:670–5.
51. Humar A, Kumar D, Preiksaitis J, et al. A trial of valganciclovir prophylaxis for cytomegalovirus prevention in lung transplant recipients. *Am J Transplant* **2005**; 5:1462–8.
52. Paya C, Humar A, Dominguez E, et al; Valganciclovir Solid Organ Transplant Study Group. Efficacy and safety of valganciclovir vs. oral ganciclovir for prevention of cytomegalovirus disease in solid organ transplant recipients. *Am J Transplant* **2004**; 4:611–20.
53. Luan FL, Kommareddi M, Ojo AO. Impact of cytomegalovirus disease in D+/R-kidney transplant patients receiving 6 months low-dose valganciclovir prophylaxis. *Am J Transplant* **2011**; 11:1936–42.
54. Onor IO, Todd SB, Meredith E, et al. Evaluation of clinical outcomes of prophylactic versus preemptive cytomegalovirus strategy in liver transplant recipients. *Transpl Int* **2013**; 26:592–600.
55. Couzi L, Helou S, Bachelet T, et al. High incidence of anticytomegalovirus drug resistance among D+R- kidney transplant recipients receiving preemptive therapy. *Am J Transplant* **2012**; 12:202–9.
56. Witzke O, Hauser IA, Bartels M, Wolf G, Wolters H, Nitschke M; VIPP Study Group. Valganciclovir prophylaxis versus preemptive therapy in cytomegalovirus-positive renal allograft recipients: 1-year results of a randomized clinical trial. *Transplantation* **2012**; 93:61–8.
57. van der Beek MT, Berger SP, Vossen AC, et al. Preemptive versus sequential prophylactic-preemptive treatment regimens for cytomegalovirus in renal transplantation: comparison of treatment failure and antiviral resistance. *Transplantation* **2010**; 89:320–6.
58. McGillicuddy JW, Weimert NA, Taber DJ, et al. Can preemptive cytomegalovirus monitoring be as effective as universal prophylaxis when implemented as the standard of care in patients at moderate risk? *Transplantation* **2010**; 89:1218–23.
59. Khoury JA, Storch GA, Bohl DL, et al. Prophylactic versus preemptive oral valganciclovir for the management of cytomegalovirus infection in adult renal transplant recipients. *Am J Transplant* **2006**; 6:2134–43.
60. Durand CM, Marr KA, Arnold CA, et al. Detection of cytomegalovirus DNA in plasma as an adjunct diagnostic for gastrointestinal tract disease in kidney and liver transplant recipients. *Clin Infect Dis* **2013**; 57:1550–9.
61. Costa C, Curtioni A, Sidoti F, et al. Detection of human cytomegalovirus in transbronchial biopsies from lung transplant recipients. *Arch Virol* **2013**; 158:1461–5.
62. Harvala H, Stewart C, Muller K, et al. High risk of cytomegalovirus infection following solid organ transplantation despite prophylactic therapy. *J Med Virol* **2013**; 85:893–8.
63. Boillat Blanco N, Pascual M, Venetz JP, Nseir G, Meylan PR, Manuel O. Impact of a preemptive strategy after 3 months of valganciclovir cytomegalovirus prophylaxis in kidney transplant recipients. *Transplantation* **2011**; 91:251–5.
64. Kuypers DR, Claes K, Evenepoel P, Maes B, Vanrenterghem Y. A simplified strategy for clinical management of late cytomegalovirus infection after oral ganciclovir prophylaxis in renal recipients. *J Antimicrob Chemother* **2005**; 55:391–4.
65. Kute VB, Vanikar AV, Shah PR, et al. Post-renal transplant cytomegalovirus infection: study of risk factors. *Transplant Proc* **2012**; 44:706–9.
66. Walker JK, Scholz LM, Scheetz MH, et al. Leukopenia complicates cytomegalovirus prevention after renal transplantation with alemtuzumab induction. *Transplantation* **2007**; 83:874–82.
67. Vu D, Shah T, Ansari J, et al. Interferon-gamma gene polymorphism +874 A/T is associated with an increased risk of cytomegalovirus infection among Hispanic renal transplant recipients. *Transpl Infect Dis* **2014**; 16:724–32.
68. Abate D, Saldan A, Mengoli C, et al. Comparison of cytomegalovirus (CMV) enzyme-linked immunosorbent spot and CMV quantiferon gamma interferon-releasing assays in assessing risk of CMV infection in kidney transplant recipients. *J Clin Microbiol* **2013**; 51:2501–7.
69. Kielberger L, Bouda M, Jindra P, Reischig T. Pharmacoeconomic impact of different regimens to prevent cytomegalovirus infection in renal transplant recipients. *Kidney Blood Press Res* **2012**; 35:407–16.
70. Kim SI, Kim CJ, Kim YJ, et al. Antiviral prophylaxis versus preemptive therapy to prevent cytomegalovirus infection and related death in liver transplantation: a retrospective study with propensity score matching. *Transplant Proc* **2012**; 44:787–90.
71. Manuel O, Pascual M, Perrottet N, et al. Ganciclovir exposure under a 450 mg daily dosage of valganciclovir for cytomegalovirus prevention in kidney transplantation: a prospective study. *Clin Transplant* **2010**; 24:794–800.
72. Weclawiak H, Kamar N, Mengelle C, et al. Pre-emptive intravenous ganciclovir versus valganciclovir prophylaxis for de novo cytomegalovirus-seropositive kidney-transplant recipients. *Transpl Int* **2010**; 23:1056–64.
73. Smith JM, Corey L, Bittner R, et al. Subclinical viremia increases risk for chronic allograft injury in pediatric renal transplantation. *J Am Soc Nephrol* **2010**; 21:1579–86.
74. Eid AJ, Brown RA, Arthurs SK, et al. A prospective longitudinal analysis of cytomegalovirus (CMV)-specific CD4+ and CD8+ T cells in kidney allograft recipients at risk of CMV infection. *Transpl Int* **2010**; 23:506–13.
75. Park JM, Lake KD, Arenas JD, Fontana RJ. Efficacy and safety of low-dose valganciclovir in the prevention of cytomegalovirus disease in adult liver transplant recipients. *Liver Transpl* **2006**; 12:112–6.
76. Mengelle C, Rostaing L, Weclawiak H, Rossignol C, Kamar N, Izopet J. Prophylaxis versus pre-emptive treatment for prevention of cytomegalovirus infection in CMV-seropositive orthotopic liver-transplant recipients. *J Med Virol* **2015**; 87:836–44.
77. Gracia-Ahufinger I, Ferrando-Martínez S, Montejo M, et al. Pre-transplant thymic function is associated with the risk of cytomegalovirus disease after solid organ transplantation. *Clin Microbiol Infect* **2015**; 21:511.e1–7.
78. Puttarajappa C, Bhattarai M, Mour G, et al. Cytomegalovirus infection in high-risk kidney transplant recipients receiving thymoglobulin induction—a single-center experience. *Clin Transplant* **2016**; 30:1159–64.
79. Cameron BM, Kennedy SE, Rawlinson WD, Mackie FE. The efficacy of valganciclovir for prevention of infections with cytomegalovirus and Epstein-Barr virus after kidney transplant in children. *Pediatr Transplant* **2017**; 21. doi: 10.1111/ptr.12816.
80. Werzowa J, Schwaiger B, Hecking M, et al. Prophylactic CMV therapy does not improve three-yr patient and graft survival compared to preemptive therapy. *Clin Transplant* **2015**; 29:1230–8.
81. Byrns JS, Pilch NW, Taber DJ. Impact of pharmacist involvement in early identification and enrollment in patient assistance programs on CMV outcomes in transplantation. *J Pharm Pract* **2016**; 29:97–102.
82. Kaminski H, Couzi L, Garrigue I, Moreau JF, Déchanet-Merville J, Merville P. Easier control of late-onset cytomegalovirus disease following universal prophylaxis through an early antiviral immune response in donor-positive, recipient-negative kidney transplants. *Am J Transplant* **2016**; 16:2384–94.
83. Kamar N, Mengelle C, Esposito L, et al. Predictive factors for cytomegalovirus reactivation in cytomegalovirus-seropositive kidney-transplant patients. *J Med Virol* **2008**; 80:1012–7.
84. Reischig T, Kacer M, Jindra P, Hes O, Lysak D, Bouda M. Randomized trial of valganciclovir versus valacyclovir prophylaxis for prevention of cytomegalovirus in renal transplantation. *Clin J Am Soc Nephrol* **2015**; 10:294–304.
85. Martín-Gandul C, Pérez-Romero P, Blanco-Lobo P, et al; Spanish Network for Research in Infectious Diseases (REIPI). Viral load, CMV-specific T-cell immune response and cytomegalovirus disease in solid organ transplant recipients at higher risk for cytomegalovirus infection during preemptive therapy. *Transpl Int* **2014**; 27:1060–8.
86. Reischig T, Jindra P, Hes O, Svecová M, Klaboch J, Treska V. Valacyclovir prophylaxis versus preemptive valganciclovir therapy to prevent cytomegalovirus disease after renal transplantation. *Am J Transplant* **2008**; 8:69–77.
87. Atabani SF, Smith C, Atkinson C, et al. Cytomegalovirus replication kinetics in solid organ transplant recipients managed by preemptive therapy. *Am J Transplant* **2012**; 12:2457–64.
88. Müller V, Perrakis A, Meyer J, et al. The value of pre-emptive therapy for cytomegalovirus after liver transplantation. *Transplant Proc* **2012**; 44:1357–61.
89. Sun HY, Cacciarelli TV, Wagener MM, Singh N. Preemptive therapy for cytomegalovirus based on real-time measurement of viral load in liver transplant recipients. *Transpl Immunol* **2010**; 23:166–9.
90. Pillet A, Mengelle C, Basse G, et al. Monitoring HCMV infection with quantitative real-time PCR in HCMV-positive orthotopic liver transplant recipients, and predictive factors for treatment of the first episode of HCMV viremia. *Transplant Proc* **2006**; 38:2335–8.
91. Wadhawan M, Gupta S, Goyal N, et al. Cytomegalovirus infection: its incidence and management in cytomegalovirus-seropositive living related liver transplant recipients: a single-center experience. *Liver Transpl* **2012**; 18:1448–55.
92. Watcharananan SP, Louhapanawat S, Chantratita W, Jirasiritham S, Sumethkul V. Cytomegalovirus viremia after kidney transplantation in Thailand: predictors of symptomatic infection and outcome. *Transplant Proc* **2012**; 44:701–5.

93. Mossad SB. Preventing CMV viremia and disease 3–6 months after renal transplantation. *Am J Transplant* **2010**; 10:2184; author reply 2185.
94. Hammond SP, Martin ST, Roberts K, et al. Cytomegalovirus disease in lung transplantation: impact of recipient seropositivity and duration of antiviral prophylaxis. *Transpl Infect Dis* **2013**; 15:163–70.
95. Smallwood GA, de Vera ME, Davis L, Martinez E, Stieber AC, Heffron TG. Preemptive ganciclovir for CMV viremia in liver transplantation. *Transplant Proc* **2001**; 33:1814–5.
96. Aldridge RW, Mattes FM, Rolando N, et al. Effects of donor/recipient human leukocyte antigen mismatch on human cytomegalovirus replication following liver transplantation. *Transpl Infect Dis* **2015**; 17:25–32.
97. Sood S, Haifer C, Yu L, et al. Targeted individual prophylaxis offers superior risk stratification for cytomegalovirus reactivation after liver transplantation. *Liver Transpl* **2015**; 21:1478–85.
98. Caston JJ, Castells L, Varo E, et al. Impact of cytomegalovirus infection on severe hepatitis C recurrence in patients undergoing liver transplantation. *Transplantation* **2016**; 100:593–9.
99. Schneider M, Mاتيقي T, Kundi M, et al. Clinical significance of the single nucleotide polymorphism TLR2 R753Q in heart transplant recipients at risk for cytomegalovirus disease. *J Clin Virol* **2016**; 84:64–9.
100. Leone F, Gigliotti P, Mauro MV, et al. Early cytomegalovirus-specific T-cell response and estimated glomerular filtration rate identify patients at high risk of infection after renal transplantation. *Transpl Infect Dis* **2016**; 18(2):191–201.
101. Asberg A, Humar A, Rollag H, et al; VICTOR Study Group. Oral valganciclovir is noninferior to intravenous ganciclovir for the treatment of cytomegalovirus disease in solid organ transplant recipients. *Am J Transplant* **2007**; 7:2106–13.
102. Razonable RR, Åsberg A, Rollag H, et al. Virologic suppression measured by a cytomegalovirus (CMV) DNA test calibrated to the World Health Organization international standard is predictive of CMV disease resolution in transplant recipients. *Clin Infect Dis* **2013**; 56:1546–53.
103. Fryer JF, Heath AB, Minor PD; Collaborative Study Group. A collaborative study to establish the 1st WHO International Standard for human cytomegalovirus for nucleic acid amplification technology. *Biologicals* **2016**; 44:242–51.
104. Durier N, Ananworanich J, Apornpong T, et al. Cytomegalovirus viremia in Thai HIV-infected patients on antiretroviral therapy: prevalence and associated mortality. *Clin Infect Dis* **2013**; 57:147–55.
105. Fielding K, Koba A, Grant AD, et al. Cytomegalovirus viremia as a risk factor for mortality prior to antiretroviral therapy among HIV-infected gold miners in South Africa. *PLoS One* **2011**; 6:e25571.
106. Foster GR, Afdhal N, Roberts SK, et al; ASTRAL-2 Investigators; ASTRAL-3 Investigators. Sofosbuvir and velpatasvir for HCV genotype 2 and 3 infection. *N Engl J Med* **2015**; 373:2608–17.
107. Feld JJ, Jacobson IM, Hézode C, et al; ASTRAL-1 Investigators. Sofosbuvir and velpatasvir for HCV genotype 1, 2, 4, 5, and 6 infection. *N Engl J Med* **2015**; 373:2599–607.

## APPENDIX I: MEMBERS OF THE CMV FORUM

Rekha Abichandani, MD, MS; Shire  
 Barbara Alexander, MD, MHS; Duke University Medical Center  
 Robin Avery, MD, FIDSA; Johns Hopkins School of Medicine  
 Fausto Baldanti, MD; Fondazione IRCCS Policlinico San Matteo  
 Susan Barnett, PhD; GlaxoSmithKline  
 Paul Baum, MD, PhD; Roche Molecular Diagnostics  
 M. Michelle Berrey, MD, MPH; Chimerix, Inc.  
 Debra Birnkrant, MD; US Food and Drug Administration  
 Emily Blumberg, MD; Hospital of the University of Pennsylvania  
 Michael Boeckh, MD, PhD; Fred Hutchison Cancer Research Center/University of Washington  
 David Boutolleau, PharmD, PhD; Université Pierre et Marie Curie & Hôpitaux Universitaires La Pitié-Salpêtrière–Charles Foix  
 Terry Bowlin, PhD; Microbiotix, Inc  
 Jennifer Brooks, MS, MSN; Microbiotix, Inc  
 Roy Chemaly, MD, MPH, FACP, FIDSA; University of Texas MD Anderson Cancer Center

Sunwen Chou, MD; Oregon Health & Science University  
 Gavin Cloherty, PhD; Abbott Diagnostics  
 William Cruikshank, PhD; Oxford Immunotec  
 Lesia Dropulic, MD; National Institutes of Health  
 Hermann Einsele, MD; University of Würzburg  
 Jay Erdman, MS; Astellas Pharma, Inc.  
 Gary Fahle; National Institutes of Health  
 Lynn Fallon, BS, RN; CTI Clinical Trial & Consulting Services  
 Heather Gillis; Oxford Immunotec  
 Dimitri Gonzalez, MS; Advanced Biological Laboratories  
 Paul Griffiths, MD, DSc, FRCPath; University College London Medical School  
 Kurt Gunter, MD, FASCP; Cell Medica, Inc  
 Hans Hirsch, MD, MSc; Universität Basel  
 Aimee Hodowanec, MD; US Food and Drug Administration  
 Atul Humar, MD, MSc, FRCPC; Toronto General Hospital  
 Peter Hunt, MD; University of California, San Francisco  
 Filip Josephson, MD, PhD; Swedish Medical Products Agency  
 Takashi Komatsu, PhD; US Food and Drug Administration  
 Camille Kotton, MD, FIDSA; Harvard Medical School and Massachusetts General Hospital  
 Philip Krause, MD; US Food and Drug Administration  
 Frank Kuhr, PhD; Qiagen, Inc  
 Christopher Lademacher, MD, PhD; Astellas Pharma  
 Randall Lanier, PhD; Chimerix, Inc  
 Tadd Lazarus, MD; Qiagen  
 John Leake, MD, MPH; Quest Diagnostics  
 Randi Leavitt, MD, PhD; Merck Research Laboratories  
 Sandra Nusinoff Lehrman, MD; Merck Research Labs  
 Li Li, MS; US Food and Drug Administration  
 Per Ljungman, MD, PhD; Karolinska Institutet and Karolinska University Hospital  
 Paula Isabelle Lodding; University of Copenhagen  
 Jens Lundgren, MD, DMSc; Rigshospitalet, University of Copenhagen  
 Francisco (Paco) Martinez-Murillo, PhD; US Food and Drug Administration  
 Howard Mayer, MD; Shire  
 Megan McCutcheon, BS; Oxford Immunotec  
 John McKinnon, MD, MSc; Henry Ford Hospital, Wayne State University  
 Thomas Mertens, MD; Ulm University and University Clinic of Ulm  
 Veronica Miller, PhD; Forum for Collaborative Research  
 Kevin Modarress, PhD; Qiagen  
 Johann Mols; GlaxoSmithKline  
 Sally Mossman, PhD; GlaxoSmithKline Vaccines  
 Yoshihiko Murata, MD, PhD; Merck  
 David Murawski; Quest Diagnostics  
 Jeffrey Murray, MD, MPH; US Food and Drug Administration  
 Yoichiro Natori, MD; University of Toronto

Garrett Nichols, MD, MS; Chimerix, Inc  
Jules O'Rear, PhD; US Food and Drug Administration  
Karl Peggs, MBBCh, MA, MRCP, FRCPath; University College  
London Medical School & Cancer Institute  
Andreas Pikis, MD; US Food and Drug Administration  
Mark Prichard, PhD; University of Alabama, Birmingham  
School of Medicine  
Raymund Razonable, MD; Mayo Clinic  
Marcie Riches, MD, MS; Moffitt Cancer Center

Jeff Roberts, MD; US Food and Drug Administration  
Wael Saber, MD, MS; Medical College of Wisconsin and Center  
for International Blood & Marrow Transplant Research  
Chalom Sayada, MD, PhD; Advanced Biological Laboratories, Inc  
Mary Singer, MD, PhD; US Food and Drug Administration  
Thomas Stamminger, MD; University of Erlangen–Nuremberg  
Anna Wijatyk, MD; Shire  
Dong Yu, PhD; Novartis Vaccines and Diagnostics, Inc  
Bernhardt Zeiher, MD; Astellas Pharma Global Development, Inc