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1 **Synthesis, cytotoxicity, antioxidant activity and molecular modeling**
2 **of new NSAIDs-EBS derivatives**

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1 **Abstract:**

2 Two series of NSAIDs-EBS derivatives (**5a-j** and **9a-i**) based on the
3 hybridization of nonsteroidal anti-inflammatory drugs (NSAIDs) skeleton and
4 Ebselen moiety were synthesized. Their cytotoxicity was evaluated against five types
5 of human cancer cell lines, BGC-823 (human gastric cancer cell line), SW480 (human
6 colon adenocarcinoma cells), MCF-7 (human breast adenocarcinoma cells), HeLa
7 (human cervical cancer cells), A549 (human lung carcinoma cells). Moreover, the
8 most active compound **5j** showed IC₅₀ values below 3 μM in all cancer cell lines and
9 with remarkable anticancer activity against MCF-7 (1.5 μM) and HeLa (1.7 μM). The
10 redox properties of the NSAIDs-EBS derivatives prepared herein were conducted by
11 2, 2-didiphenyl-1-picrylhydrazyl (DPPH), bleomycin dependent DNA damage and
12 glutathione peroxidase (GPx)-like assays. Finally, TrxR1 inhibition activity assay and
13 molecular docking study revealed NSAIDs-EBS derivatives could serve as potential
14 TrxR1 inhibitor.

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18 **Keywords:** Selenium; NSAIDs; Ebselen; Anticancer; Molecular modeling

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1 **Introduction**

2 Non-steroidal anti-inflammatory drugs (NSAIDs) are a class of medications
3 commonly used to treat symptoms of inflammatory diseases such as osteoarthritis and
4 rheumatoid arthritis, and are routinely been used across the world [1, 2]. Moreover,
5 numerous evidence from epidemiological and preclinical studies have shown that
6 NSAIDs used in combination with different therapies, such as chemotherapy and
7 immunotherapy or even radiation, represented an attractive strategy to increase
8 anticancer efficacy and reduce toxicity [3-6]. Based on the fact that NSAIDs display
9 their anticancer activities, the chemical modifications of their structures have
10 demonstrated stronger cytotoxicity and chemo-preventive than corresponding NSAID
11 alone [7, 8]. NSAIDs framework modification has become a structure-based
12 medicinal chemistry strategy to design novel anticancer agents in the past decades [9-
13 12].

14 Selenium is an essential trace mineral nutrient with multiple roles in the growth
15 and function of living animal cells, and it effectively inhibits tumorigenesis in both
16 animal models and epidemiological studies. Twenty-five selenoproteins in the human
17 body exert specific biological functions. Selenium compounds have attracted huge
18 interest in the past decades as chemotherapeutic and chemo-preventive agents.
19 Several epidemiological studies have reported an inverse association between the
20 nutritional selenium status and cancer risk. Specifically, selenazo compounds have
21 received great attention owing to their chemical properties, pharmaceutical
22 applications, and low toxicity [13-15]. Ebselen (EBS, **Fig 1**) is the most potential
23 compound in cancer prevention, some EBS-related compounds have been reported to
24 exhibit anticancer activity (**Fig 1**) [16-21].

25 In previous study, we have reported the synthesis of a series of novel NSAIDs-
26 Selenium derivatives and screened their anticancer activity by *in vitro* study, the
27 modification of NSAIDs scaffolds with Se functionalities (-SeCN, -Se-Se-, -SeCF₃)
28 demonstrated potent inhibition of human tumor cell [22-25]. Because the
29 pharmacological effects of Ebselen, including antioxidant and anticancer activities,
30 twenty new NSAIDs-EBS derivatives were designed by the fusion of NSAIDs

1 fragment and Ebselen in a new molecule [Fig 2]. The hybrid compounds were
 2 evaluated for anticancer activities *in vitro*. Furthermore, the antioxidant potential of
 3 the compounds was investigated by employing DPPH, bleomycin-dependent DNA
 4 damage and GPx-like assays. Finally, Thioredoxin Reductase (TrxR1) inhibition
 5 activity assay and molecular docking study (TrxR1 as docking protein) were
 6 performed, in order to predict the target and anticancer activity of the prepared
 7 NSAIDs-EBS hybrid compounds.

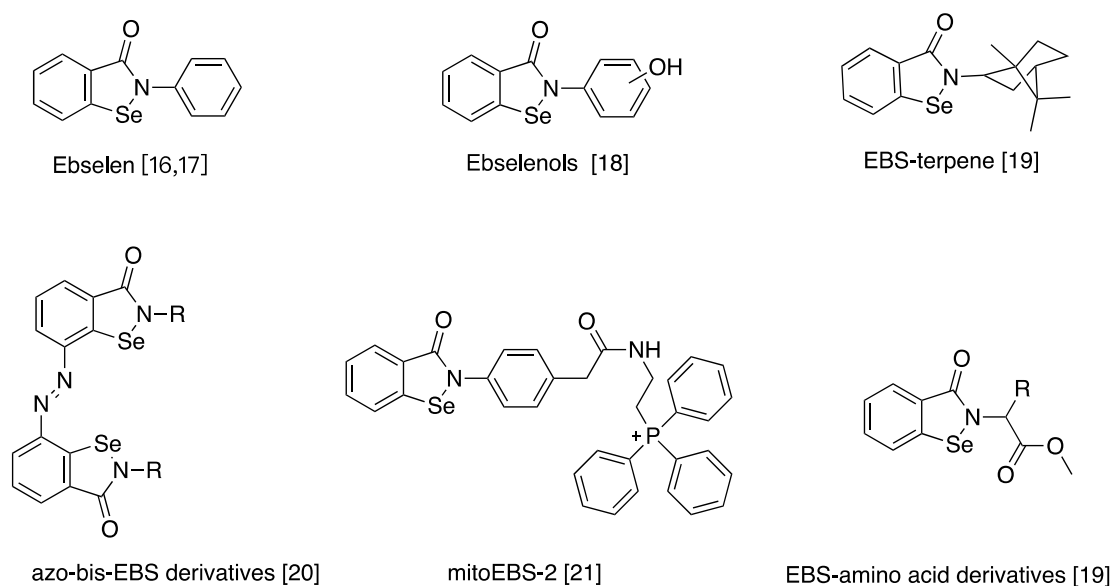


Fig. 1. EBS-related compounds with anticancer activity

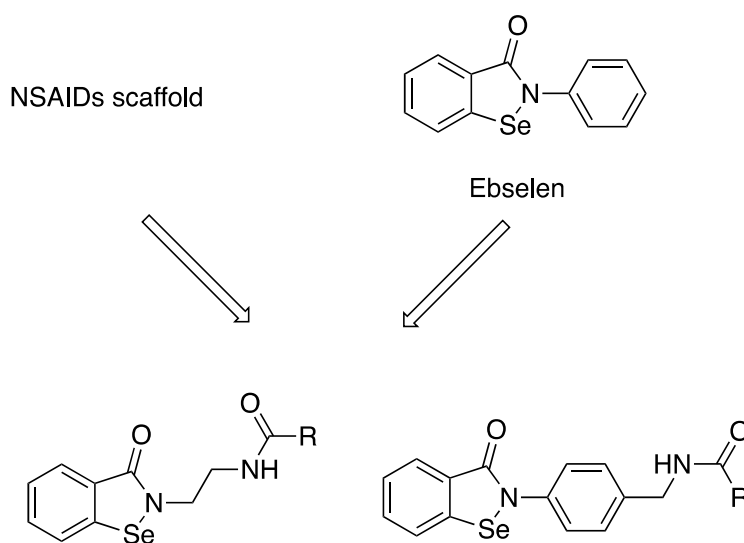


Fig. 2. NSAIDs-EBS derivatives

2. Results and Discussion

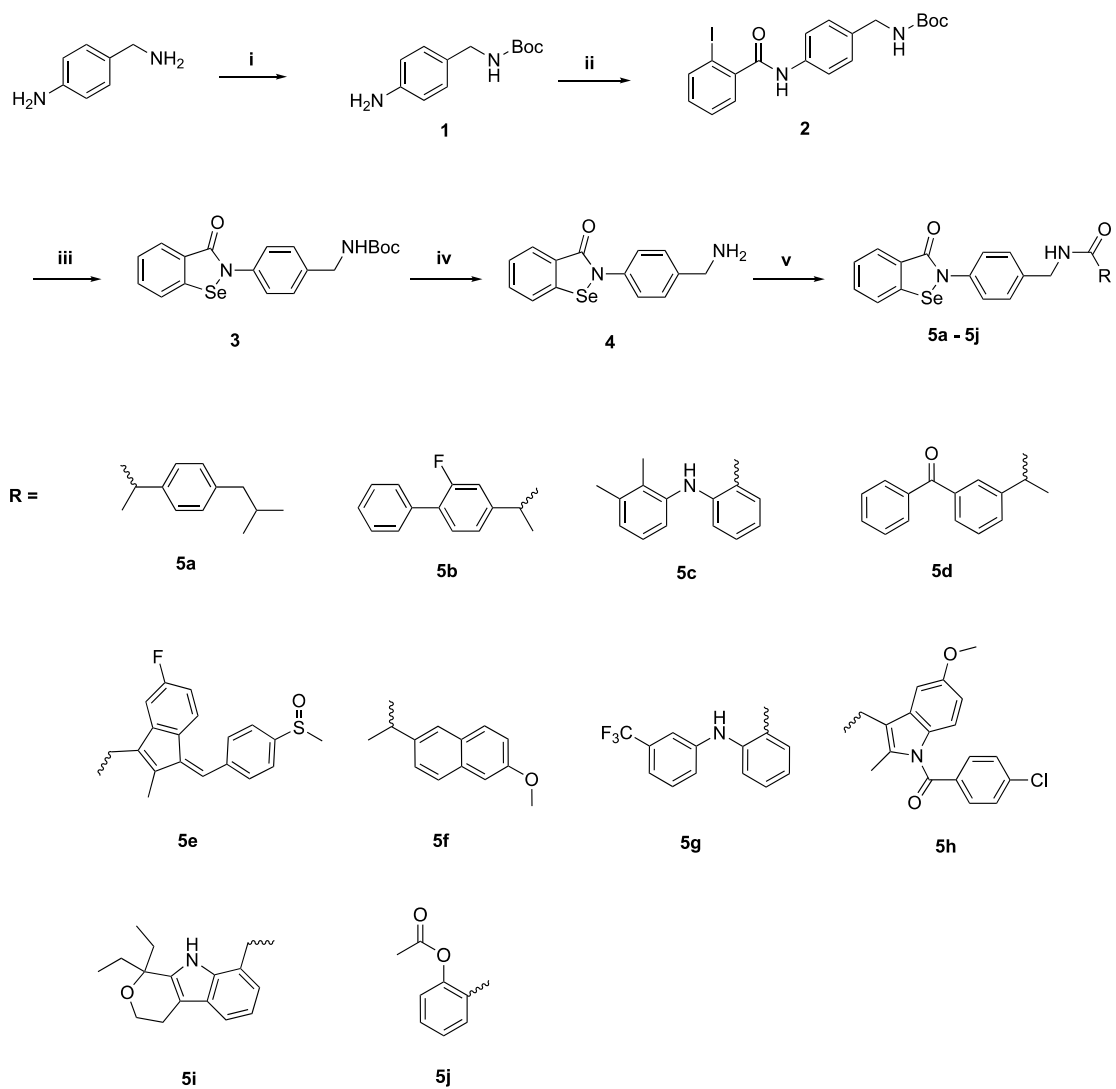
1 **2.1 Chemistry**

2 The synthesis strategies of compounds **5a-j** are outlined in Scheme 1. First,
3 4-aminobenzylamine reacted with di-tert-butyl dicarbonate to give intermediate
4 **1**. The reactions of **1** with o-iodobenzoyl chloride in the present of TEM
5 provided compound **2**. The reaction of **2** with KSeCN and following
6 deprotection produced the EBS intermediate **4**. Finally, the target products **5a-j**
7 were obtained by reacting compound **4** with commercially available NSAIDs in
8 the present of EDCI and HOBt as condensation agent (**Scheme 1**).

9 The synthesis of target compounds **9a-i** is shown in Scheme 2. The
10 reaction of o-iodobenzoyl chloride with tert-butyl (2-aminoethyl) carbamate
11 afforded the intermediate **6**. Compound **6** was reacted with KSeCN and
12 following deprotection produced the EBS intermediate **8**. Finally, the target
13 products **9a-i** were obtained by reacting compound **8** with commercially available
14 NSAIDs in the present of EDCI and HOBt as condensation agent (**Scheme 2**).

15 The purity of all final compounds was 95% or higher and their chemical
16 structures were characterized using ^1H NMR, ^{13}C NMR and HRMS (ESI).

17



1

2 **Scheme 1.** i) $(\text{Boc})_2\text{O}$, THF, 0-25 °C; ii) TEA, DCM, 25 °C; iii) CuI, Cs_2CO_3 ,

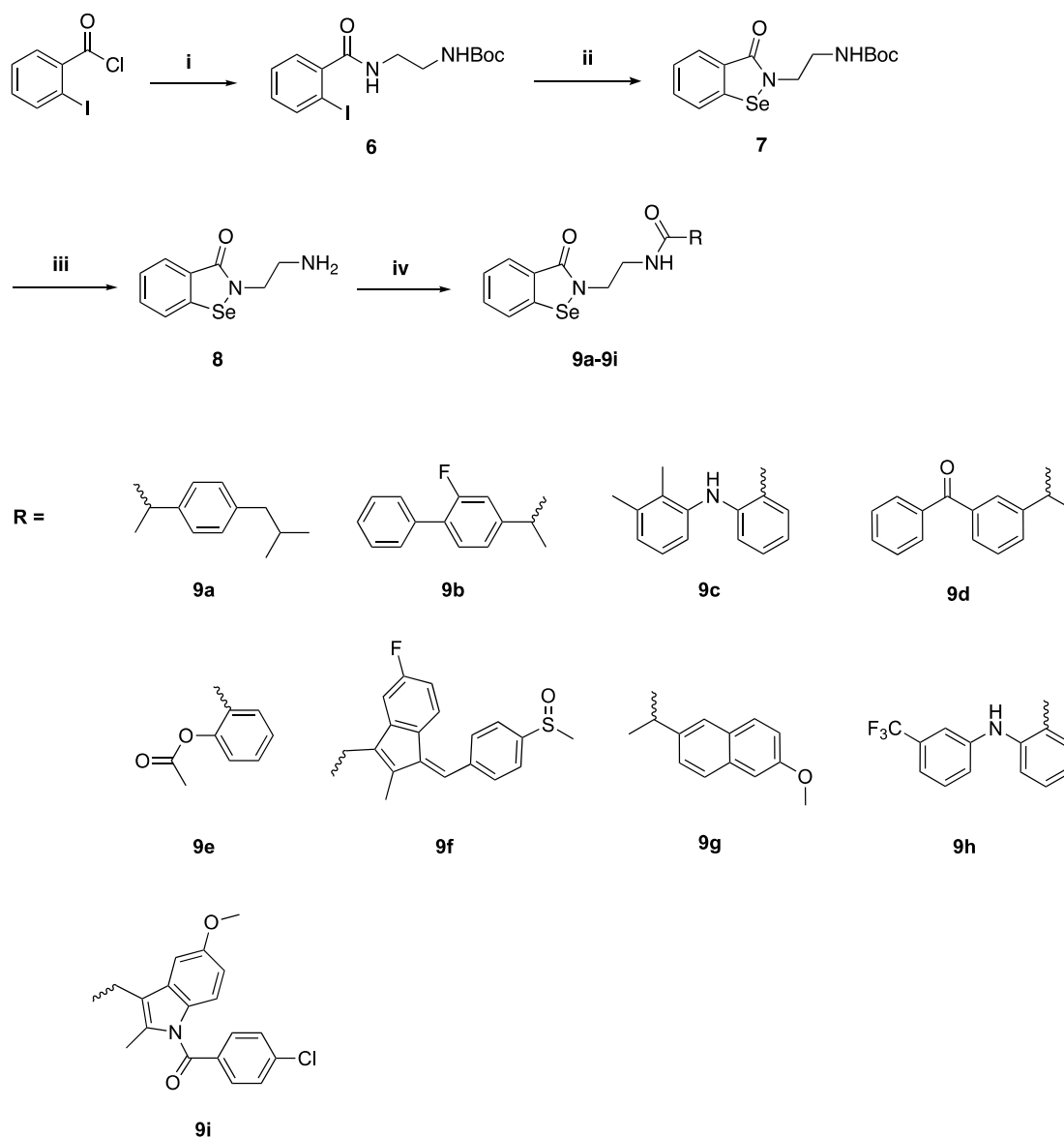
3 KSeCN, 1,10-phenanthroline, DMF, 100 °C; iv) TFA, DCM, r. t.; v) NSAIDs, EDCI,

4 HOBT, TEA, DCM.

5

6

1



2

3 **Scheme 2.** i) N-Boc-ethylenediamine, TEA, DCM, 0°C; ii) CuI, Cs₂CO₃, KSeCN,
4 1,10-phenanthroline, DMF, 100 °C; iii) TFA, DCM, 25 °C; iv) NSAIDs, EDCI,
5 HOBT, TEA, DCM, 25 °C.

6

7 2.2. Cytotoxicity

8 MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay was
9 conducted to evaluate the potential antiproliferative activities against human tumor
10 cell lines derived from various human cancer types: BGC-823 (human gastric cancer
11 cell line), SW480 (human colon adenocarcinoma cells), MCF-7 (human breast

1 adenocarcinoma cells), HeLa (human cervical cancer cells), A549 (human lung
2 carcinoma cells) of target compounds **5a-j** and **9a-i**, doxorubicin was selected as
3 reference standard (Table 1).

4 As shown in **Table 1**, most of the NSAIDs-EBS derivatives exhibited good
5 antiproliferative activity with IC₅₀ values at micromolar level, while the selected
6 patent NSAIDs (Aspirin, Ibuprofen and Naproxen) and Ebselen are inactive against
7 all cells even in the maximum dose of 50 μM. The IC₅₀ values obtained for the
8 NSAIDs-EBS derivatives showed that the fusion of NSAIDs scaffold and ebselen
9 moiety in a new molecule result in the significant effect on cancer cell line.

10 The results in table **1** showed that the cytotoxic activity of NSAIDs-EBS
11 derivatives containing a phenyl moiety between two nitrogen atom (**5a-i**) is better
12 than corresponding NSAIDs-EBS derivatives containing ethyl moiety (**9a-i**),
13 considering the lipophilicity and electron withdrawing effect.

14 Furthermore, the cytotoxic activity of compounds **5a**, **5b**, **5c**, **5d**, **5e**, **5g**, **5h**, **5i**
15 and **5j** displayed IC₅₀ values below 10 μM against BGC-823 cells. The most active
16 compounds of these two series are **5c** and **5j**. These two compounds show IC₅₀ values
17 below 5 μM in all of tested cancer cell lines. Compound **5j** emerges the most potent
18 agent with IC₅₀ values below 3 μM in all cancer cell lines and with remarkable
19 anticancer activity against MCF-7 (1.5 μM) and HeLa (1.7 μM).

20

1 **Table 1.** Cytotoxic activity expressed by IC₅₀ of NSAIDs-EBS derivatives (**5a-j** and
 2 **9a-i**) on different cancer cell lines.

Compound	IC ₅₀ (μ M) ^a				
	BGC-823	SW-480	MCF-7	HeLa	A549
Aspirin ^b	>50	>50	>50	>50	>50
Ibuprofen ^b	>50	>50	>50	>50	>50
Naproxen ^b	>50	>50	>50	>50	>50
Ebselen ^b	>50	>50	>50	>50	>50
5a	7.5±0.7	10.4±1.1	12.4±1.2	9.3±0.8	8.1±0.8
5b	6.5±0.4	8.3±0.8	11.3±1.1	9.5±0.9	12.3±1.2
5c	3.4±0.2	4.5±0.4	4.2±0.3	3.8±0.2	3.6±0.2
5d	11.2±1.2	12.5±1.2	8.4±0.8	9.2±0.9	13.2±1.3
5e	9.5±0.9	9.8±0.9	7.4±0.7	8.4±0.8	8.3±0.8
5f	13.5±1.3	7.4±0.7	13.8±1.3	9.7±0.9	11.4±1.1
5g	8.4±0.8	13.7±1.2	14.2±1.1	7.5±1.3	12.7±1.2
5h	9.2±0.9	7.5±0.7	11.3±1.1	14.3±1.4	10.3±1.0
5i	9.8±1.5	12.3±1.2	8.8±0.7	12.6±1.3	9.8±0.9
5j	2.4±0.2	2.8±0.2	1.5±0.1	1.7±0.1	2.1±0.2
9a	12.5±1.2	11.4±1.1	15.7±1.5	10.6±1.1	9.4±0.8
9b	9.5±0.6	11.3±1.1	14.9±1.4	10.4±1.0	13.6±1.3
9c	6.5±0.7	7.2±0.7	5.3±0.5	8.8±0.8	5.2±0.5
9d	13.6±1.4	14.6±1.5	10.3±1.0	11.3±1.1	15.6±1.5
9e	11.4±1.1	14.2±1.4	14.5±1.4	12.7±1.2	10.6±1.0
9f	16.6±1.6	10.4±1.0	14.4±1.4	11.3±1.1	13.3±1.3
9g	13.5±1.3	15.7±1.5	16.5±1.6	8.8±0.8	14.7±1.4
9h	11.4±1.1	9.4±0.9	12.6±1.2	15.2±1.5	12.2±1.2
9i	17.4±1.7	14.3±1.4	10.3±1.0	13.6±1.3	11.2±1.1
Doxorubicin^c	9.8±0.9	12.4±1.2	12.8±1.2	11.5±1.1	9.4±0.9

3 ^a IC₅₀ values are indicated as the mean ± SD (standard error) of at least three
 4 independent experiments.

5 ^b Patent NSAIDs and Ebselen.

6 ^c Standard benchmark compound.

7

8 2.3. Antioxidant activity

9 Reactive Oxygen Species (ROS) is actually a collective term that is used to
 10 describe oxygen-derived small and highly reactive molecules, such as superoxide
 11 anion (O₂⁻), hydroxyl radical (OH·), peroxy radical (ROO·) and alkoxy radical (RO·)
 12 [26]. ROS play essential roles in altering protein structure, thereby changing its
 13 function and participate in many pathological processes [27]. Various human diseases,
 14 including different types of cancer, are associated with a disturbed intracellular redox
 15 balance and oxidative stress (OS) [28].

1 Owing to the fact that a number of synthetic organoselenium compounds have
2 been synthesized for their use as redox-modulators in the last few years [29], the
3 antioxidant activity of compounds (**5c**, **5j**) are further estimated employing different
4 biochemical assays such as DPPH, bleomycin-dependent DNA damage and Gpx-like
5 assays [30].

6 7 2.3.1. Radical scavenging capacity (DPPH) assay.

8 The DPPH chemical assay is considered to be the rapid tools to evaluate the
9 radical-scavenging capability of organic selenides [31]. The antioxidant activity of a
10 compound is checked by its ability to decolorize DPPH radical (purple color in
11 methanol) to DPPHH (colorless) and the corresponding radical-scavenging activity is
12 estimated by the decrease in the absorbance at 517 nm [32]. Vitamin C was used as a
13 positive control (**Table 2**). Antioxidant activity was calculated as follows:

14
$$\% \text{ Antioxidant activity} = [(\text{control absorbance} - \text{sample absorbance}) / \text{control}$$

15
$$\text{absorbance}] \times 100\%$$

16 As shown in **Table 2**, NSAIDs-EBS derivatives **5f** and **5h** were the most active
17 compounds in this assay, demonstrating a good free-radical scavenging activity
18 compared to Vitamin C.

19 20 2.3.2. Bleomycin DNA damage assay.

21 Bleomycin (BLM) is a radiomimetic antitumor antibiotic first isolated
22 from *Streptomyces verticillus* [33]. BLM is widely used in clinical chemotherapy for
23 the treatment of different types of cancer, namely testicular cancer, lymphoma, lung
24 cancer, cervical cancer and cancers of the head and neck [34-35]. The bleomycin-iron
25 DNA damage assay has been routinely used as a preliminary method to test potential
26 of drugs and organic selenium compound [36]. As shown in **Table 2**, compounds **5a**,
27 **5h**, **5j** and **9g** induced DNA degradation significantly more than other tested
28 compounds.

1 **Table 2** Redox modulation activity of NSAID-EBS derivatives.

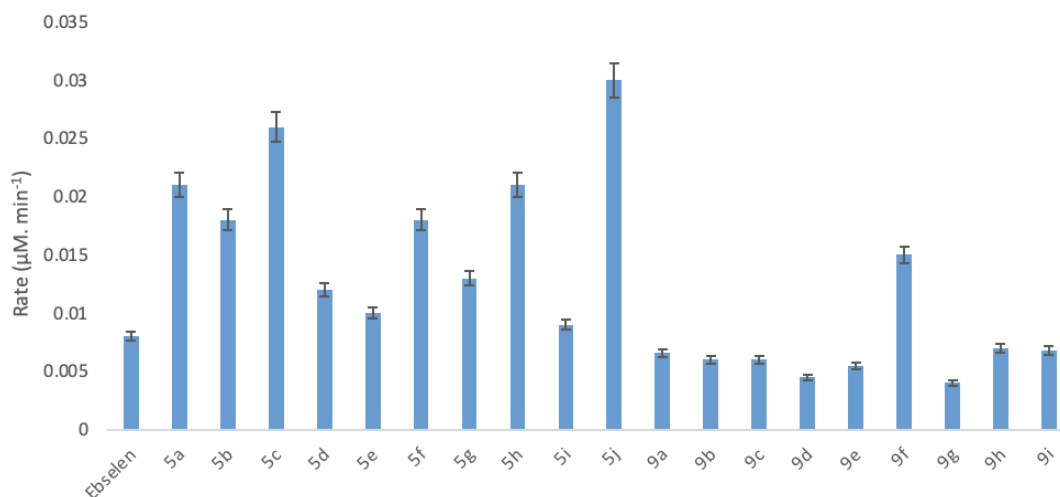
Compd. No.	DPPH		Bleomycin-dependent DNA damage
	Inhibition %	Fold	assay Absorbance
Vitamin C	97.2±1.3	1	292±2.73
5a	42.6±2.3	0.4	106.3±0.43
5b	58.2±3.6	0.6	57.3±0.33
5c	54.3±4.6	0.5	72.4±0.33
5d	31.3±2.9	0.3	85.3±1.67
5e	42.9±2.1	0.4	72.4±0.52
5f	68.6±2.7	0.7	82.8±0.84
5g	51.5±1.2	0.5	56.1±0.41
5h	78.7±3.3	0.8	101.3±1.51
5i	52.1±4.3	0.5	97.4±1.45
5j	57.3±3.1	0.5	108.3±0.39
9a	33.5±2.1	0.3	68.4±1.32
9b	42.5±2.4	0.4	85.7±2.12
9c	37.4±2.1	0.4	72.4±1.33
9d	33.3±1.6	0.3	87.6±1.20
9e	41.4±2.2	0.4	91.4±1.27
9f	29.0±1.0	0.3	68.4±1.33
9g	28.6±2.6	0.3	123.1±2.47
9h	41.7±2.0	0.4	77.7±1.32
9i	27.3±2.3	0.3	92.4±1.26

2

3 2.3.3. Glutathione peroxidase-like activity.

4 Glutathione peroxidase (GPx) is a selenoprotein that protects cells by catalyzing
5 the reduction of peroxides with the stoichiometric reductant glutathione (GSH) [37].
6 The GPx activity of NSAIDs-EBS derivatives was estimated by the decrease in
7 absorbance (340 nm) due to the oxidation of NADPH to NADP⁺. Ebselen was used as
8 the positive control.

1 The results shown in **Fig. 3** indicated that compounds **5a**, **5b**, **5c**, **5f**, **5h**, **5j** and
 2 **9f** displayed a GPx-like activity better than other derivatives. Compounds **9a-i**, for
 3 which the linker is ethyl group between NSAIDs fragment and Ebselen moiety,
 4 exhibited weaker GPx-like activity than that of phenyl group as linker except
 5 compound **9f**. Compound **5j** was the most active derivatives in this assay, up to 3 fold
 6 to the GPx mimetic ebselen.



7
 8 **Fig. 3.** GPx-like activity assay of NSAID-EBS hybrid compounds in μM. Min⁻¹.

9

10 2.4. TrxR1 inhibition activity.

11 The principle of enzyme inhibition experiment is that DTNB [5,5 '-dithiobis - (2-
 12 nitrobenzoic acid)] is one of the substrates of TrxR1. In vitro experiments, DTNB will
 13 be decomposed into TNB (2-nitro-5-thiobenzoic acid) by TrxR1 in the presence of
 14 NADPH. TNB has specific UV absorption at 412 nm. Therefore, the reaction rate of
 15 enzymatic decomposition reaction can be reflected by measuring the UV absorption at
 16 412 nm per unit time of the reaction system, and then measuring enzyme activity [38].
 17 Auranofin was used as the positive control.

18 As shown in Table 3, all NSAIDs-EBS derivatives exhibited strong inhibition
 19 against TrxR1, compounds **5a**, **5b**, **5c**, **5d**, **5e**, **5f**, **5g**, **5h**, **5i** and **5j** shown EC₅₀ values
 20 below 20 nm, which is better than Auranofin. The result showed compound **5a-j** have
 21 potential candidate as inhibitor of TrxR1.

22

1 **Table 3 TrxR1 inhibition activity of NASIDs-EBS derivatives**

Compd.	TrxR1 EC ₅₀
No.	(μ M)
5a	17.2 \pm 1.38
5b	11.2 \pm 1.87
5c	14.3 \pm 0.85
5d	19.6 \pm 1.72
5e	10.4 \pm 1.42
5f	14.6 \pm 1.34
5g	11.5 \pm 1.21
5h	15.7 \pm 1.33
5i	17.1 \pm 2.31
5j	8.8 \pm 0.43
9a	23.5 \pm 3.11
9b	28.5 \pm 2.82
9c	26.6 \pm 2.23
9d	23.6 \pm 1.25
9e	21.4 \pm 2.21
9f	27.0 \pm 1.34
9g	28.4 \pm 2.61
9h	24.8 \pm 2.53
9i	24.5 \pm 2.42
Auranofin	22.4 \pm 1.62

2

3 2.5. Docking Studies

4 The binding mode between organoselenium compounds and Mammalian TrxR1
5 protein was described by docking studies. TrxR1 consists of several functional
6 domains, including FAD and NAD binding domains at the N-terminal, and the
7 dimerization interface domain at the flexible C-terminal side [39-41]. It has been
8 reported that flexible docking can simulate the interaction between small molecules
9 and TrxR1^[4]. Therefore, compounds **5c** and **5j** were docked into the TrxR1 protein
10 (PDB id: 1H6V) using Flexible Docking Protocol as reported in the literature [42].

1 The distances between the selenium atom of all two compounds and Cys497/Cys498
 2 of TrxR1 were measured and focused on because it is closely related to the
 3 accessibility of cysteine thiol attack selenides. These compounds showed acceptable
 4 docking results (**Table 4-5**).

5 For compound **5j**, pose 4 showed a good docking conformation with the
 6 relatively high value of -CDOCKER energy (12.562 kcal/mol) and a relatively close
 7 distance between the selenium atom and Cys498 (9.773 Å) (**Table 5**, Pose 4). This
 8 good conformation may be related to the key hydrogen bond interaction between the
 9 oxygen of acetyl group and SER483 (2.29 Å). In addition, hydrogen bonding between
 10 the oxygen of 3-oxobenzod[1,2]selenazol-2(3H)-yl)benzyl and TRP407 (2.00 Å) is
 11 also important (**Figure 5**). Although the pose 5 of compound **5c** showed no hydrogen
 12 bond, it had an acceptable value of -CDOCKER energy (7.350 kcal/mol) and distance
 13 between the selenium atom and Cys498 (6.614 Å). There were many hydrophobic
 14 interactions, including hydrophobic (Pi-Alkyl) between two different benzene rings
 15 and CYS498 (5.141 Å, 5.137 Å), hydrophobic (Alkyl) between methyl groups on
 16 benzene ring and CYS498 (4.190 Å), and hydrophobic (Pi-Alkyl) between phenyl of
 17 3-oxobenzod[1,2]selenazol-2(3H)-yl)benzyl and LEU409 (5.469 Å) (**Table 4**, Pose
 18 **5; Figure 4**).

19 **Table 4** Ligand-protein poses for compound **5c**

Pose Index	CDOCKER_ENERGY	CDOCKER_INTERRATIO ENERGY	LibDock Score	LibDock Pose	Distance Cys497Se	Distance Cys498Se
1	13.2384	45.4329	61.7485	7	22.446	17.008
2	10.4149	43.5291	66.0141	6	20.851	22.024
3	9.7976	42.6044	62.2043	8	12.735	5.175
4	8.60531	40.4135	69.1193	2	22.848	20.817
5	7.34978	39.2558	87.5773	4	8.561	6.614
6	7.09795	41.2406	72.2109	5	11.294	8.087
7	7.06524	40.4968	83.4649	5	11.488	8.983
8	6.99063	38.2927	53.7714	7	11.280	14.775
9	6.84262	41.0232	69.3691	7	11.386	7.247
10	6.65588	39.1307	96.9686	1	14.168	8.893
11	6.35112	38.4162	71.5831	1	12.521	4.748
12	6.35059	38.618	81.4432	6	12.000	7.920
13	5.98894	39.1252	63.7073	3	21.559	19.587
14	5.54376	38.0319	53.3036	10	11.086	7.030
15	4.873	39.0683	57.8956	4	8.054	10.010
16	4.61831	36.9053	102.476	1	7.777	5.214
17	4.43294	38.7698	61.2802	4	15.480	8.839
18	4.25345	36.326	78.3978	1	12.174	6.446
19	3.82841	35.5458	64.8576	2	5.041	7.519

20	3.42857	38.0731	99.8759	2	7.867	4.961
21	3.34678	36.3131	62.9525	10	6.237	8.013
22	2.84919	37.2215	67.7144	4	14.199	11.248
23	2.75952	34.6775	56.7099	8	22.439	19.095
24	2.39014	34.6317	63.1427	10	12.714	8.074
25	1.69794	35.5682	65.1989	8	8.839	9.121
26	1.65597	34.7061	68.534	3	19.010	21.709
27	0.761431	33.8651	59.5824	9	14.171	9.310
28	0.457764	32.3594	54.8453	10	12.305	7.603
29	0.102792	31.8219	65.1714	9	10.777	5.253
30	-4.37105	26.9452	55.4452	10	9.992	6.198
31	-4.9625	30.776	59.7369	3	3.255	8.833
32	-6.46586	28.6328	71.1944	8	10.521	7.237

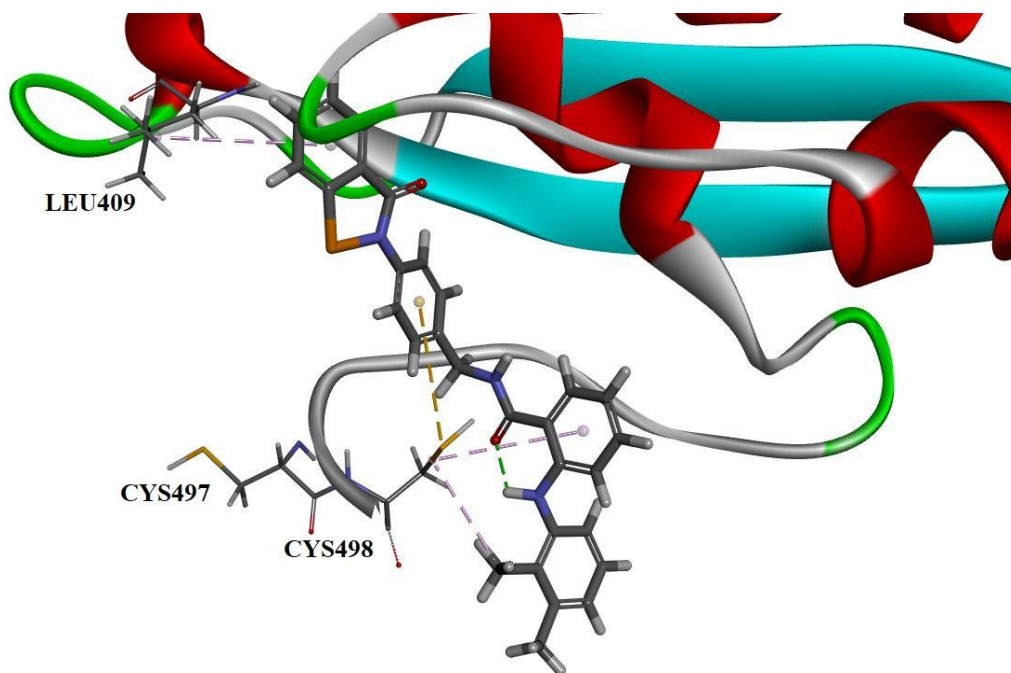
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Table 5 Ligand-protein poses for compound **5j**

Pose Index	CDOCKER_ENERGY	CDOCKER_INTERRATIO N_ENERGY	LibDockS core	LibDock Pose	Distance Cys497Se	Distance Cys498Se
1	19.8943	44.2099	64.1592	6	9.110	6.912
2	19.1219	46.7132	70.4112	6	20.387	32.317
3	18.4552	43.1668	78.5763	5	11.084	11.073
4	17.5838	42.5621	63.3701	8	7.990	6.253
5	17.4763	41.6662	54.3429	8	13.337	7.076
6	17.4124	43.2487	75.6808	4	9.017	8.893
7	16.1704	39.5507	56.622	9	9.765	6.149
8	15.7242	40.3922	83.1296	2	8.880	8.007
9	15.2206	40.1879	76.0685	3	15.860	12.024
10	15.2121	40.9488	86.7815	1	10.155	7.855
11	14.0462	39.2595	77.2953	6	12.905	6.510
12	13.9291	38.1134	73.0708	5	10.329	9.048
13	13.3664	38.6014	76.3795	7	10.564	3.642
14	13.1608	39.1776	73.2676	8	13.721	12.239
15	12.7202	38.1457	98.5742	1	13.135	6.068
16	12.7057	37.448	87.1604	1	11.449	9.654
17	12.5957	37.812	61.7665	7	13.710	6.822
18	12.0462	37.1745	79.0099	4	9.027	7.055
19	11.8174	37.5311	102.361	1	9.922	9.167
20	11.7229	37.8333	79.1876	3	11.555	8.871
21	11.5495	36.7249	59.7242	9	12.927	9.550
22	11.46	36.1866	73.0299	6	20.179	18.749
23	11.0925	34.6597	67.1314	10	10.872	8.150
24	9.93392	33.6135	91.6464	3	9.116	7.502
25	9.76676	38.4745	58.9347	8	9.822	10.982
26	7.13759	35.5936	71.6435	9	11.223	8.436
27	5.71796	32.2846	69.6629	10	8.831	7.709
28	5.05737	30.4126	65.6731	8	12.929	7.701
29	2.94544	30.5268	54.4481	10	10.113	6.789
30	1.81129	26.1195	51.8166	9	20.884	21.979

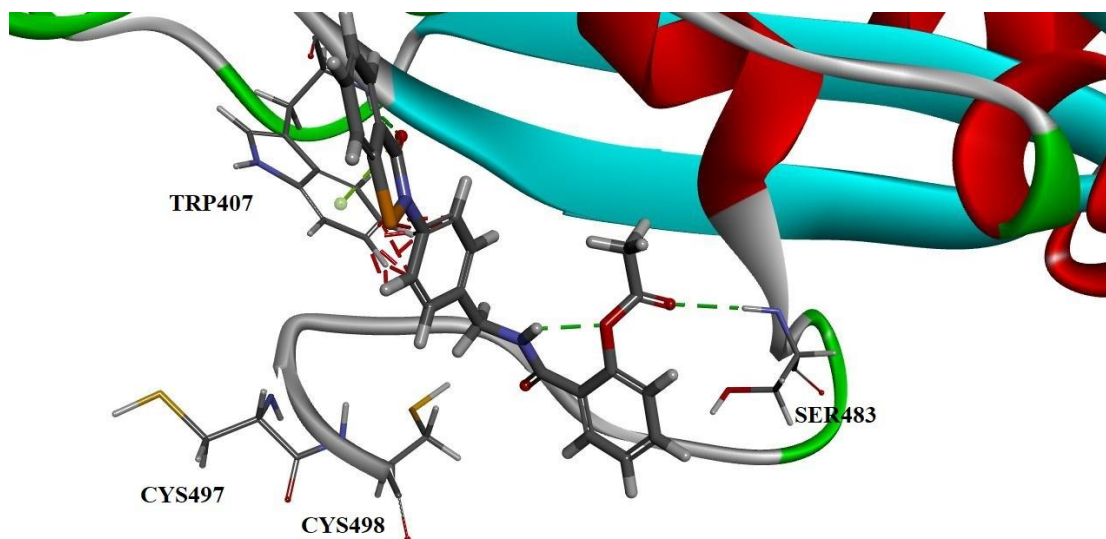
3



1

2 **Fig. 4.** The pose 5 of **5c**. Four interactions were shown: Hydrophobic (Pi-Alkyl)
 3 between two different benzene rings and CYS498 (5.141 Å, 5.137 Å), hydrophobic
 4 (Alkyl) between methyl groups on benzene ring and CYS498 (4.190 Å), and
 5 hydrophobic (Pi-Alkyl) between phenyl of 3-oxobenzo[d][1,2]selenazol-2(3H)-
 6 yl)benzyl and LEU409 (5.469 Å).

7



8

9 **Fig. 5.** The pose 4 of **5j**. Two interactions were shown: hydrogen bonding between
 10 the oxygen of acetyl group and SER483 (2.29 Å) and hydrogen bonding between the
 11 oxygen of 3-oxobenzo[d][1,2]selenazol-2(3H)-yl)benzyl and TRP407 (2.00 Å).

12

3. Conclusions

In summary, two series of NSAIDs-EBS derivatives were synthesized and characterized. Five human cell lines (BGC-823, SW480, MCF-7, HeLa and A549) were selected to test cytotoxicity of the compounds. Compound **5j** showed most potent cytotoxicity activity with IC₅₀ values below 3 μ m against five cancer cell lines. Moreover, most of the NSAIDs-EBS derivatives exhibited moderate to good CPx-like activity compared to ebselen. Finally, TrxR1 inhibition activity assay and in flexible docking study performed into TrxR1 enzyme, compound **5j** showed a moderate binding energies and binding mode that the distance between the selenium atom and Cys497/Cys498.

Overall, considering the potency of these NSAIDs-EBS derivatives on cancer cell viability, antioxidant activity and docking assay, the further study will focus on design of this new type of potential NSAIDs-EBS anticancer agents.

4. Experimental section

4.1. General methods

All chemical reagents for the synthesis of the compounds were purchased from Macklin (Shanghai, China) or TCI (Shanghai, China) and used without further purification unless stated otherwise. Thin-layer chromatography (TLC) was performed on aluminium pre-coated sheets (E. Merck Silica gel 60 F254). Melting points were recorded on an Electrothermal apparatus and are uncorrected. NMR spectra were recorded in CDCl₃ on a Bruker Avance 400 MHz (for ¹H) and 100 MHz (for ¹³C) spectrometer with 5 mm PABBO probe. The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, and m = multiplet. Chemical shifts (δ) are reported in parts per million (ppm) downfield from TMS and the coupling constants (*J*) are expressed in Hertz (Hz). High-resolution MS were performed on a SCIEX, TripleTOF 5600+, operating in ionization mode.

4.2. Experimental procedures

4.2.1. Procedure for the synthesis of compound **1**

1 To a solution of 2-(4-aminophenyl)ethylamine (2.0 g) in THF (20 mL) was
2 added (Boc)₂O (4.3 g) at 0°C. Then the mixture was stirred at 25°C for 3 hrs. TLC
3 showed the reaction was complete. The mixture was concentrated under reduced
4 pressure. The crude product was purified by column chromatography on silica gel
5 (PE/EA = 50:1 to 5:1) to afford compound **2** as white solid (2 g) in 55% yield. ¹H
6 NMR (400 MHz, CDCl₃): 1.45 (s, 9H, 3-CH₃), 3.66 (br, 2H, -NH), 4.18 (d, J = 5.8 Hz,
7 2H, -CH₂), 4.73 (br, 1H, -NH), 6.66-6.61 (m, 2H), 7.10-7.03 (m, 2H). ¹³C NMR (100
8 MHz, CDCl₃): 28.6, 44.5, 79.4, 115.3, 128.9, 129.0, 145.8.

9 10 *4.2.2. procedure for the synthesis of compound 2*

11 To a solution of compound **1** (2.0 g) in DCM (40 mL) was added TEA (1.36 g)
12 at 0°C, then **3** (1.36 g) was added slowly into the mixture. The mixture was stirred at
13 25°C for 0.5 hour. TLC showed the reaction was complete. The mixture was diluted
14 with H₂O (40 mL), the aqueous layer was extracted with DCM (20 mL×2), the
15 combined organic layer was washed with brine (20 mL× 2), dried over Na₂SO₄,
16 filtered and the filtrate was concentrated under reduced pressure. The crude product
17 was purified by beating (DCM/MeOH=4:1) to afford compound **2** (2.8 g) as a white
18 solid in 69% yield. ¹H NMR (400 MHz, CDCl₃): 1.45 (s, 9H, 3-CH₃), 4.28 (s, 2H, -
19 CH₂), 4.90 (brs, 1H, -NH), 7.14 (t, 1H, J = 8.00 Hz, ArH), 7.28 (s, 1H, ArH), 7.41 (t,
20 1H, J = 8.00 Hz, ArH), 7.50 (d, 1H, J = 8.00 Hz, ArH), 7.59 (d, 2H, J = 8.00 Hz, ArH),
21 7.68 (brs, 1H, ArH), 7.89 (d, 1H, J = 8.00 Hz, ArH). ¹³C NMR (100 MHz, CDCl₃):
22 28.4, 44.7, 79.8, 92.7, 115.3, 127.7, 128.9, 129.0, 130.9, 131.3, 141.7, 142.5, 145.8,
23 164.9.

24 *4.2.3. procedure for the synthesis of compound 3*

25 To a solution of compound **2** (1.9 g) in DMF (18 mL) was added CuI (799 mg),
26 Cs₂CO₃ (3.43 g), KSeCN (726 mg) and 1,10-phenanthroline (757 mg). Then the
27 mixture was stirred at 100 °C for 0.5 hour. TLC showed the reaction was complete.
28 The mixture was cooled to 25°C and then diluted with H₂O (40 mL) and ethyl acetate
29 (EA) (20 mL), the aqueous layer was extracted with EA (20 mL×2), the combined
30 organic layer was washed with brine (20 mL×2), dried over Na₂SO₄, filtered and the

1 filtrate was concentrated under reduced pressure. The crude product was purified by
2 beating (MeOH, 5mL) to afford compound 3 (0.83 g) as a yellow solid in 49% yield.
3 ¹H NMR (400 MHz, DMSO): 1.41 (s, 9H, 3-CH₃), 4.14 (s, 2H, -CH₂), 7.31 (d, 1H, J
4 = 8.00 Hz, ArH), 7.45-7.50 (m, 2H, ArH), 7.57 (d, 1H, J = 8.00 Hz, ArH), 7.68 (t, 1H,
5 J = 8.00 Hz, ArH), 7.91 (d, 1H, J = 8.00 Hz, ArH), 8.09 (d, 1H, J = 8.00 Hz, ArH).
6 ¹³C NMR (100 MHz, DMSO): 28.7, 44.9, 79.6, 121.4, 127.1, 127.5, 128.8, 131.1,
7 131.4, 133.8, 136.2, 137.4, 143.2, 167.6.

8

9 4.2.4. procedure for the synthesis of compound 4

10 To a solution of compound 3 (430 mg) in DCM (5 mL) was added TFA (1 mL).
11 The mixture was stirred at 25°C for 1 hour. TLC showed the reaction was complete.
12 The mixture was concentrated under reduced pressure to afford crude compound 4.
13 The pH of crude was adjusted to about value 9 by TEA. The mixture was used for the
14 next step without purification.

15

16 4.2.5. General procedure for the synthesis of compounds 5a-5j

17 To a solution of patent NSAIDs (1.0 eq) in DCM (5 mL) was added EDCI (1.2
18 eq.), HOBT (1.2 eq.) and TEA (3.0 eq.) and compound 4 (1.0 eq). The mixture was
19 stirred at 25°C for 16 hrs. TLC showed the reaction was complete. The mixture was
20 diluted with H₂O, the aqueous layer was extracted with DCM, the combined organic
21 layer was washed with brine, dried over Na₂SO₄, filtered and the filtrate was
22 concentrated under reduced pressure. The crude product was purified by beating to
23 afford the desired product.

24

25 4.2.5.1. 2-(4-isobutylphenyl)-N-(4-(3-oxobenzo[d][1,2]selenazol-2(3H)-yl)benzyl)

26 Propenamide (5a).

27 Yield: 85 %. White solid. Mp: 103-105°C. ¹H NMR (400 MHz, DMSO-6d): δ 0.85 (d,
28 6H, J = 4.00 Hz, 2-CH₃), 1.36 (d, 3H, J = 8.00 Hz, -CH₃), 1.77-1.84 (m, 1H, -CH),
29 2.41 (d, 2H, J = 8.00 Hz, -CH₂), 3.65 (q, 1H, J = 8.00 Hz, -CH), 4.26 (d, 2H, J = 8.00
30 Hz, -CH₂), 7.09 (d, 2H, J = 8.00 Hz, ArH), 7.21 (d, 2H, J = 8.00 Hz, ArH), 7.25 (d,

1 2H, J = 8.00 Hz, ArH), 7.47 (d, 1H, J = 8.00 Hz, ArH), 7.52 (d, 2H, J = 8.00 Hz,
2 ArH), 7.66-7.70 (m, 1H, ArH), 7.89-7.91 (m, 1H, ArH), 8.10 (d, 1H, J = 8.00 Hz,
3 ArH), 8.47-8.50 (m, 1H, -NH). ¹³C NMR (100 MHz, DMSO-6d): δ 18.9, 22.6, 30.1,
4 42.1, 44.7, 45.3, 125.0, 126.3, 126.7, 127.5, 128.2, 128.4, 129.0, 129.3, 132.7, 137.7,
5 138.7, 139.4, 139.8, 140.0, 165.4, 174.0. HRMS calcd. For C₂₇H₂₈N₂O₂Se [M+H]⁺:
6 493.1316, found 493.1389 [M+H]⁺.

7

8 4.2.5.2.2-(2-fluoro-[1,1'-biphenyl]-4-yl)-N-(4-(3-oxobenzod[1,2]selenazol-2(3H)-
9 yl)benzyl)propenamide (**5b**).

10 Yield: 82 %. White solid. Mp: 97-99°C. ¹H NMR (400 MHz, DMSO-6d): δ 1.43 (d,
11 3H, J = 8.00Hz, -CH₃), 3.78 (q, 1H, J = 4.00 Hz, -CH), 4.24-4.36 (m, 2H, -CH₂),
12 7.27-7.30 (m, 4H, ArH), 7.38-7.41 (m, 1H, ArH), 7.46-7.51 (m, 4H, ArH), 7.54-7.58
13 (m, 4H, ArH), 7.67-7.71 (m, 1H, ArH), 7.91 (d, 1H, J = 8.00 Hz, ArH), 8.09 (d, 1H, J
14 = 8.00 Hz, ArH), 8.63 (brs, 1H, -NH). ¹³C NMR (100 MHz, DMSO-6d): δ 18.9, 42.3,
15 45.1, 115.3 (d, J = 23 Hz), 124.3 (d, J = 3 Hz), 125.1, 126.3, 126.7, 126.9 (d, J =
16 13Hz), 128.2, 128.4, 128.9, 129.1, 129.2, 130.5 (d, J = 4 Hz), 132.7, 135.,5, 137.6,
17 138.8, 139.4, 144.5 (d, J = 8 Hz), 158.1, 160.5, 165.5, 173.3. HRMS calcd. For
18 C₂₉H₂₃FN₂O₂Se [M+H]⁺: 531.0987, found 531.0962 [M+H]⁺.

19

20 4.2.5.3. 2-((2,3-dimethylphenyl)amino)-N-(4-(3-oxobenzod[1,2]selenazol-2(3H)-
21 yl)benzyl)benzamide (**5c**).

22 Yield: 78 %. White solid. Mp: 110-112°C. ¹H NMR (400 MHz, CDCl₃): δ 2.20 (s, 3H,
23 -CH₃), 2.32 ((s, 3H, -CH₃), 4.61 (s, 2H, -CH₂), 6.64-6.68 (m, 1H, ArH), 6.75-6.77(m,
24 1H, ArH), 6.92 (d, 1H, J = 8.00 Hz, ArH), 6.95(d, 1H, J = 8.00 Hz, ArH), 7.05-7.09
25 (m, 1H, ArH), 7.17-7.23 (m, 2H, ArH), 7.40 (d, 2H, J = 8.00 Hz, ArH), 7.45-7.49 (m,
26 2H, ArH), 7.58 (d, 2H, J = 8.00 Hz, ArH), 7.63-7.68 (m, 2H, ArH), 8.10 (d, 1H, J =
27 8.00 Hz, ArH), 9.26 (s, 1H, -NH). ¹³C NMR (100 MHz, CDCl₃): δ 14.0, 20.7, 43.3,
28 114.9, 116.4, 116.8, 121.1, 123.8, 125.7, 125.8, 126.6, 127.4, 127.5, 128.7, 129.4,
29 131.1, 132.5, 132.6, 137.0, 137.7, 138.1, 138.3, 139.5, 147.4, 165.9, 169.7. HRMS
30 calcd. For C₂₉H₂₅N₃O₂Se[M+H]⁺: 528.119, found 528.1172 [M+H]⁺.

1

2 4.2.5.4. 2-(3-benzoylphenyl)-N-(4-(3-oxobenzod[1,2]selenazol-2(3H)-
3 yl)benzyl)propanamide (**5d**).

4 Yield: 80 %. White solid. Mp: 90-92°C. ¹H NMR (400 MHz, CDCl₃): δ 1.56 (d, 3H, J
5 = 8.00 Hz, -CH₃), 3.68 (q, 1H, J = 8.00 Hz, -CH), 4.35-4.38 (m, 2H, -CH₂), 7.19 (d,
6 2H, J = 8.00 Hz, ArH), 7.42-7.49 (m, 6H, ArH), 7.56-7.61 (m, 2H, ArH), 7.64-7.66
7 (m, 3H, ArH), 7.75-7.77 (m, 3H, ArH), 8.06 (s, 1H, -NH). ¹³C NMR (100 MHz,
8 CDCl₃): δ 18.7, 43.1, 46.9, 123.9, 125.7, 126.6, 127.4, 128.4, 128.5, 128.8, 129.1,
9 129.2, 129.3, 130.1, 131.6, 132.6, 137.0, 137.8, 138.1, 138.2, 141.8, 165.8, 173.6,
10 196.6. HRMS calcd. For C₃₀H₂₄N₂O₃Se[M+H]⁺: 541.103, found 541.1001 [M+H]⁺.

11

12 4.2.5.5. (Z)-2-(5-fluoro-2-methyl-1-(4-(methylsulfinyl)benzylidene)-1H-inden-3-yl)-N-
13 (4-(3-oxobenzod[1,2]selenazol-2(3H)-yl)benzyl)acetamide (**5e**).

14 Yield: 85%. White solid. Mp: 130-132°C. ¹H NMR (400 MHz, CDCl₃): δ 2.21 (s, 3H,
15 -CH₃), 2.80 (s, 3H, -CH₃), 3.59 (s, 2H, -CH₂), 4.43 (s, 2H, -CH₂), 6.10 (s, 1H, -NH),
16 6.56-6.61 (m, 1H, ArH), 6.87-6.89 (m, 1H, ArH), 7.16-7.21 (m, 4H, ArH), 7.44-7.52
17 (m, 3H, ArH), 7.64-7.67 (m, 4H, ArH), 7.70-7.72 (m, 2H, ArH), 8.08 (d, 1H, J = 4.00
18 Hz, ArH). ¹³C NMR (100 MHz, CDCl₃): δ 10.7, 33.8, 43.1, 43.9, 106.1 (d, J = 23 Hz),
19 111.3 (d, J = 23 Hz), 123.8, 124.0, 125.7, 126.6, 127.4, 128.5, 128.9, 129.4, 129.6,
20 130.3, 132.3, 132.7, 136.7, 137.7, 138.3, 138.9, 139.4, 141.4, 145.6, 146.2, 162.2,
21 164.6, 165.8, 169.2. HRMS calcd. For C₃₄H₂₇FN₂O₃SSe [M+H]⁺: 643.097, found
22 643.0956 [M+H]⁺.

23

24 4.2.5.6. 2-(6-methoxynaphthalen-2-yl)-N-(4-(3-oxobenzod[1,2]selenazol-2(3H)-
25 yl)benzyl)propanamide (**5f**).

26 Yield: 85 %. White solid. Mp: 88-90°C. ¹H NMR (400 MHz, DMSO-6d): δ 1.45 (d,
27 3H, J = 4.00 Hz, -CH₃), 3.82 (q, 1H, J = 4.00 Hz, -CH), 3.86 (s, 3H, -CH₃), 4.27 (s,
28 2H, -CH₂), 7.13-7.16 (m, 1H, ArH), 7.25 (d, 2H, J = 8.00 Hz, ArH), 7.28 (d, 1H, J =
29 4.00 Hz, ArH), 7.46-7.49 (m, 2H, ArH), 7.52 (d, 2H, J = 8.00 Hz, ArH), 7.66-7.70 (m,
30 1H, ArH), 7.73 (s, 1H, ArH), 7.76-7.80 (m, 2H, ArH), 7.89 (d, 1H, J = 8.00 Hz, ArH),
31 8.09 (d, 1H, J = 8.00 Hz, ArH), 8.56 (brs, 1H, -NH). ¹³C NMR (100 MHz, DMSO-6d):

1 δ 19.0, 42.2, 45.5, 55.6, 160.2, 119.1, 125.0, 125.8, 126.3, 126.7, 127.0, 127.1, 128.4,
2 128.8, 128.8, 128.9, 129.6, 132.7, 133.6, 137.7, 137.8, 138.7, 139.4, 157.5, 165.4,
3 173.9. HRMS calcd. For $C_{28}H_{24}N_2O_3Se[M+H]^+$: 517.103, found 517.1019 $[M+H]^+$.

4
5

6 4.2.5.7. *N*-(4-(3-oxobenzod[1,2]selenazol-2(3H)-yl)benzyl)-2-((3-
7 (trifluoromethyl)phenyl)amino)benzamide (**5g**)

8 Yield: 77%. White solid. Mp: 121-123°C. 1H NMR (400 MHz, DMSO- d_6): δ 1.93 (s,
9 2H, -CH₂), 6.99 (t, 1H, J = 8.00 Hz, ArH), 7.23 (d, 1H, J = 8.00 Hz, ArH), 7.36-7.43
10 (m, 6H, ArH), 7.47-7.50 (m, 2H, ArH), 7.58 (d, 2H, J = 8.00 Hz, ArH), 7.68 (t, 1H, J
11 = 8.00 Hz, ArH), 7.78 (d, 1H, J = 8.00 Hz, ArH), 7.90 (d, 1H, J = 8.00 Hz, ArH), 8.09
12 (d, 1H, J = 8.00 Hz, ArH), 9.20 (brs, 1H, -NH), 9.71 (s, 1H, -NH). ^{13}C NMR (100
13 MHz, DMSO- d_6): δ 42.6, 114.5 (q, J_{C-F} = 4.0 Hz), 117.4, 117.5 (q, J_{C-F} = 4.0 Hz),
14 120.4, 121.6, 122.1, 124.2 (q, J_{C-F} = 271 Hz, -CF₃), 125.1, 126.3, 126.7, 128.4, 128.5,
15 128.9, 129.5, 130.8 (q, J_{C-F} = 32 Hz), 131.0 132.5 (d, J = 22 Hz), 137.5, 138.8, 139.4,
16 143.1, 143.6, 165.4, 168.9. HRMS calcd. For $C_{28}H_{20}F_3N_3O_2Se[M+H]^+$: 568.0751,
17 found 568.0739 $[M+H]^+$.

18
19

4.2.5.8.

20 2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1H-indol-3-yl)-N-(4-(3-
21 oxobenzod[1,2]selenazol-2(3H)-yl)benzyl)acetamide (**5h**)

22 Yield: 82%. White solid. Mp: 91-93°C. 1H NMR (400 MHz, DMSO- d_6): δ 2.26 (s,
23 3H, -CH₃), 3.61 (s, 2H, -CH₂), 3.75 (s, 3H, -OCH₃), 4.30 (s, 2H, -CH₂), 7.72 (d, 1H, J
24 = 4.00 Hz, ArH), 6.97 (d, 1H, J = 8.00 Hz, ArH), 7.15 (s, 1H, ArH), 7.32 (d, 2H, J =
25 8.00 Hz, ArH), 7.49 (t, 1H, J = 8.00 Hz, ArH), 7.55 (d, 2H, J = 8.00 Hz, ArH), 7.63-
26 7.66 (m, 2H, ArH), 7.69-7.71 (m, 3H, ArH), 7.90 (d, 1H, J = 8.00 Hz, ArH), 8.09 (d,
27 1H, J = 8.00 Hz, ArH), 8.61 (s, brs, 1H, -NH). ^{13}C NMR (100 MHz, DMSO- d_6): δ
28 13.9, 31.7, 42.4, 55.9, 102.3, 111.9, 114.8, 115.1, 125.0, 126.3, 126.7, 128.4, 128.5,
29 128.9, 129.5, 130.8, 131.3, 131.6, 132.7, 134.7, 135.7, 137.7, 138.0, 138.8, 139.3,
30 156.1, 165.5, 168.4, 170.0. HRMS calcd. For $C_{33}H_{26}ClN_3O_3Se[M+H]^+$: 644.0855,
31 found 644.0841 $[M+H]^+$.

1

2 4.2.5.9.2-(1,8-diethyl-1,3,4,9-tetrahydropyrano[3,4-b]indol-1-yl)-N-(4-(3-

3 oxobenzo[d][1,2]selenazol-2(3H)-yl)benzyl)acetamide (**5i**).

4 Yield: 85%. White solid. Mp: 96-98°C. ¹H NMR (400 MHz, DMSO-6d): δ 0.68 (t,
5 3H, J = 8.00 Hz, -CH₃), 1.25 (t, 3H, J = 8.00 Hz, -CH₃), 2.06 (q, 2H, J = 8.00 Hz, -
6 CH₂), 2.60-2.71 (m, 2H, -CH₂), 2.74-2.78 (m, 1H -CH-), 2.83 (q, 2H, J = 8.00 Hz, -
7 CH₂), 2.91-2.95 (m, 1H -CH-), 3.98 (s, 2H, -CH₂), 4.26-4.35 (m, 2H, -CH₂), 6.88-6.95
8 (m, 2H, ArH), 7.19 (d, 2H, J = 8.00 Hz, ArH), 7.24 (d, 1H, J = 4.00 Hz, ArH), 7.45 (d,
9 2H, J = 8.00 Hz, ArH), 7.50 (d, 1H, J = 8.00 Hz, ArH), 7.69 (t, 1H, J = 8.00 Hz, ArH),
10 7.90 (d, 1H, J = 8.00 Hz, ArH), 8.10 (d, 1H, J = 8.00 Hz, ArH), 8.17 (brs, 1H, -NH),
11 10.54 (s, 1H, -NH). ¹³C NMR (100 MHz, DMSO-6d): δ 8.3, 14.9, 22.4, 24.2, 31.1,
12 42.1, 44.4, 60.4, 76.0. HRMS calcd. For C₃₁H₃₁N₃O₃Se[M+H]⁺: 574.1609, found
13 574.1571 [M+H]⁺.

14

15 4.2.5.10. 2-((4-(3-oxobenzo[d][1,2]selenazol-2(3H)-yl)benzyl)carbamoyl)phenyl
16 acetate (**5j**).

17 Yield: 82%. White solid. Mp: 112-114°C. ¹H NMR (400 MHz, CDCl₃): δ 2.14 (s, 3H,
18 -CH₃), 4.59 (d, 2H, J = 4.00 Hz, -CH₂), 6.75 (s, 1H, -NH), 7.09 (d, 1H, J = 8.00 Hz,
19 ArH), 7.29 (t, 1H, J = 8.00 Hz, ArH), 7.39 (d, 2H, J = 8.00 Hz, ArH), 7.46 (t, 2H, J =
20 8.00 Hz, ArH), 7.60 (d, 2H, J = 8.00 Hz, ArH), 7.64-7.69 (m, 2H, ArH), 7.77 (d, 1H, J
21 = 8.00 Hz, ArH), 8.08 (d, 1H, J = 8.00 Hz, ArH). ¹³C NMR (100 MHz, CDCl₃): δ
22 20.9, 43.4, 123.3, 123.8, 125.7, 126.4, 126.6, 127.4, 128.2, 128.9, 129.4, 129.8, 132.0,
23 132.7, 136.7, 137.6, 138.6, 148.0, 165.6, 165.8, 169.3. HRMS calcd. For
24 C₂₃H₁₈N₂O₄Se[M+H]⁺: 466.0432, found 467.0534 [M+H]⁺.

25

26 4.2.6. procedure for the synthesis of compound **6**

27 To a solution of N-Boc-Ethylenediamine (9.0 g) and TEA (6.83 g) in DCM (200
28 mL) was added o-iodobenzoyl chloride (15.0 g) in portions at 0°C. Then the mixture
29 was stirred at 0°C for 0.5 hour. TLC showed the reaction was complete. Then H₂O
30 (200 mL) was added into the mixture. The aqueous layer was extracted with DCM (20
31 mL×2), the combined organic layer was washed with brine (50 mL×1), dried over

1 Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure. The crude
2 product was slurried by MeOH to afford the compound **6** (18.5 g) in 84% yield. ¹H
3 NMR (400 MHz, CDCl₃): 1.40 (s, 3H, 3-CH₃), 3.36-3.40 (m, 2H, -CH₂), 3.51-3.55 (m,
4 2H, -CH₂), 5.09 (brs, 1H, -NH), 6.58 (brs, 1H, -NH), 7.05-7.07 (m, 1H, ArH), 7.33-
5 7.35 (m, 2H, ArH), 7.81-7.83 (m, 2H, ArH). ¹³C NMR (100 MHz, CDCl₃): 27.4, 36.5,
6 52.2, 89.6, 92.8, 127.5, 130.7, 131.2, 141.8, 142.7, 167.8.

7

8 *4.2.7. procedure for the synthesis of compound 7*

9 To a solution of compound **6** (18.5 g) in DMF (180 mL) was added CuI (9.0 g),
10 Cs₂CO₃ (38.66 g), KSeCN (8.19 g) and 1,10-phenanthroline (8.54 g). Then the
11 mixture was stirred at 100 °C for 40 minutes. TLC showed the reaction was complete.
12 The mixture was cooled to 25°C and then diluted with H₂O (400 mL) and EA (200
13 mL), the aqueous layer was extracted with EA (200 mL×1), the combined organic
14 layer was washed with brine (200 mL×1), dried over Na₂SO₄, filtered and the filtrate
15 was concentrated under reduced pressure. The crude product was slurried by EA to
16 afford compound **7** (7.0 g) as yellow solid in 43.5 % yield. ¹H NMR (400 MHz,
17 CDCl₃): 1.42 (s, 3H, 3-CH₃), 3.44-3.48 (m, 2H, -CH₂), 3.95-3.98 (m, 2H, -CH₂), 5.03
18 (brs, 1H, -NH), 7.41-7.45 (m, 1H, ArH), 7.58-7.65 (m, 2H, ArH), 8.03-8.06 (m, 1H,
19 ArH). ¹³C NMR (100 MHz, CDCl₃): 27.6, 39.5, 54.2, 89.4, 127.3, 128.9, 131.4, 132.3,
20 133.6, 143.4, 170.5.

21

22 *4.2.8. procedure for the synthesis of compound 8*

23 To a solution of compound **7** (300 mg) in DCM (5 mL) was added TFA (1 mL).
24 The mixture was stirred at 25°C for 1 hour. TLC showed the reaction was complete.
25 The mixture was concentrated under reduced pressure to afford crude product **6**. The
26 pH of crude product was adjusted to about 9 by TEA. The mixture was used for the
27 next step without purification.

28

29 *4.2.9. General procedure for the synthesis of compounds 9a-9j*

1 To a solution of compound **8** (1.0 eq) in DCM (20 mL) was added EDCI (1.2
2 eq.), HOBt (1.2 eq), TEA (3.0 eq) and NSAIDs (1.2 eq). The mixture was stirred at
3 25°C for 16 hrs. TLC showed the reaction was complete. The mixture was diluted
4 with H₂O (20 mL), the aqueous layer was extracted with DCM (15 mL×2), the
5 combined organic layer was washed with brine (15 mL×2), dried over Na₂SO₄,
6 filtered and the filtrate was concentrated under reduced pressure. The crude product
7 was purified by column chromatography on silica gel to afford the desired product.

8
9 4.2.9.1. *2-(4-isobutylphenyl)-N-(2-(3-oxobenzod[1,2]selenazol-2(3H)-*
10 *yl)ethyl)propanamide (9a)*

11 Yield: 80 %. White solid. Mp: 113-115°C. ¹H NMR (400 MHz, CDCl₃): δ 0.86 (d, 6H,
12 J = 4.00 Hz, 2-CH₃), 1.48 (d, 3H, J = 8.00 Hz, -CH₃), 1.78 (q, 1H, J = 4.00 Hz, -CH),
13 2.39 (d, 2H, J = 8.00 Hz, -CH₂), 3.50-3.52 (m, 3H, -CH, -CH₂), 3.85-3.97 (m, 1H, -
14 CH), 6.20-6.22 (m, 1H, ArH), 7.01 (d, 2H, J = 8.00 Hz, ArH), 7.16 (d, 2H, J = 8.00
15 Hz, ArH), 7.42-7.44 (m, 1H, ArH), 7.60-7.61 (m, 1H, ArH), 8.00 (brs, 1H, -NH). ¹³C
16 NMR (100 MHz, CDCl₃): δ 18.3, 22.4, 30.2, 40.2, 44.0, 45.0, 46.7, 124.0, 126.3,
17 126.7, 127.3, 128.8, 129.5, 132.2, 138.3, 140.6, 168.0, 175.1. HRMS calcd. For
18 C₂₂H₂₆N₃O₂Se [M+H]⁺: 431.1237, found 431.1209 [M+H]⁺.

19
20 4.2.9.2. *2-(2-fluoro-[1,1'-biphenyl]-4-yl)-N-(2-(3-oxobenzod[1,2]selenazol-2(3H)-*
21 *yl)ethyl)propanamide (9b)*

22 Yield: 82%. Yellow solid. Mp: 102-104°C. ¹H NMR (400 MHz, CDCl₃): δ 1.51 (d,
23 3H, d = 8.00 Hz, -CH₃), 3.54-3.61 (m, 3H, -CH, -CH₂), 3.86-4.03 (m, 2H, -CH₂), 6.62
24 (brs, 1H, -NH), 7.08-7.12 (m, 2H, ArH), 7.25-7.28 (m, 1H, ArH), 7.34-7.38 (m, 2H,
25 ArH), 7.40-7.48 (m, 4H, ArH), 7.53-7.60 (m, 2H, ArH), 7.97 (d, 1H, J = 8.00 Hz,
26 ArH). ¹³C NMR (100 MHz, CDCl₃): δ 18.3, 40.5, 44.1, 46.5, 115.3 (d, J = 23 Hz),
27 123.6, 124.1, 126.4, 126.6, 127.7, 128.4, 128.7, 128.9, 130.9, 132.3, 135.5, 138.2,
28 142.6 158.5, 161.0, 168.2, 174.2. HRMS calcd. For C₂₄H₂₁FN₂O₂Se [M+H]⁺: 469.083,
29 found 469.0800 [M+H]⁺.

30

1 4.2.9.3. 2-((2,3-dimethylphenyl)amino)-N-(2-(3-oxobenzod[1,2]selenazol-2(3H)-
2 yl)ethyl)benzamide (**9c**). Yield: 80%. White solid. Mp: 131-133°C. ¹H NMR (400
3 MHz, CDCl₃): δ 2.17 (s, 3H, -CH₃), 2.30 (s, 3H, -CH₃), 3.75 (t, 2H, J = 4.00 Hz, -
4 CH₂), 4.11 (t, 2H, J = 4.00 Hz, -CH₂), 6.68 (t, 1H, J = 8.00 Hz, ArH), 6.89 (d, 1H, J =
5 8.00 Hz, ArH), 6.93 (d, 1H, J = 8.00 Hz, ArH), 7.05 (t, 1H, J = 8.00 Hz, ArH), 7.13-
6 7.20 (m, 2H, ArH), 7.40-7.43 (m, 2H, ArH), 7.52 (d, 1H, J = 8.00 Hz, ArH), 7.56-
7 7.62 (m, 2H, ArH), 9.31 (s, 1H, -NH). ¹³C NMR (100 MHz, CDCl₃): δ 13.9, 20.7,
8 41.0, 44.4, 114.9, 116.2, 116.9, 120.9, 124.1, 125.6, 125.7, 126.4, 126.6, 127.8, 128.8,
9 130.9, 132.3, 138.0, 138.3, 139.6, 147.3, 168.4, 170.2. HRMS calcd. For
10 C₂₄H₂₃N₃O₂Se [M+H]⁺: 466.1033, found 466.0999 [M+H]⁺.

11

12 4.2.9.4. 2-(3-benzoylphenyl)-N-(2-(3-oxobenzod[1,2]selenazol-2(3H)-
13 yl)ethyl)propenamide (**9d**).

14 Yield: 82%. White solid. Mp: 116-118°C. ¹H NMR (400 MHz, CDCl₃): δ 1.53 (d, 3H,
15 J = 8.00 Hz, -CH₃), 3.46-3.60 (m 2H, -CH₂), 3.65 (q, 1H, J = 8.00 Hz, -CH), 3.79-
16 3.86 (m, 1H, -CH-), 3.96-4.02 (m, 1H, -CH-), 6.58 (brs, 1H, -NH), 7.32-7.39 (m, 2H,
17 ArH), 7.47 (t, 2H, J = 8.00 Hz, ArH), 7.54-7.60 (m, 4H, ArH), 7.68 (d, 1H, J = 8.00
18 Hz, ArH), 7.77-7.79 (m, 3H, ArH), 7.93 (d, 1H, J = 8.00 Hz, ArH). ¹³C NMR (100
19 MHz, CDCl₃): δ 18.3, 40.6, 44.0, 46.9, 124.2, 126.3, 126.6, 128.4, 128.6, 129.0, 130.2,
20 131.6, 132.2, 132.7, 137.3, 137.9, 138.4, 141.6, 168.0, 174.2, 196.8. HRMS calcd. For
21 C₂₅H₂₂N₂O₃Se [M+H]⁺: 479.0874, found 479.0831 [M+H]⁺.

22

23 4.2.9.5. (Z)-2-(5-fluoro-2-methyl-1-(4-(methylsulfinyl)benzylidene)-1H-inden-3-yl)-N-
24 (2-(3-oxobenzod[1,2]selenazol-2(3H)-yl)ethyl)acetamide (**9e**).

25 Yield: 82%. White solid. Mp: 127-129°C. ¹H NMR (400 MHz, CDCl₃): δ 2.33 (s, 3H,
26 -CH₃), 3.52-3.54 (m, 2H, -CH₂), 3.61 (s, 2H, -CH₂), 3.73 (s, 3H, -CH₃), 3.87-3.90 (m,
27 2H, -CH₂), 6.59 (d, 1H, J = 8.00 Hz, ArH), 6.62 (brs, 1H, -NH), 6.80 (d, 1H, J = 8.00
28 Hz, ArH), 6.83 (d, 1H, J = 4.00 Hz, ArH), 7.34-7.38 (m, 1H, ArH), 7.45-7.47 (m, 2H,
29 ArH), 7.56-7.60 (m, 2H, ArH), 7.75-7.78 (m, 3H, ArH). ¹³C NMR (100 MHz, CDCl₃):
30 δ 100.8, 112.1, 112.5, 115.1, 124.1, 126.3, 126.4, 128.7, 129.2, 130.4, 131.0, 131.4,

1 132.3, 133.8, 136.6, 138.0, 139.4, 156.1, 168.0, 168.5, 170.8. HRMS calcd. For
2 C₂₅H₂₉FN₂O₃SSe [M+H]⁺: 581.0813, found 581.0796 [M+H]⁺.

3

4 4.2.9.6. 2-(6-methoxynaphthalen-2-yl)-N-(2-(3-oxobenzod[1,2]selenazol-2(3H)-
5 yl)ethyl)propanamide (**9f**).

6 Yield: 85%. White solid. Mp: 125-127°C. ¹H NMR (400 MHz, DMSO-d₆): δ 1.42 (d,
7 3H, J = 8.00 Hz, -CH₃), 3.32-3.38 (m, 2H, -CH₂), 3.70-3.83 (m, 3H, -CH, -CH₂), 3.85
8 (s, 3H, -OCH₃), 7.12 (d, 1H, J = 8.00 Hz, ArH), 7.24 (s, 1H, ArH), 7.40-7.44 (m, 2H,
9 ArH), 7.60-7.64 (m, 1H, ArH), 7.69 (d, 2H, J = 8.00 Hz, ArH), 7.74 (d, 1H, J = 8.00
10 Hz, ArH), 7.82 (d, 1H, J = 8.00 Hz, ArH), 8.03 (d, 1H, J = 8.00 Hz, ArH), 8.20 (brs,
11 1H, -NH). ¹³C NMR (100 MHz, DMSO-d₆): δ 18.9, 43.2, 45.7, 55.6, 106.1, 119.0,
12 125.8, 126.2, 126.3, 127.0, 127.8, 128.2, 128.8, 129.6, 132.0, 133.6, 137.6, 140.0,
13 157.4, 167.0, 174.3. HRMS calcd. For C₂₃H₂₂N₂O₃Se[M+H]⁺: 445.0874, found
14 445.0989 [M+H]⁺.

15

16 4.2.9.7. N-(2-(3-oxobenzod[1,2]selenazol-2(3H)-yl)ethyl)-2-((3-
17 (trifluoromethyl)phenyl)amino)benzamide (**9g**).

18 Yield: 80%. White solid. Mp: 99-101°C. ¹H NMR (400 MHz, CDCl₃): δ 3.75 (t, 2H, J
19 = 8.00 Hz, -CH₂), 4.10 (t, 2H, J = 8.00 Hz, -CH₂), 7.18 (d, 1H, J = 4.00 Hz, ArH),
20 7.28-7.41 (m, 4H, ArH), 7.55-7.62 (m, 4H, ArH), 8.00 (d, 1H, J = 8.00 Hz, ArH), 9.61
21 (s, 1H, ArH). ¹³C NMR (100 MHz, CDCl₃): δ 41.1, 44.4, 115.9, 116.1 (q, J_{C-F} = 4.0
22 Hz), 118.2 (q, J_{C-F} = 4.0 Hz), 118.9, 119.3, 122.8, 124.0 (q, J_{C-F} = 271 Hz, -CF₃),
23 124.1, 126.4, 126.5, 128.2, 128.8, 129.8, 131.6 (q, J_{C-F} = 32 Hz), 132.3 (d, J = 12 Hz),
24 138.3, 142.5, 144.3, 168.6, 169.8. HRMS calcd. For C₂₃H₁₈F₃N₃O₂Se [M+H]⁺:
25 506.0594, found 506.0560 [M+H]⁺.

26

27 4.2.9.8. 2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1H-indol-3-yl)-N-(2-(3-
28 oxobenzod[1,2]selenazol-2(3H)-yl)ethyl)acetamide (**9h**).

29 Yield: 80%. White solid. Mp: 116-118°C. ¹H NMR (400 MHz, CDCl₃): 2.33 (s, 3H, -
30 CH₃), 3.52-3.61 (m, 2H, -CH₂), 3.61 (s, 2H, -CH₂), 3.73 (s, 3H, -OCH₃), 3.87-3.90 (m,

1 2H, -CH₂), 6.58-6.61 (m, 1H, ArH), 6.62 (brs, 1H, -NH), 6.80 (d, 1H, J = 8.00 Hz,
2 ArH), 6.83 (d, 1H, J = 4.00 Hz, ArH), 7.34-7.37 (m, 1H, ArH), 7.46 (d, 2H, J = 8.00
3 Hz, ArH), 7.57-7.60 (m, 2H, ArH), 7.75-7.79 (m, 3H, ArH). ¹³C NMR (100 MHz,
4 CDCl₃): δ 13.3, 32.1, 41.0, 43.8, 55.7, 100.8, 112.1, 112.5, 115.1, 124.1, 126.3, 126.4,
5 128.7, 129.2, 130.4, 131.0, 131.4, 132.3, 133.8, 136.6, 138.0, 139.4, 156.1, 168.0,
6 168.5, 170.8. HRMS calcd. For C₂₈H₂₄ClN₃O₄Se [M+H]⁺: 582.0699, found
7 582.0655[M+H]⁺.

8

9 4.2.9.9. *2-(3,6-diethyl-1,3,4,5-tetrahydropyrano[4,3-b]indol-3-yl)-N-(2-(3-*
10 *oxobenzo[d][1,2]selenazol-2(3H)-yl)ethyl)acetamide(9i).*

11 Yield: 85%. White solid. Mp: 87-89°C. ¹H NMR (400 MHz, CDCl₃): δ 0.74 (t, 3H, J
12 = 8.00 Hz, -CH₃), 1.32 (t, 3H, J = 8.00 Hz, -CH₃), 1.85-1.92 (m, 2H, -CH₂), 2.04-2.09
13 (m, 2H, -CH₂), 2.76 (q, 2H, J = 8.00 Hz, -CH₂), 2.81-2.88 (m, 2H, -CH₂), 3.56 (q, 2H,
14 J = 8.00 Hz, -CH₂), 3.90-3.98 (m, 2H, -CH₂), 4.00-4.04 (m, 2H, -CH₂), 6.99 (d, 1H, J
15 = 8.00 Hz, ArH), 7.03-7.06 (m, 1H, ArH), 7.17 (brs, 1H, -NH), 7.32 (d, 1H, J = 8.00
16 Hz, ArH), 7.83-7.42 (m, 1H, ArH), 7.53-7.60 (m, 2H, ArH), 8.00 (d, 1H, J = 8.00 Hz,
17 ArH), 9.52 (s, 1H, -NH). ¹³C NMR (100 MHz, CDCl₃): δ 7.7, 13.9, 22.4, 24.2, 30.9,
18 40.2, 44.0, 44.6, 60.6, 75.5, 107.9, 115.8, 119.5, 120.2, 124.1, 126.3, 126.4, 126.6,
19 126.9, 128.8, 132.3, 134.7, 136.1, 138.2, 168.1, 171.9. HRMS calcd. For
20 C₂₆H₂₉N₃O₃Se [M+H]⁺: 512.1452, found 512.1413 [M+H]⁺.

21

22 4.3. cell viability assay

23 Five human cancer cell lines BGC-823, SW480, MCF-7, HeLa and A549 cells
24 were maintained in RPMI 1640 medium with 10% fetal bovine serum (FBS) and 100
25 units/mL of penicillin and streptomycin (Thermo Fisher Scientific, shanghai, China)
26 at 37 °C and 5% CO₂ in a humidified atmosphere. Cells were passaged at
27 preconfluent densities, using a solution containing 0.05% trypsin and 0.5 mM EDTA.

28 All the tested NSAIDs-EBS derivatives were evaluated in vitro for their
29 cytotoxicity activity against five cancer cell lines by 3-(4,5-dimethylthiazol-2-yl)-2,5-
30 diphenyl-2H-tetrazolium bromide (MTT) assay according to the method as described

1 before [43]. Exponentially growing cells were harvested and plated in 96-well plates
2 at a concentration of 1×10^4 cells / well. After 24 h incubation at 37 °C under a
3 humidified 5% CO₂ to allow cell attachment, the cells in the wells were respectively
4 treated with target compounds at various concentrations for 24 h, 48 h and 72 h. The
5 concentration of DMSO was always kept below 1.25%, which was found to be non-
6 toxic to the cells. Three hours prior to experiment termination, MTT solution (20 µL
7 of 5.0 mg/mL solution) was added to each well and incubated at 37°C. At the
8 termination time point, the medium/MTT mixtures were removed, and the formazan
9 crystals formed by the mitochondrial dehydrogenase activity of vital cells were
10 dissolved in 100 µL of DMSO per well. The optical densities were measured at 570
11 nm using a 96-well multiscanner (Dynex Technologies, MRX Revelation; Chantilly,
12 VA, USA).

13

14 4.4. DPPH free radical scavenging activity

15 DPPH free radical scavenging activity of corresponding compounds was
16 measured according to the method as previous reported with little optimization [44].
17 Briefly, 20 mL of test samples at different concentrations was mixed with 180 mL of
18 or DPPH solution for 30 min in the dark. Then, the change in absorbance at 517 nm
19 for DPPH was measured on a microplate reader. Ascorbic acid (vitamin C) and
20 ebselen were used as a positive control, DMSO was used as a negative control.

21

22 4.5. Bleomycin-dependent DNA damage

23 The reaction mixture contained DNA (0.5 mg/mL), bleomycin sulfate
24 (0.05 mg/mL), MgCl₂ (5 mM), FeCl₃ (50 mM), and tested compound in a conc. of
25 0.1 mg/mL. L-ascorbic acid was used as positive control. The mixture was incubated
26 at 37°C for 1h. The reaction was terminated by addition of 0.05 mL EDTA (0.1 M).
27 The color was developed by adding 0.5 mL TBA (1% w/v) and 0.5 mL HCl (25% v/v),
28 followed by heating at 80°C for 30 minutes. After cooling in ice water, the extent of
29 DNA damage was measured by increase in absorbance at 532 nm [45].

30

1 4.6. Glutathione peroxidase-like activity

2 GPx kit (Biodiagnostic, Egypt) was used for the determination of GPx according
3 to Paglia et al [46]. The reaction mixture contained 1ml assay buffer (50mM
4 phosphate buffer containing 0.1% Triton X-100) and 0.1ml NADPH reagent (24
5 mmol Glutathione, 12 unit Glutathione reductase and 4.8 mmol NADPH) and 0.01ml
6 (41 mM) tested compounds and the reaction was started by the addition of H₂O₂ (0.8
7 mM). The contents were mixed well and the absorbances were recorded at 340 nm
8 over a period of 3 min against deionized water. The change of absorbance per minute
9 (A₃₄₀ nm/min) was estimated using ebselen (41 mM) as positive control. The values
10 represented in Fig 3 are expressed after background correction for the reaction with
11 H₂O₂ and GSH. In case of colored compounds, their activities were estimated after
12 subtracting their own absorbances at the used wave length.

14 4.7. Colorimetric detection of rat TrxR1 activity

15 Activity of TrxR1 (Abcam) was assayed using DTNB as substrate. The reactions
16 on 96 well plate were ran in final volume of 100 ml, 100 mM potassium phosphate,
17 pH 7.0, containing 1 mM EDTA, 0.1 mg/ml BSA, 5 nM of TrxR1 and 0.2mM
18 NADPH. Concentration range of compounds for EC₅₀ determination was 0.25-25 mM.
19 Reaction mixture was incubated for 15min on plate shaker at room temperature, after
20 which DTNB was added to final concentration of 5 mM. Enzyme kinetics was
21 monitored on TECAN Infinite M1000 PRO microplate reader, by measuring increase
22 in absorbance at 412 nm for 20 min.

24 4.8. Molecular Modeling

25 4.8.1 Protein and Ligand Preparation

26 The mammalian TrxR1 protein (PDB ID: 1H6V) was obtained from Protein Data
27 Bank. The other subunits of TrxR1 were deleted and only one monomer F was
28 prepared by Protein Preparation Wizard in Maestro 11.5 (Schrödinger, LLC, New
29 York, NY, 2019.). Specifically, subunits F was assigned in sequence, hydrogen was
30 added, ionization and tautomerism were adjusted, hydrogen bond distribution was

1 optimized, water was removed, and structure was minimized. The LigPrep utility in
2 Maestro 11.5 was used to perform ligand preparation applying OPLS2005 force field.
3 The Epik utility is used to generate tautomers and possibly ionized states, and then
4 minimize the resulting 3D conformation.

5 6 4.8.2 Ligand Docking

7 The docking task was completed on Discovery Studio Client 2018. The binding
8 site of TrxR1 was defined as a docking sphere with dimensions X: 27.757, Y: 6.510,
9 Z: 33.698 and R: 15 Å. 10 protein conformations of TrxR1 protein were generated
10 with a maximum alteration of 8 residues, which were typed in CHARMM field force.
11 Under the conformation method FAST, every ligand was generated several
12 conformations. With all other parameters as default, compound **5c** and **5j** were docked
13 into protein structure in the Flexible Docking Protocol. For each pose, the distance
14 between the compound's selenium atom and the sulfur atom of either Cys497 or
15 Cys498 was calculated by the distance monitor in the Discovery Studio. For each
16 ligand, average -CDocker energy and average selenium-sulfur distance were
17 calculated.

18 19 **Statistical analysis**

20 Data were given as mean \pm SD of three independent experiments, graphs and
21 curve fitting were using origin Version 8.0 (OriginLab Corporation, Northampton,
22 USA). P value less than 0.05 was considered statistically significant.

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1

2 **References**

- 3 [1] H. Ribeiro, I. Rodrigues, L. Napoleão, L. Lira, D. Marques, M. Veríssimo, J. P.
4 Andrade, M. Dourado, Non-steroidal anti-inflammatory drugs (NSAIDs), pain and
5 aging: Adjusting prescription to patient features, *Biomed. Pharmacother.* 150
6 (2022) 112958.
- 7 [2] S. Bindu, S. Mazumder, U. Bandyopadhyay, Non-steroidal anti-inflammatory
8 drugs (NSAIDs) and organ damage: A current perspective. *Biochem. Pharmacol.*
9 180 (2020) 114147.
- 10 [3] S. A. Mirzaei, F. Dinmohammadi, A. Alizadeh, F. Elahian, Inflammatory pathway
11 interactions and cancer multidrug resistance regulation, *Life Sci.* 235 (2019)
12 116825.
- 13 [4] K. Mortezaee, W. Parwaie, E. Motevaseli, H. Mirtavoos-Mahyari, A. E. Musa, D.
14 Shabeeb, F. Esmaily, M. Najafi, B. Farhood, Targets for improving tumor
15 response to radiotherapy, *Int. Immunopharmacol.* 76 (2019) 105847.
- 16 [5] S. Ramos-Inza, A. C. Ruberte, C. Sanmartín, A. K. Sharma, D. Plano, NSAIDs:
17 old acquaintance in the pipeline for cancer treatment and prevention-structural
18 modulation, mechanisms of action, and bright future, *J. Med. Chem.* 64 (2021)
19 16380-16421.
- 20 [6] A.P. Fernandes, V. Gandin, Selenium compounds as therapeutic agents in cancer,
21 *Biochimica. et. Biophysica. Acta.* 1850 (2015) 1642-1660.
- 22 [7] D. Basudhar, G. Bharadwaj, R. Y. Cheng, S. Jain, S. Shi, J. L. Heinecke, R. J.
23 Holland, L. A. Ridnour, V. M. Caceres, R. C. Spadari-Bratfisch, N. Paolocci, C. A.
24 Velazquez-Martinez, D. A. Wink, K. M. Miranda, Synthesis and chemical and
25 biological comparison of nitroxyl- and nitric oxide-releasing diazeniumdiolate-
26 based aspirin derivatives, *J. Med. Chem.* 56 (2013) 7804-7820.
- 27 [8] J. L. Williams, N. Nath, J. Chen, T. R. Hundley, J. Gao, L. Kopelovich, K. Kashfi,
28 B. Rigas, Growth inhibition of human colon cancer cells by nitric oxide (NO)-
29 donating aspirin is associated with cyclooxygenase-2 induction and beta-

- 1 catenin/T-cell factor signaling, nuclear factor-kappaB, and NO synthase 2
2 inhibition: implications for chemoprevention, *Cancer Res.* 63 (2003) 7613-7618.
- 3 [9] Y.A. Ammar, M.A. Salem, E.A. Fayed, M.H. Helal, M.S.A. El-Gaby, H. K.
4 Thabet, Naproxen derivatives: Synthesis, reactions, and biological applications,
5 *Synth. Commun.* 47(15) (2017) 1341-1367.
- 6 [10] D. Plano, D. N. Karella, M. K. Pandey, J. E. Spallholz, S. Amin, A. K. Sharma,
7 Design, synthesis, and biological evaluation of novel selenium (Se-NSAID)
8 molecules as anticancer agents, *J. Med. Chem.* 59 (2016) 1946-1959.
- 9 [11] D. Desai, N. Kaushal, U. H. Gandhi, R. J. Arner, C. D'Souza, G. Chen, H. Vunta,
10 K. El-Bayoumy, S. Amin, K. S. Prabhu, Synthesis and evaluation of the anti-
11 inflammatory properties of selenium-derivatives of celecoxib, *Chem. Biol.*
12 *Interact.* 188 (2010) 446-456.
- 13 [12] D. Desai, I. Sinha, K. Null, W. Wolter, M. A. Suckow, T. King, S. Amin, R.
14 Sinha, Synthesis and antitumor properties of selenocoxib-1 against rat prostate
15 adenocarcinoma cells, *Int. J. Cancer.* 127 (2010) 230-238.
- 16 [13] A. C. Ruberte, C. Sanmartin, C. Aydillo, A. K. Sharma, D. Plano, Development
17 and therapeutic potential of selenazo compounds, *J. Med. Chem.* 63 (4) (2020)
18 1473-1489.
- 19 [14] P. Collery, Strategies for the development of selenium-based anticancer drugs, *J.*
20 *Trace Elem. Med. Biol.* 50 (2018) 498-507.
- 21 [15] C. Sanmartin, D. Plano, A. K. Sharma, J. A. Palop, Selenium compounds,
22 apoptosis and other types of cell death: an overview for cancer therapy, *Int. J.*
23 *Mol. Sci.* 13 (8) (2012) 9649-9672.
- 24 [16] F. Martini, S. G. Rosa, I. P. Klann, B. C. W. Fulco, F. B. Carvalho, F. L.
25 Rahmeier, M. C. Fernandes, C. W. Nogueira, A multifunctional compound
26 ebselen reverses memory impairment, apoptosis and oxidative stress in a mouse
27 model of sporadic Alzheimer's disease, *J. Psychiatr. Res.* 109 (2019) 107-117.
- 28 [17] D. Bartolini, P. Torquato, M. Piroddi, F. Galli, Targeting glutathione S-
29 transferase P and its interactome with selenium compounds in cancer therapy,
30 *Biochim. Biophys. Acta, Gen. Subj.* 1863 (2019) 130-143.

- 1 [18] S. Kumar, J. J. Yan, J. F. Poon, V. P. Singh, X. Lu, M. K. Ott, L. Engman, S.
2 Kumar, Multifunctional antioxidants: Regenerable radical-trapping and
3 hydroperoxide-decomposing ebselenols, *Angew. Chem., Int. Ed.* 55 (2016)
4 3729-3733.
- 5 [19] A. J. Pacula, K. B. Kaczor, J. Antosiewicz, A. Janecka, A. Dlugosz, T. Janecki,
6 A. Wojtczak, J. Scianowski, New chiral ebselen analogues with antioxidant and
7 cytotoxic potential, *Molecules*. 22 (2017) 492.
- 8 [20] V. P. Singh, J. F. Poon, J. J. Yan, X. Lu, M. K. Ott, R. J. Butcher, P. J. Gates, L.
9 Engman, Nitro-, azo-, and amino derivatives of ebselen: synthesis, structure, and
10 cytoprotective effects, *J. Org. Chem.* 82 (2017) 313-321.
- 11 [21] D. A. Stoyanovsky, J. F. J. iang, M. P. Murphy, M. Epperly, X. L. Zhang, S. Li, J.
12 Greenberger, V. Kagan, H. Bayir, Design and synthesis of a mitochondria-
13 targeted mimic of glutathione peroxidase, mitoebsele-2, as a radiation mitigator.
14 *ACS Med. Chem. Lett.* 5 (2014) 1304-1307.
- 15 [22] L. Liu, S. Li, X. Li, M. Zhong, Y. Lu, J. Yang, Y. Zhang, X. He, Synthesis of
16 NSAIDs-Se derivatives as potent anticancer agents, *Med. Chem. Res.* 27
17 (2018) 2071-2078.
- 18 [23] Y. Nie, M. Zhong, S. Li, X. Li, Y. Zhang, Y. Zhang, X. He, Synthesis and
19 potential anticancer activity of some novel selenocyanates and diselenides, *Chem.*
20 *Biodivers.* 17(5) (2020) e1900603.
- 21 [24] X. He, M. Zhong, S. Li, X. Li, Y. Li, Z. Li, Y. Gao, F. Ding, D. Wen, Y. Lei, Y.
22 Zhang, Synthesis and biological evaluation of organoselenium (NSAIDs-SeCN
23 and SeCF₃) derivatives as potential anticancer agents, *Eur. J. Med. Chem.* 208
24 (2020) 112864.
- 25 [25] X. He, Y. Nie, M. Zhong, S. Li, X. Li, Y. Guo, Z. Liu, Y. Gao, F. Ding, D. Wen,
26 Y. Zhang. New organoselenides (NSAIDs-Se derivatives) as potential anticancer
27 agents: Synthesis, biological evaluation and in silico calculations, *Eur. J. Med.*
28 *Chem.* 218 (2021) 113384.
- 29

- 1 [26] S. Bedouhène, F. Moulti-Mati, M. Hurtado-Nedelec, P. M. Dang, J. El-Benna,
2 Luminol-amplified chemiluminescence detects mainly superoxide anion
3 produced by human neutrophils, *Am. J. Blood. Res.* 7 (2017) 41-48.
- 4 [27] B.M. Sahoo, B.K. Banik, P. Borah, A. Jain, Reactive oxygen species (ROS): key
5 components in cancer therapies. *Anticancer Agents Med. Chem.* 22(2) (2022)
6 215-222.
- 7 [28] J. E. Klaunig, Oxidative stress and cancer, *Curr. Pharm. Des.* 24 (40) (2018)
8 4771-4778.
- 9 [29] I. Rohn, N. Kroepfl , M. Aschner , J. Bornhorst , D. Kuehnelt , T. Schwerdtle,
10 Selenoneine ameliorates peroxide-induced oxidative stress in *C. elegans*, *J. Trace.*
11 *Elem. Med. Bio.* 55 (2019) 78-81.
- 12 [30] G. Bjørklund, M. Shanaida, R. Lysiuk, H. Antonyak, I. Klishch, V. Shanaida, M.
13 Peana, Selenium: an antioxidant with a critical role in anti-aging, *Molecules.*
14 27(19) (2022) 6613.
- 15 [31] C. P. Kaushik, R. Luxmi, Synthesis, antibacterial, and antioxidant activities of
16 naphthyl-linked disubstituted 1,2,3-triazoles, *J. Heterocycl. Chem.* 57 (2020)
17 2400-2409.
- 18 [32] B. Bocchini, B. Goldani, F. S. S. Sousa, P. T. Birmann, C.A. Brüning, E. J.
19 Lenardão, Santi C, Savegnago L, Alves D, Synthesis and antioxidant activity of
20 new selenium-containing quinolines. *Med. Chem.* 17(6) (2021) 667-676.
- 21 [33] A. Sentkowska, K. Pyrzyńska, Investigation of antioxidant activity of selenium
22 compounds and their mixtures with tea polyphenols. *Mol. Biol. Rep.* 46(3) (2019)
23 3019-3024.
- 24 [34] S.S. Karshieva, G. Babayeva, V. S. Pokrovsky, Y. M. Shlyapnikov, E. A.
25 Shlyapnikova, A. E. Bugrova, A.S. Kononikhin, E. N. Nikolaev, I.L. Kanev,
26 Antitumor effect of bleomycin nanoaerosol in murine carcinoma model.
27 *Molecules.* 28 (10) (2023) 4157.
- 28 [35] U. Galm, M. H. Hager, S.G. Van Lanen, J. Ju, J. S. Thorson, B. Shen. Antitumor
29 antibiotics: bleomycin, enediynes, and mitomycin, *Chem. Rev.* 105 (2005) 739-
30 758.

- 1 [36] Shahabi R, Anissian A, Javadmoosavi SA, Nasirinezhad F, Protective and anti-
2 inflammatory effect of selenium nano-particles against bleomycin-induced
3 pulmonary injury in male rats. *Drug. Chem. Toxicol.* 44 (1) (2021) 92-100.
- 4 [37] L. Flohé, S. Toppo, L. Orian. The glutathione peroxidase family: Discoveries and
5 mechanism. *Free Radic. Biol. Med.* 187 (2022) 113-122.
- 6 [38] R. Gencheva, E. S. J. Arnér, Thioredoxin Reductase Inhibition for Cancer
7 Therapy, *Rev. pharmacol. Toxicol.* 62 (2022) 177-196.
- 8 [39] S. Gromer, L.A. Wessjohann, J. Eubel, W. Brandt, Mutational studies confirm.
9 the catalytic triad in the human selenoenzyme thioredoxin reductase predicted by
10 molecular modeling, *Chembiochem.* 7 (2006) 1649-1652.
- 11 [40] W. Brandt, L. A. Wessjohann, The functional role of selenocysteine (Sec) in the.
12 catalysis mechanism of large thioredoxin reductases: proposition of a swapping
13 catalytic triad including a Sec-His-Glu state, *Chembiochem.* 6 (2005) 386-394.
- 14 [41].E.S.J. Arnér, Targeting the selenoprotein thioredoxin reductase 1 for anticancer
15 therapy. *Adv Cancer Res.* 136 (2017) 139-151.
- 16 [42] S. Shaaban, A. Negm, A. M. Ashmawy, D. M. Ahmed, L. A. Wessjohann.
17 Combinatorial synthesis, in silico, molecular and biochemical studies of
18 tetrazole-derived organic selenides with increased selectivity against
19 hepatocellular carcinoma, *Eur. J. Med. Chem.* 122 (2016) 55-71.
- 20 [43] V. Gandin, P. Khalkar, J. Braude, A.P. Fernandes, Organic selenium compounds
21 as potential chemotherapeutic agents for improved cancer treatment. *Free. Radic.*
22 *Biol. Med.* 127 (2018) 80-97.
- 23 [44] İ. Gulcin, Antioxidants and antioxidant methods: an updated overview, *Arch.*
24 *Toxicol.* 94 (3) (2020) 651-715.
- 25 [45] A. B. A. El-Gazzar, M. M. Youssef, A. M. S. Youssef, A. A. Abu-Hashem, F. A.
26 Badria, Design and synthesis of azolopyrimidoquinolines, pyrimidoquinazolines
27 as anti-oxidant, anti-inflammatory and analgesic activities, *Eur. J. Med. Chem.*
28 44 (2009) 609-624.

1 [46] N. M. Giles, G. I. Giles, J. E. Holley, N. J. Gutowski, C. Jacob, Targeting
2 oxidative stress-related diseases: organochalcogen catalysts as redox sensitizers,
3 Biochem. Pharmacol. 66 (2014) 2021-2028.
4