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**USE OF GENOTYPIC HIV DNA TESTING: A
DELPHI-TYPE CONSENSUS** Short running title:
Delphi Consensus on Genotypic HIV DNA Testing

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► **To cite this version:**

Elisabeth Andre-Garnier, Laurence Bocket, Thomas Bourlet, Laurent Hocqueloux, Quentin Lepiller, et al.. USE OF GENOTYPIC HIV DNA TESTING: A DELPHI-TYPE CONSENSUS Short running title: Delphi Consensus on Genotypic HIV DNA Testing. *Journal of Antimicrobial Chemotherapy*, 2024, 10.1093/jac/dkae007 . hal-04423751

HAL Id: hal-04423751

<https://hal.sorbonne-universite.fr/hal-04423751>

Submitted on 29 Jan 2024

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30 **Abstract**

31 Objectives:

32 As many disparities in the clinical use of HIV DNA sequencing are observed, a DELPHI-type
33 consensus was initiated in France to homogenize use, techniques, and interpretation of results.

34 Methods:

35 Based on a literature review and clinical experience, a Steering Committee (SC) of eight
36 virologists and one infectious disease specialist formulated statements. Statements were
37 submitted to an independent and anonymous electronic vote of virologists and HIV clinicians
38 in France, between October and December 2022.

39 Results:

40 The SC developed 20 statements grouped into six categories: *clinical situations for the use of*
41 *HIV DNA genotyping; techniques for performing HIV DNA genotyping; consideration of*
42 *APOBEC mutations; genotyping results reporting; recycling of antiretrovirals; availability of*
43 *HIV DNA genotyping tests and delays*. Twenty-one virologists and 47 clinicians participated
44 in two voting rounds and 18/20 (90%) assertions reached a ‘strong’ consensus. For example,
45 that prior genotyping on HIV DNA is useful for clinical decision-making when considering
46 switching to some long-acting regimens or to reduce the number of antiretroviral agents in
47 virologically suppressed patients for whom RNA data are unavailable / not exploitable / not
48 sufficiently informative. Two statements achieved no consensus: reporting any detected viral
49 minority population for discussion in multidisciplinary meetings (virologists), and possible
50 risk of virologic failure when using a second generation InSTI + XTC regimen in patients
51 with undetectable viral load ≥ 1 year and in the presence of a documented *M184V* mutation < 5
52 years (clinicians).

53 Conclusion:

54 This DELPHI-type consensus will facilitate the strengthening and harmonization of good
55 practice when performing HIV DNA sequencing.

56 **1. Introduction**

57 Human Immunodeficiency Virus-1 infection has become a manageable chronic disease with
58 the availability of antiretroviral therapies.^{1,2} Lifelong treatment is currently required to obtain
59 and maintain viral suppression. Either prior to initiation of ART or in the event of suboptimal
60 response to ART, HIV drug resistance testing using plasma HIV RNA plays a key role in
61 guiding treatment choices and optimization.^{1,2} When switching to a new ART regimen due to
62 toxicities, for simplification, drug reduction, or a long-acting regimen, it is also recommended
63 to first check HIV genotyping data.² In these situations, HIV viral load (VL) usually under 50
64 copies/mL does not allow amplification for RNA drug resistance testing.³

65 In recent years, there has been growing interest in how HIV drug resistance testing using
66 cellular HIV DNA could assist in clinical decision-making in the event of switching ART,
67 especially when plasma HIV RNA genotype testing is not possible.⁴⁻⁶ The 2022 European
68 AIDS Clinical Society (EACS) guidelines state that “Proviral DNA genotyping may be useful
69 in persons with multiple virologic failures, unavailable resistance history or low-level viremia
70 at the time of switch”.² European and French guidelines indicate that it is possible to perform
71 genotypic resistance tests on HIV DNA from Peripheral Blood Mononuclear Cells (PBMCs)
72 in the absence of historical data on plasma viral RNA.^{2,7} This test should be interpreted with
73 caution since it has a good positive predictive value but a low negative predictive value.⁷

74 However, while these guidelines provide general guidance on the indications for cell-
75 associated total HIV DNA resistance testing, practical recommendations to virologists and
76 HIV clinicians are lacking, particularly regarding frequent specific ART switch situations,
77 technique, and interpretation of results.

78 Since many disparities in clinical practice have been observed, both in the literature and in
79 clinical practice, a modified DELPHI consensus research project was conducted in France

80 with the aim of homogenizing situations in which HIV DNA sequencing could be used and
81 guiding interpretation of results.

82 **2. Material and methods**

83 The Delphi method is an iterative consensus approach based on information collected from a
84 panel of voters with expertise in the subject under consideration.⁸⁻¹⁶ This approach has been
85 widely used in many therapeutic areas and several times in HIV care.¹⁷⁻²⁸ Using this
86 structured approach, voting experts give their opinion individually and anonymously, and
87 express their degree of agreement on statements in order to achieve consensus on a specific
88 and well-defined subject.

89 In accordance with both French and international methodologies,^{9-12,29} our study was
90 structured as a modified national Delphi consensus and conducted among French hospital
91 clinicians and virologists between September and December 2022. The opinion of voting
92 experts was collected during two assessment rounds using a questionnaire developed by a
93 Steering Committee (SC) (Figure 1).

94 As recommended by the French National Authority for Health (HAS), voters specified their
95 level of agreement with the statements using a 9-point Likert scale ranging from 1 “Strongly
96 disagree” to 9 “Strongly agree”.²⁹⁻³¹ The percentage of scores and the median were calculated
97 for each statement separately in each voting round. Consensus for a statement was considered
98 ‘strong’ when >75% of the scores were ≥ 7 and the median score was ≥ 8 , ‘good’ when only
99 one of these two parameters was satisfied, and ‘lacking’ when none of the parameters was
100 satisfied.^{9,10,32}

101 *Steering Committee (SC)*

102 The SC included one infectious disease specialist and eight virologists directed by the last
103 author of this article. Two initial SC meetings were held in June and August 2022.

104 *Voting Group*

105 Two voter profiles were identified: virologists and HIV clinicians. A list of voters was
106 compiled based on the following criteria: experience, acquired knowledge and expertise in
107 HIV care, presenting in national conferences or involvement in HIV care projects, with
108 recruitment throughout France, including French overseas territories. The voters were invited
109 via individual e-mails to participate in online voting, with personalized access via a dedicated
110 website. Questions on techniques for performing HIV DNA genotyping were voted on by
111 virologists only. The anonymity of both voting groups was guaranteed. Voters had no
112 interaction with the SC, and SC members did not vote.²⁹

113 *Voting Round #1*

114 During this first round of voting, a free text space for comments was made available enabling
115 voters to develop or explain their opinion for each statement. At the end of the first round,
116 scores and voter comments were summarized for each statement.

117 A third SC meeting took place in November 2022 to discuss the Round #1 results:

- 118 • Statements that achieved a ‘strong’ consensus (i.e., $\geq 75\%$ of scores ≥ 7 AND median ≥ 8)
119 were validated in full and included in the final summary.
- 120 • Statements that achieved a ‘good’ consensus (i.e., $\geq 75\%$ of scores ≥ 7 OR median ≥ 8)
121 were discussed and proposed for Voting Round #2 only when the SC was able to develop
122 a revised version based on analysis of voter comments.
- 123 • Statements that did not achieve consensus were re-worded by the SC based on feedback
124 from voters and submitted for Voting Round #2.

125 *Voting Round #2*

126 Only voters from Voting Round #1 were invited to participate in Voting Round #2 to assess
127 the statements amended by the SC from Voting Round #1 results. The free text comment
128 option was deleted but replaced with an ‘I don’t know’ option instead of the scoring response.

129 Votes including this ‘I don’t know’ option were excluded from the analysis. Following the
130 results of Voting Round #2, the SC closed the process.

131 *Ethical considerations*

132 This research was conducted in accordance with the Declaration of Helsinki. All personal data
133 transmitted for the study was separated from the results and anonymized, pursuant to the
134 French data protection law (GDPR – General Data Protection Regulation).

135 **3. Results**

136 Based on a literature analysis, existing guidelines and clinical experience, the SC initially
137 developed 21 statements (two were subsequently merged resulting in 20 statements) divided
138 into 6 key areas: *clinical situations for the use of HIV DNA genotyping; techniques for*
139 *performing HIV DNA genotyping; consideration of APOBEC mutations; genotyping results*
140 *reporting; recycling of antiretrovirals; availability of HIV DNA genotyping tests and delays.*

141 *Participation*

142 Voters in Round #1 included 21 virologists and 47 clinicians. All virologists (21/21, 100%)
143 and 40 clinicians out of 47 (85.1%) from Round #1 actively voted in Round#2.

144 A summary of the characteristics of voters is shown in Table 1. The virologists were 76%
145 (n=16) full-time hospital workers, 10% (n=2) part-time and 14% (n=3) engineers (‘Others’).
146 Their median experience in performing HIV DNA sequencing was 10 years (IQR [5-12]) and
147 the median number of HIV DNA genotypes performed per year was 225 (IQR [42.5-425]).
148 Clinicians were 94% (n=44) full time hospital workers and 6% (n=3) part time. Their median
149 experience with people living with HIV (PLWH) management was 25 years (IQR [15-31.5])
150 and the median number of patients they followed per year was 270 (IQR [200-400]).

151 All virologists and clinicians had extensive experience in HIV care-related activities over the
152 previous five years, such as writing conference abstracts (76% and 83% respectively), writing
153 scientific publications (76% and 74%), participating in research projects (100% and 91%),

154 involved in training (81% and 81%), belonging to a professional or associated group (76%
155 and 81%) and speaking at scientific events (52% and 68%).

156 *Statements (Table 2)*

157 After Voting Round #1, 9/21 statements achieved a ‘strong’ consensus ($\geq 75\%$ votes ≥ 7 and
158 median ≥ 8); 5/21 statements achieved a ‘good’ consensus ($\geq 75\%$ votes ≥ 7 or median ≥ 8) and
159 7 statements lacked a consensus: 12 statements were revised by the SC for Voting Round #2,
160 including all those which achieved a ‘good’ consensus and all those which did not achieve a
161 consensus, of which two were merged resulting in 20 statements. After Voting Round #2,
162 9/11 revised statements achieved a ‘strong’ consensus, and two statements did not achieve a
163 consensus. In total, 18/20 statements (90%) achieved consensus. The distribution of
164 cumulative votes, medians and results are provided in Table 2.

165 *Consensus statement results (See Table S1 for Consensus results according to voter group,*
166 *and Table S2 for Statements, detailed virologists and clinicians voting results, and cumulative*
167 *results for both groups).*

168 Clinical situations for the use of HIV DNA genotyping

169 In the context of a therapeutic decision requiring genotyping data, there was a ‘strong’
170 consensus from voters on the recommendation to perform HIV DNA genotyping when HIV
171 RNA is non-amplifiable, when cumulative HIV RNA genotyping is not available and/or when
172 the historical genotype is incomplete or unusable. Voters recognized with a ‘strong’
173 consensus that for the following therapeutic targets - reverse transcriptase, protease, integrase
174 - HIV DNA sequencing has a good positive predictive value towards mutation detection
175 (excluding APOBEC mutations) and an imperfect negative predictive value.

176 Voters also ‘strongly’ agreed that, in a virologically suppressed patient, in the absence of
177 exploitable or sufficiently informative RNA data, and when considering a drug-reduction /
178 simplification of the antiretroviral (ARV) regimen:

179 - for a switch to some long-acting regimens, prior HIV DNA genotyping is useful for clinical
180 decision-making,

181 - for sequential dosing (4 days out of 7 or 5 days out of 7) without changing any ARV in the
182 current regimen, it is not mandatory to prior perform HIV DNA genotyping for clinical
183 decision-making,

184 - for a reduced ARV number regimen, prior HIV DNA genotyping may be useful for clinical
185 decision-making.

186 Techniques for performing HIV-DNA genotyping

187 Virologists validated with a ‘strong’ consensus that, in current practice, HIV DNA
188 genotyping has a decreased performance (sensitivity, representativeness of viral populations)
189 when the DNA quantity is very low. It can be performed indifferently from whole blood,
190 mononuclear cells isolated from peripheral blood or blood cell pellets, and although
191 performance could be increased by performing duplicate, duplicate is not feasible in clinical
192 practice.

193 Concerning HIV DNA genotyping techniques, virologists agreed with a ‘strong’ consensus
194 that Sanger or ultra-deep sequencing (UDS) could be used. However, there was an absence of
195 consensus on the relevance of discussing any viral minority population (i.e. variants below 15
196 to 20% of the viral population) detected after using UDS techniques in a multidisciplinary
197 meeting in the absence of defined clinically relevant detection threshold, according to the
198 current state of knowledge (‘no consensus’; with the exclusion of 1/21 (4.7%) virologists who
199 answered ‘I don’t know’).

200 Consideration of APOBEC mutations

201 The cytidine deaminases APOBEC3F and 3G enzymes might introduce G to A nucleotide
202 mutations that can impair crucial enzymatic sites or generate stop codons that reduce the
203 amount of replication competent proviruses.³³⁻³⁶ Voters validated with a ‘strong’ consensus

204 that the detection of the *M184I* mutation in HIV DNA is suggestive of the presence of a
205 defective genome in the APOBEC enzyme when associated with other evocative mutations
206 (e.g., *M41I*, *M230I* on reverse transcriptase) and/or stop codons. They also recognized with a
207 ‘strong’ consensus that, when resistance mutations attributable to APOBEC are present, their
208 significance should be interpreted with caution according to the clinical context and
209 therapeutic history of the patient and should be indicated in the HIV DNA genotyping
210 analysis report.

211 Reporting of genotyping results

212 With a ‘strong’ consensus, virologists and clinicians felt that the clinical interpretation of
213 resistance mutations on HIV DNA genotyping should be discussed in multidisciplinary
214 meetings. They also agreed that the detection via HIV DNA sequencing of new resistance
215 mutations (excluding APOBEC and stop codons), which were previously undetected, must be
216 considered for the switch decision and subsequent patient follow-up.

217 ARV recycling

218 Virologists and clinicians agreed with a ‘strong’ consensus that, in a patient with an
219 undetectable viral load for at least one year and with documented *M184V* substitution on the
220 current DNA genotype and/or on an RNA genotype performed within the last five years, the
221 use of a 2nd generation InSTI (Integrase Strand Transfer Inhibitor) + XTC (Lamivudine or
222 Emtricitabine) + 1 NRTI combination is at low risk of virological failure over time.
223 Virologists validated with a ‘strong’ consensus that, under the same conditions, the use of a
224 2nd generation InSTI + XTC combination may present a risk of virological failure over time.
225 However, clinicians remained divided on this possible virological risk and their vote did not
226 reach a consensus (‘no consensus’, no clinicians answered ‘I don't know’).

227 With a ‘strong’ consensus, virologists and clinicians validated that the use/recycling of
228 NNRTIs, if resistance to this class was detected in HIV-DNA and/or in previous historical
229 genotypes, is associated with a greater risk of virological failure, independently of the

230 duration of undetectable viral load, particularly in drug-reduction strategies using this ARV
231 class.

232 Availability of HIV DNA genotyping tests and time to report results

233 With a ‘strong’ consensus, virologists and clinicians felt that genotypic HIV DNA testing
234 should be accessible in clinical practice to all clinicians managing PLWH, and that results
235 from these tests should be available within 30 days.

236 **4. Discussion**

237 This consensus research, using the DELPHI method, aims at harmonizing HIV DNA
238 sequencing practices.

239 All five assertions on clinical situations for use of HIV DNA genotyping developed by the SC
240 were validated with a ‘strong’ consensus by the voters. Although HIV DNA sequencing is not
241 routinely recommended² and does not systematically reveal the same results as those
242 previously detected by cumulative plasma RNA genotyping in virologically controlled
243 patients,^{4,5} it is useful to perform in several clinical circumstances. This is the case when
244 historical HIV RNA resistance data are insufficient and/or incomplete, or when the viral load
245 is too low to proceed with HIV RNA sequencing. A recent study - based on a very large
246 genotypic database in France - describing the prevalence of genotypic baseline risk factors for
247 some long-acting regimen failures among ARV-naive patients showed that 10.1% of patients
248 displayed one baseline virological risk factor for virologic failure.³⁷ These findings emphasize
249 the need to check the genotypic resistance profile prior to initiating a long-acting regimen to
250 limit the potential risk of virologic failure and the emergence of resistance.

251 However, in the case of a virologically suppressed patient, in the event of a decision to reduce
252 or simplify sequential treatment (4 days or 5 days out of 7) without changing the regimen,
253 there was a ‘strong’ consensus that prior genotyping of HIV DNA is not essential to clinical
254 decision-making, even in the absence of usable or sufficiently informative RNA data. This

255 matches literature findings showing that triple combination therapy of a 2nd generation InSTI
256 + XTC + 1 NRTI administered every 4 or 5 days maintains control of HIV replication in
257 virologically suppressed PLWH while reducing cumulative exposure to ARV.^{38,39}

258 There was a ‘strong’ consensus from virologist voters that HIV DNA sequencing should be
259 performed when the viral quantity is sufficiently high (since the quantity of HIV DNA
260 influences the quality of the results obtained), that it can be used from different blood sample
261 matrix, indifferently by Sanger or UDS, and that duplicates increase test performance
262 (although this cannot be used in current clinical practice). Nevertheless, knowing HIV-DNA
263 genotyping underestimate resistance detection due to a phenomenon of dilution of resistant
264 species in the reservoir regardless of the sequencing method used, UDS methods might
265 improve resistance detection in HIV-DNA due to their greater sensitivity.⁴⁰⁻⁴² Virologists
266 were unable to reach a consensus on the fact that, given the current state of knowledge, it may
267 be worthwhile reporting any minority viral population detected for discussion in a
268 multidisciplinary discussion. They also didn’t support the idea that it might be useful to report
269 any minority viral population detected for multidisciplinary discussion in the current context
270 of an undefined detection threshold for UDS techniques.

271 Although the 1% threshold for UDS techniques was found to be close to the sensitivity
272 obtained in historical HIV RNA resistance tests,⁴¹ it was difficult for the SC to generate a
273 statement for voting with such a detection threshold. This is due to the variability of this
274 threshold depending on the UDS technique used, and the lack of solid evidence on the impact
275 of a minority variant as low as 1% on virological failure for newer ARVs with a high barrier
276 to resistance. Considering that UDS on HIV-DNA is now affordable in clinical practice and
277 may become the future the potential new gold standard in the future, the definition of a
278 technical cut-off to warrant enough sequencing accuracy and a clinical cut-off to establish the
279 clinical relevance of minority variants on treatment switch in virologically suppressed patients
280 are still unmet needs. So further research into these thresholds for both RNA- and DNA-based
281 techniques is warranted.

282 As shown in the literature,^{43,44} the detection of the *M184I* mutation in HIV DNA suggests the
283 presence of a defective genome due to the APOBEC enzyme when associated with other
284 suggestive mutations (See Table S3 for the list of mutations)³⁶ and/or stop codons, and a
285 ‘strong’ consensus was reached on this statement. The presence of *M184I* mutation can impair
286 the activity of XTC and possibly some nucleoside reverse transcriptase translocation
287 inhibitors (NRTTIs). These mutations should be considered possible artifacts if they occur at
288 the same threshold at which multiple signature APOBEC mutations are also present.⁴⁵ When
289 resistance mutations attributable to APOBEC are detected, it is recommended that their
290 significance should be interpreted with caution³⁷ and should be indicated in the HIV DNA
291 genotyping analysis.

292 The French National Authority for Health already recommends that the interpretation of
293 results from a DNA-based genotypic resistance test requires consultation between clinician
294 and virologist.⁴⁶ In this context, a ‘strong’ consensus was reached on the need to discuss
295 clinical interpretation of resistance mutations obtained by HIV DNA genotyping at
296 multidisciplinary discussions. This was also the case regarding clinical decisions about
297 switching ART and patient follow-up in newly detected resistance mutations.

298 The question of how resistance mutations are 'archived' over time remains important for the
299 potential re-use of specific ARVs. A recent study investigated the kinetics of the *M184V*
300 mutation in proviral HIV DNA in long-term virologically suppressed patients.⁴⁷ The authors
301 showed significant progressive clearance of the *M184V* mutation in proviral HIV DNA over
302 the five years of the study. In the presence of a detected M184V substitution over the past 5
303 years, the SC looked for consensus statements on ARV recycling practices. In this context,
304 the SC proposed statements on ARV recycling practices in the event of the presence of an
305 M184V substitution detected within the last five years.

306 Regardless of the finding of an *M184V* mutation in the DNA genotype and clearance kinetics
307 of the mutation, it has been observed that, in patients virologically suppressed for at least one
308 year, the use of a 2nd generation InSTI + XTC + 1 NRTI regimen presents a low risk of

309 virological failure over time.⁴⁸ The voters ‘strongly’ endorsed this statement. However, when
310 a M184V mutation has been documented over the past five years in a virologically suppressed
311 patient, the virologist voters ‘strongly’ agreed that the use of a 2nd generation InSTI + XTC
312 regimen could present a risk of virological failure over time, as described in some literature.⁴⁹
313 For documented NNRTI mutations, there was a ‘strong’ consensus that the recycling of this
314 ARV class is associated with an increased risk of virological failure, irrespective of the
315 duration of viral suppression, particularly in drug reduction strategies and long-acting
316 regimens using this ARV class.⁵⁰
317 Since HIV DNA sequencing adds an important contribution to many clinical situations and
318 patient follow-up,² there was a ‘strong’ consensus that it should be accessible to all
319 practitioners. Also, that its results should be received within one month. The literature rarely
320 provides such an indication of time in which to report results but, with current HIV DNA
321 sequencing methods being faster than before, this timeframe seems reasonable.⁵¹
322 The Delphi method is known as a structured procedure which enables many experts to be
323 consulted individually and anonymously on a specific subject while guaranteeing free
324 expression of each voter. However, this approach has some limitations associated with voters’
325 profiles, statements elaboration and criteria considered to achieve a consensus.⁵² Our research
326 sought to limit these potential biases as far as possible to ensure maximum objectivity.
327 Although voters were recruited only in France, they were selected on objective criteria based
328 on their experience and expertise in HIV care and HIV virology. These criteria yielded a voter
329 sample with reassuring characteristics: a median of 10 years’ experience performing HIV
330 DNA sequencing in the virologists’ group and a median of 25 years’ experience in PLWH
331 management in the clinicians’ group. As far as the SC statements are concerned, a literature
332 review made it possible to identify key questions raised in clinical practice and propose
333 precisely worded statements. In terms of the threshold used to reach consensus, our study was
334 based on a rigorous two-criteria approach. This strict and demanding definition lends a high
335 degree of credibility to our results. To ensure the virologist panel represented the whole of

336 France territory, the SC supported identification of some virologists. Finally, our research was
337 conducted with a continuous and complete separation between voters who voted
338 anonymously and SC members who neither participated in the vote nor interacted directly
339 with voters. The constraint inherent in this separation was the absence of direct exchanges
340 between voters and SC members: such exchanges could have been useful when revising
341 statements for voting Round #2. Furthermore, like all Delphi-type consensus, the findings
342 represent good practices for virologists and clinicians who remain masters of their own
343 practice and must adapt findings to individual patient circumstances.

344 In conclusion, in this consensus research using the Delphi method, 18/20 (90%) statements
345 achieved a consensus. Only two assertions did not reach consensus. Virologist voters
346 remained divided on the value of discussing any minority population detected at a
347 multidisciplinary meeting, and the clinician voters remained divided on the possible
348 virological risk of using a combination of a 2nd generation InSTI + XTC in HIV suppressed
349 patients of more than one year in the presence of a documented M184V mutation of less than
350 five years.

351 Our consensus findings constitute a solid basis for implementation and homogenization of
352 practice regarding the use of DNA HIV sequencing, its performance, and its reporting,
353 particularly when needing to reduce the number of ARV agents and when using some long-
354 acting regimens.

355 **Acknowledgements**

356 The SC would like to thank all virologists and clinicians who agreed to participate and vote
357 on the statements in both rounds, and who made it possible to achieve this consensus.

358 **Funding**

359 This consensus research was funded by Gilead Sciences.

360 **Transparency declarations**

361 Carried out with the institutional support of Gilead Sciences S.A.S. Operational and
362 publishing aspects of the interviews were managed by Medica Education Corpus agency.

363 Elisabeth ANDRE-GARNIER reports grants from Gilead Sciences, MSD and ViiV
364 Healthcare

365 Laurence BOCKET reports grants from Gilead Sciences, MSD and ViiV Healthcare

366 Thomas BOURLET reports grants from MSD and GILEAD Sciences, not in the scope of the
367 present work.

368 Laurent HOCQUELOUX reports non-financial support from Gilead Sciences, MSD and ViiV
369 Healthcare, payments for advisory board participation from Gilead Sciences and ViiV
370 Healthcare, and personal fees from Gilead Sciences, MSD and ViiV Healthcare, all outside
371 the submitted work.

372 Quentin LEPILLER reports grants from Gilead Sciences, MSD and ViiV Healthcare.

373 Anne MAILLARD reports grants from Gilead Sciences, MSD and ViiV Healthcare.

374 Sandrine REIGADAS, Guillaume BARRIERE and François DURAND are employees of
375 Gilead Sciences S.A.S.

376 Brigitte MONTES reports grants from Gilead Sciences, MSD and ViiV Healthcare.

377 Karl STEFIC reports payments for advisory board participation from Gilead Sciences and
378 ViiV Healthcare.

379 Anne-Geneviève MARCELIN reports grants from Gilead Sciences, MSD and ViiV
380 Healthcare, payments for advisory board participation from Gilead Sciences, MSD and ViiV
381 Healthcare.

382 **References**

383 1. Panel on Antiretroviral Guidelines for Adults and Adolescents. Department of Health
384 and Human Services. *Guidelines for the use of antiretroviral agents in adults and*

- 385 *adolescents with HIV.*
- 386 [https://clinicalinfo.hiv.gov/sites/default/files/guidelines/documents/adult-adolescent-](https://clinicalinfo.hiv.gov/sites/default/files/guidelines/documents/adult-adolescent-arv/guidelines-adult-adolescent-arv.pdf)
- 387 [arv/guidelines-adult-adolescent-arv.pdf](https://clinicalinfo.hiv.gov/sites/default/files/guidelines/documents/adult-adolescent-arv/guidelines-adult-adolescent-arv.pdf)
- 388 2. European AIDS Clinical Society 2022. *GUIDELINES Version 11.1.*
- 389 https://www.eacsociety.org/media/guidelines-11.1_final_09-10.pdf
- 390 3. Assoumou L, Charpentier C, Recordon-Pinson P *et al.* Prevalence of HIV-1 drug
- 391 resistance in treated patients with viral load ≥ 50 copies/mL: a 2014 French nationwide
- 392 study. *J Antimicrob Chemother* 2017; **72**: 1769-1773.
- 393 4. Wirlden M, Soulie C, Valantin MA, *et al.* Historical HIV-RNA resistance test results
- 394 are more informative than proviral DNA genotyping in cases of suppressed or residual
- 395 viraemia. *J Antimicrob Chemother* 2011; **66**: 709-12.
- 396 5. Delaugerre C, Braun J, Charreau I, *et al.* Comparison of resistance mutation patterns
- 397 in historical plasma HIV RNA genotypes with those in current proviral HIV DNA
- 398 genotypes among extensively treated patients with suppressed replication. *HIV Med*
- 399 2012; **13**: 517-25.
- 400 6. Boukli N, Boyd A, Collot M *et al.* Utility of HIV-1 DNA genotype in determining
- 401 antiretroviral resistance in patients with low or undetectable HIV RNA viral loads. *J*
- 402 *Antimicrob Chemother* 2018; **73**: 3129-3136.
- 403 7. Morlat, C. Groupe d'experts pour la prise en charge du VIH 2017. *Optimisation d'un*
- 404 *traitement antirétroviral en situation de succès virologique. Prise en charge médicale*
- 405 *des personnes vivant avec le VIH.* [https://cns.sante.fr/actualites/prise-en-charge-du-](https://cns.sante.fr/actualites/prise-en-charge-du-vih-recommandations-du-groupe-dexperts/)
- 406 [vih-recommandations-du-groupe-dexperts/](https://cns.sante.fr/actualites/prise-en-charge-du-vih-recommandations-du-groupe-dexperts/)
- 407 8. Dalkey NC. The Delphi method: An experimental study of group opinion. In N. C.
- 408 Dalkey, D. L. Rourke, R. Lewis, & D. Snyder (Eds.). *Studies in the quality of life:*
- 409 *Delphi and decision-making.* Lexington, MA: Lexington Books, 1972; 13-54.

- 410 9. Loblaw DA, Prestrud AA, Somerfield MR *et al.* American Society of Clinical
411 Oncology Clinical Practice Guidelines: formal systematic review-based consensus
412 methodology. *J Clin Oncol* 2012; **30**: 3136-3140.
- 413 10. Boukdedid R, Abdoul H, Loustau M *et al.* Using and Reporting the Delphi Method for
414 Selecting Healthcare Quality Indicators: A Systematic Review. *PLoS One* 2011; **6**:
415 e20476.
- 416 11. Diamond IR, Grant RC, Feldman BM *et al.* Defining consensus: a systematic review
417 recommends methodologic criteria for reporting of Delphi studies. *J Clin Epidemiol*
418 2014; **67**: 401-9.
- 419 12. Hasson F, Keeney S, McKenna H. Research guidelines for the Delphi survey
420 technique. *J Adv Nurs* 2000; **32**: 1008-15.
- 421 13. Hsu CC, Sandford BA. The Delphi Technique: Making Sense of Consensus. *Practical*
422 *Assessment Research & Evaluation* 2019; **12**: 1-10.
- 423 14. Humphrey-Murto S, Varpio L, Wood TJ *et al.* The Use of the Delphi and Other
424 Consensus Group Methods in Medical Education Research: A Review. *Acad Med*
425 2017; **92**: 1491-1498.
- 426 15. Richard MA, Aubin F, Beneton N *et al.* Moderate Psoriasis in Clinical Practice:
427 French Expert Consensus Using a Modified Delphi Method. *Adv Ther* 2022; **39**: 5203-
428 5215
- 429 16. Kodjikian L, Baillif S, Couturier A *et al.* Recommendations for the management of
430 diabetic macular oedema with intravitreal dexamethasone implant: A national Delphi
431 consensus study. *Eur J Ophthalmol* 2022; **32**: 2845-2856
- 432 17. Tsui S, Denison JA, Kennedy CE *et al.* Identifying models of HIV care and treatment
433 service delivery in Tanzania, Uganda, and Zambia using cluster analysis and Delphi
434 survey. *BMC Health Serv Res* 2017; **17**: 811.

- 435 18. Johnson MO, Neilands TB, Koester KA, *et al.* Detecting Disengagement From HIV
436 Care Before It Is Too Late: Development and Preliminary Validation of a Novel Index
437 of Engagement in HIV Care. *J Acquir Immune Defic Syndr* 2019; **81**: 145-152.
- 438 19. O'Connell KA, Kisteneff AV, Gill SS *et al.* HIV post-exposure prophylaxis in the
439 emergency department: An updated assessment and opportunities for HIV prevention
440 identified. *Am J Emerg Med* 2021; **46**: 323-328.
- 441 20. Fredericksen RJ, Edwards TC, Merlin JS *et al.* Patient and provider priorities for self-
442 reported domains of HIV clinical care. *AIDS Care* 2015; **27**: 1255-64
- 443 21. Cummins D, Waters D, Aggar C *et al.* Assessing Risk of HIV-Associated
444 Neurocognitive Disorder. *Nurs Res* 2019; **68**: 22-28.
- 445 22. Greacen T, Kersaudy-Rahib D, Le Gall JM *et al.* Comparing the Information and
446 Support Needs of Different Population Groups in Preparation for 2015 Government
447 Approval for HIV Self-testing in France. *PLoS One* 2016; **11**: e0152567.
- 448 23. Feyissa GT, Lockwood C, Woldie M *et al.* Evaluation of a guideline developed to
449 reduce HIV-related stigma and discrimination in healthcare settings and establishing
450 consensus. *PLoS One* 2018; **13**: e0198781.
- 451 24. Johnson MO Koester KA, Wood T *et al.* Development of an Index of Engagement in
452 HIV Care: An Adapted Internet-Based Delphi Process. *JMIR Res Protoc* 2017; **6**:
453 e224.
- 454 25. Uyei J, Li L, Braithwaite RS. HIV and Alcohol Research Priorities of City, State, and
455 Federal Policymakers: Results of a Delphi Study. *Am J Public Health* 2015; **105**: e23-
456 6.
- 457 26. Maserati R, Antinori A, Bonora S *et al.* Optimizing HIV therapy. A consensus project
458 on differences between cytidine analogues and regime compactness. *New Microbiol*
459 2014; **37**: 285-306
- 460 27. Adegbehingbe SM, Paul-Ebhohimhem V, Marais D. Development of an AFASS
461 assessment and screening tool towards the prevention of mother-to-child HIV

- 462 transmission (PMTCT) in sub-Saharan Africa-a Delphi survey. *BMC Public Health*
463 2012; **12**: 402.
- 464 28. Engler K, Ahmed S, Lessard D *et al.* Assessing the Content Validity of a New Patient-
465 Reported Measure of Barriers to Antiretroviral Therapy Adherence for Electronic
466 Administration in Routine HIV Care: Proposal for a Web-Based Delphi Study. *JMIR*
467 *Res Protoc* 2019; **8**: e12836.
- 468 29. HAS (Haute Autorité de Santé) Guide méthodologique. 2010. *Élaboration de*
469 *recommandations de bonne pratique. Méthode « Recommandations pour la pratique*
470 *clinique »*. [https://www.has-sante.fr/upload/docs/application/pdf/2011-](https://www.has-sante.fr/upload/docs/application/pdf/2011-01/guide_methodologique_consensus_formalise.pdf)
471 [01/guide_methodologique_consensus_formalise.pdf](https://www.has-sante.fr/upload/docs/application/pdf/2011-01/guide_methodologique_consensus_formalise.pdf)
- 472 30. Letrilliart L, Milliat-Guittard L, Romestaing P *et al.* Building a shared patient record
473 for breast cancer management: a French Delphi study. *Eur J Cancer Care (Engl)*
474 2009; **18**: 131-9.
- 475 31. McMillan SS, King M, Tully MP. How to use the nominal group and Delphi
476 techniques. *Int J Clin Pharm* 2016; **38**: 655-62.
- 477 32. Koene S, van Bon L, Bertini E *et al.* Outcome measures for children with
478 mitochondrial disease: consensus recommendations for future studies from a Delphi-
479 based international workshop. *J Inherit Metab Dis* 2018; **41**: 1267-1273
- 480 33. Mangeat B, Turelli P, Caron G *et al.* Broad antiretroviral defence by human
481 APOBEC3G through lethal editing of nascent reverse transcripts. *Nature* 2003; **424**:
482 99–103
- 483 34. Armitage AE, Deforche K, Chang CH *et al.* APOBEC3G-induced hypermutation of
484 human immunodeficiency virus type-1 is typically a discrete “all or nothing”
485 phenomenon. *PLoS Genet* 2012; **8**: e1002550.
- 486 35. Russell RA, Moore MD, Hu WS *et al.* APOBEC3G induces a hypermutation gradient:
487 purifying selection at multiple steps during HIV-1 replication results in levels of G-to-

488 A mutations that are high in DNA, intermediate in cellular viral RNA, and low in
489 virion RNA. *Retrovirology* 2009; **6**: 16.

490 36. Armenia D, Gagliardini R, Alteri *et al.* Temporal trend of drug-resistance and
491 APOBEC editing in PBMC genotypic resistance tests from HIV-1 infected
492 virologically suppressed individuals. *J Clin Virol* 2023; **168**: 105551.

493 37. Charpentier C, Storto A, Soulié C *et al.* Prevalence of genotypic baseline risk factors
494 for cabotegravir + rilpivirine failure among ARV-naive patients. *J Antimicrob*
495 *Chemother* 2021; **76**: 2983-2987.

496 38. Sellem B, Abdi B, Lê M *et al.* Intermittent Bictegravir/Emtricitabine/Tenofovir
497 Alafenamide Treatment Maintains High Level of Viral Suppression in Virally
498 Suppressed People Living with HIV. *J Pers Med* 2023; **13**: 583.

499 39. Landman R, de Truchis P, Assoumou L, *et al.* A 4-days-on and 3-days-off
500 maintenance treatment strategy for adults with HIV-1 (ANRS 170 QUATUOR): a
501 randomised, open-label, multicentre, parallel, non-inferiority trial. *Lancet HIV* 2022;
502 **9**: e79-e90.

503 40. St John EP, Simen BB, Turenchalk GS *et al.* A Follow-Up of the Multicenter
504 Collaborative Study on HIV-1 Drug Resistance and Tropism Testing Using 454 Ultra
505 Deep Pyrosequencing. *PLoS ONE* 2016; **11**: e0146687.

506 41. Rodriguez C, Nere ML, Demontant V *et al.* Ultra-deep sequencing improves the
507 detection of drug resistance in cellular DNA from HIV-infected patients on ART with
508 suppressed viraemia. *J Antimicrob Chemother* 2018; **73**: 3122-3128.

509 42. Balakrishna S, Loosli T, Zaheri M *et al.* Frequency matters: comparison of drug
510 resistance mutation detection by Sanger and next-generation sequencing in HIV-1. *J*
511 *Antimicrob Chemother* 2023; **78**: 656–664.

512 43. Charpentier C., Montes B., Perrier M *et al.* HIV-1 DNA ultra-deep sequencing
513 analysis at initiation of the dual therapy dolutegravir + lamivudine in the maintenance
514 DOLULAM pilot study. *J Antimicrob Chemother* 2017; **72**: 2831-2836.

- 515 44. Allavena C, Rodallec A, Leplat A, *et al.* Interest of proviral HIV-1 DNA genotypic
516 resistance testing in virologically suppressed patients candidate for maintenance
517 therapy. *J Virol Methods* 2018; **251**: 106-110.
- 518 45. Tzou PL, Kosakovsky Pond SL, Avila-Rios S *et al.* Analysis of unusual and signature
519 APOBEC-mutations in HIV-1 pol next-generation sequences. *PLoS One* 2020; **15**:
520 e0225352.
- 521 46. Morlat, C. Groupe d'experts pour la prise en charge du VIH. Résistance du VIH-1 aux
522 antirétroviraux. 2016. *Prise en charge médicale des personnes vivant avec le VIH*.
523 https://cns.sante.fr/wp-content/uploads/2017/02/experts-vih_resistance.pdf
- 524 47. Palich R, Teyssou E, Sayon S *et al.* Kinetics of Archived M184V Mutation in
525 Treatment-Experienced Virally Suppressed HIV-Infected Patients. *J Infect Dis* 2022;
526 **225**: 502-509.
- 527 48. Andreatta K, Willkom M, Martin R *et al.* Switching to
528 bictegravir/emtricitabine/tenofovir alafenamide maintained HIV-1 RNA suppression
529 in participants with archived antiretroviral resistance including M184V/I. *J*
530 *Antimicrob Chemother* 2019; **74**: 3555-3564.
- 531 49. Santoro MM, Armenia D, Teyssou E *et al.* Virological efficacy of switch to DTG plus
532 3TC in a retrospective observational cohort of suppressed HIV-1 patients with or
533 without past M184V: the LAMRES study. *J Glob Antimicrob Resist* 2022; **31**: 52-62.
- 534 50. Marcelin AG, Soulie C, Wirlden M *et al.* The Virostar study: analysis of emergent
535 resistance-associated mutations at first- or second-line HIV-1 virological failure with
536 second-generation InSTIs in two- and three-drug regimens. *HIV Glasgow congress*
537 2022. Abstract P225.
- 538 51. Alidjinou EK, Deldalle J, Hallaert C *et al.* RNA and DNA Sanger sequencing versus
539 next-generation sequencing for HIV-1 drug resistance testing in treatment-naïve
540 patients. *Antimicrob Chemother* 2017; **72**: 2823-2830.

541 52. Skinner R, Nelson RR, Chin WW *et al.* The Delphi Method Research Strategy in
542 Studies of Information Systems. *Commun Assoc Inf Syst* 2015; **37**: 31-63.

Figure 1: Modified Delphi process chart

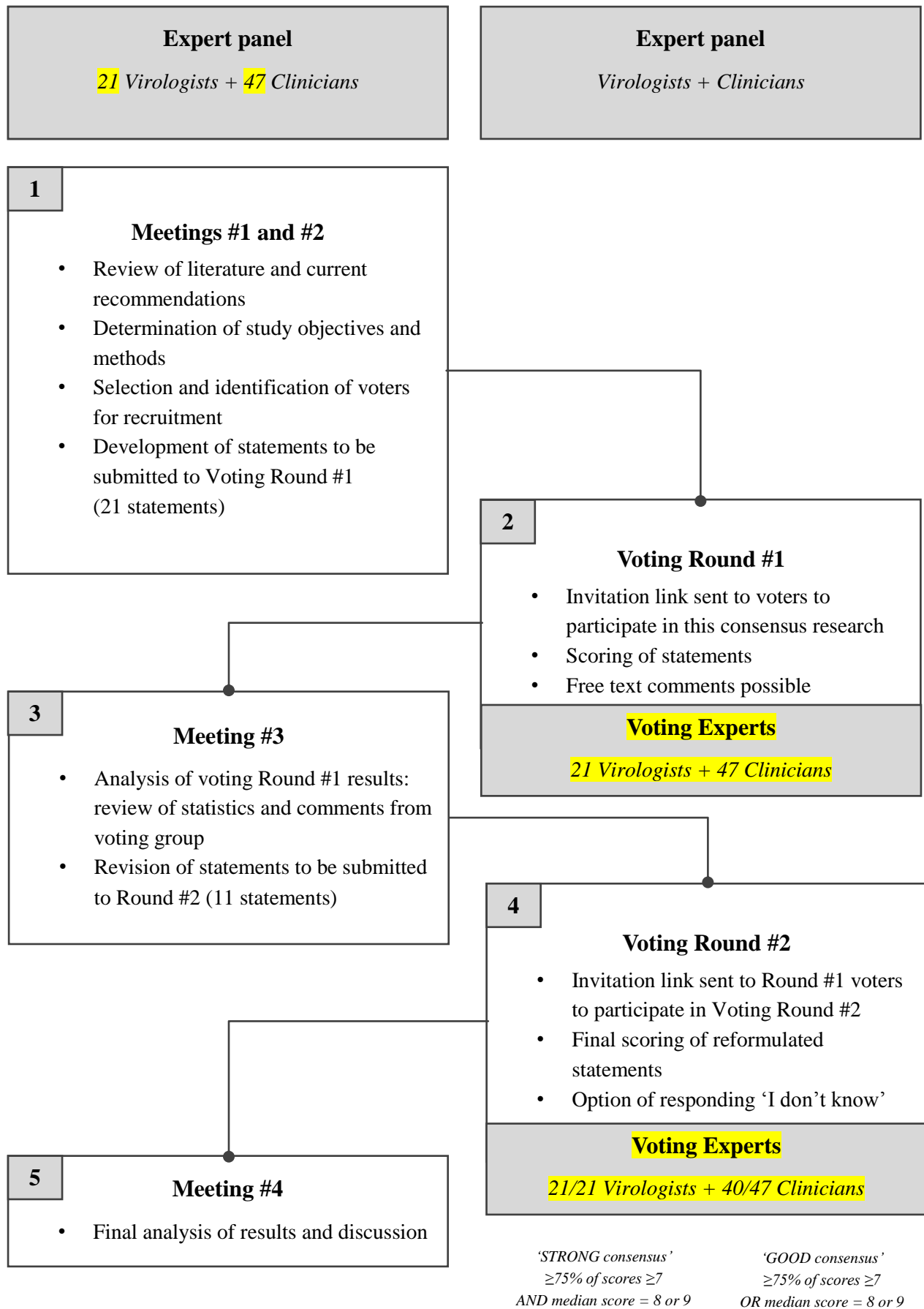


Table 1 – Characteristics of voters

Characteristic	Virologists (n = 21)	Clinicians (n = 47)
Age, median [IQR], years	46 [43-55]	56 [46-60.5]
Gender F/M, n (%)		
Female	16 (76)	18 (38)
Male	5 (24)	29 (62)
Type of practice, n (%)		
Full-time hospital workers	16 (76)	44 (94)
Part-time hospital workers	2 (10)	3 (6)
Others	3 (14)	-
Years of experience performing HIV DNA sequencing, median [IQR], years	10 [5-12]	-
Number of HIV DNA genotypes performed per year, median [IQR]	225 [42.5-425]	-
Years of experience in PLWH management, median [IQR], years	-	25 [15-31.5]
Number of PLWH seen per year, median [IQR]	-	270 [200-400]
Experience in HIV care-related activities in the past 5 years, n (%)		
Conference abstract	16 (76)	39 (83)
Scientific article	16 (76)	35 (74)
Research project (not including this study)	21 (100)	43 (91)
Involved in training	17 (81)	38 (81)
Professional or associate group or member	16 (76)	38 (81)
Speaker at scientific events	11 (52)	32 (68)

DNA: deoxyribonucleic acid; F: Female; M: Male; PLWH: People living with HIV

Table 2 - Statements and cumulative voting results for virologists and clinicians

For each statement, a total number of voters equaling 21 indicates that only virologists were invited to vote and a total number of voters different from 68, 61, or 21 indicates the use of the 'I don't know' option by voters during the second voting round.

	STATEMENTS	Scores 1-2-3 (n)	Scores 4-5-6 (n)	Scores 7-8-9 (n)	Median	Results
Clinical situations for the use of HIV DNA genotyping						
1	In the context of a therapeutic decision requiring genotyping data, when HIV RNA is not amplifiable, when cumulative HIV RNA genotyping is not available and/or in the event of an incomplete or unusable genotypic history, HIV DNA genotyping is recommended.	2.9% (2)	7.4% (5)	89.7% (61)	9	Strong consensus
2	For the following therapeutic targets - reverse transcriptase, protease, integrase - HIV DNA sequencing has a good positive predictive value (excluding APOBEC mutations) and an imperfect negative predictive value.	2.9% (2)	13.2% (9)	83.8% (57)	8	Strong consensus
3	In the context of a patient who has achieved virological success, in the event of a decision to reduce or simplify treatment to some long-acting regimens, prior genotyping on HIV DNA is useful for clinical decision-making in the absence of usable or sufficiently informative RNA data.	5.9% (4)	13.2% (9)	80.9% (55)	8	Strong consensus
4	In the context of a patient with virological success, in the event of a decision to reduce or simplify sequential treatment (4 days out of 7 or 5 days out of 7) without changing the treatment molecules, prior genotyping on HIV DNA is not essential for the clinical decision, even in the absence of usable or sufficiently informative RNA data.	13.3% (8)	10% (6)	76.7% (46)	8	Strong consensus
5	In the absence of usable or sufficiently informative RNA data, in the case of a patient with virological success, in the event of a decision to reduce/simplify to a treatment that reduces the number of ARV, prior genotyping on HIV DNA may be useful for clinical decision-making.	8.2% (5)	13.1% (8)	78.7% (48)	8	Strong consensus
Techniques for performing HIV DNA genotyping						
6	In current practice, when the amount of DNA is very low, the performance (sensitivity, representativeness of viral populations) of HIV DNA genotyping is reduced.	5.3% (1)	5.3% (1)	89.5% (17)	9	Strong consensus
7	In current practice, genotyping on HIV DNA can be performed either from whole blood, from the cells isolated from peripheral blood (PBMC) or from blood cell pellets.	14.3% (3)	9.5% (2)	76.2% (16)	8	Strong consensus
8	Duplicate DNA genotyping increases performance (sensitivity, representativeness of viral populations) but is not possible in current practice.	10.5% (2)	5.3% (1)	84.2% (16)	8	Strong consensus
9	Sanger and ultra-deep sequencing can be used to perform HIV DNA genotyping.	0% (0)	23.8% (5)	76.2% (16)	8	Strong consensus
10	For ultra-high throughput DNA sequencing techniques (ultra-deep sequencing), with the current state of knowledge, the clinically relevant detection threshold is not defined. Nevertheless, it may be interesting to report any viral minority population detected for multidisciplinary discussions.	10% (2)	25% (5)	65% (13)	7.5	NO CONSENSUS
Consideration of APOBEC mutations						
11	Detection of the <i>M184I</i> mutation in HIV DNA is suggestive of the presence of a defective genome due to the APOBEC enzyme when it is associated with other suggestive mutations (e.g., <i>M41I</i> , <i>M230I</i> on reverse transcriptase) and/or stop codons.	1.9% (1)	13.2% (7)	84.9% (45)	8	Strong consensus
12	When resistance mutations attributable to APOBEC are present, their significance should be interpreted with caution and based on the clinical context and treatment history.	5.2% (3)	5.2% (3)	89.7% (52)	9	Strong consensus
13	The presence of resistance mutations attributable to APOBEC should be reported in the HIV DNA genotyping analysis.	9.5% (2)	4.8% (1)	85.7% (18)	8	Strong consensus
Reporting of genotyping results						
14	The clinical interpretation of resistance mutations on HIV DNA genotyping should be discussed at multidisciplinary consultation meetings.	4.4% (3)	17.2% (10)	80.9% (55)	8	Strong consensus
15	In a patient with virological success, the detection on HIV DNA genotyping of new resistance mutations (excluding APOBEC & stop codons) previously undetected must be considered for the	5% (3)	10% (6)	85% (51)	8	Strong consensus

	switch decision and subsequent follow-up.					
ARV recycling						
16	In a patient with an undetectable viral load for at least one year, in the presence of a documented M184V substitution over the past five years, the use of a 2 nd generation InSTI + XTC + 1 NRTI combination presents low risk of virological failure over time.	7.4% (5)	7.4% (5)	85.3% (58)	8	Strong consensus
17	In a patient with an undetectable viral load for at least one year, in the presence of a documented M184V substitution over the past five years, the use of a 2 nd generation InSTI + XTC combination might present a risk of virological failure over time.	21.3% (13)	14.8% (9)	63.9% (39)	7	NO CONSENSUS
18	The use / recycling of NNRTIs in the event of documented resistance to ARV of this class is associated with a greater risk of virological failure, independently of the duration of undetectable viral load, especially in drug-reduction strategies using this ARV class.	1.7% (1)	18.3% (11)	80% (48)	8	Strong consensus
Test availability and delays						
19	HIV DNA genotypic tests should be accessible in clinical practice to all clinicians managing PLWH.	8.8% (6)	7.4% (5)	83.8% (57)	9	Strong consensus
20	Reports of genotypic HIV DNA test results should be sent to clinicians within 30 days.	6.6% (4)	9.8% (6)	83.6% (51)	9	Strong consensus

ARV: Antiretrovirals; APOBEC: Apolipoproteins B mRNA editing enzyme; InSTI: Integrase Strand Transfer Inhibitor; XTC: Lamivudine or Emtricitabine; PLWH: People living with HIV.

Table S1 – Consensus results according to voter groups

	STATEMENTS	Results VIROLOGISTS	Results CLINICIANS	CUMULATIVE Results
Clinical situations for the use of HIV DNA genotyping				
1	In the context of a therapeutic decision requiring genotyping data, when HIV RNA is not amplifiable, when cumulative HIV RNA genotyping is not available and/or in the event of an incomplete or unusable genotypic history, HIV DNA genotyping is recommended.	Strong consensus	Strong consensus	STRONG CONSENSUS
2	For the following therapeutic targets - reverse transcriptase, protease, integrase - HIV DNA sequencing has a good positive predictive value (excluding APOBEC mutations) and an imperfect negative predictive value.	Strong consensus	Strong consensus	STRONG CONSENSUS
3	In the context of a patient who has achieved virological success, in the event of a decision to reduce or simplify treatment to some long-acting regimens, prior genotyping on HIV DNA is useful for the clinical decision in the absence of usable or sufficiently informative RNA data.	Strong consensus	Strong consensus	STRONG CONSENSUS
4	In a virologically suppressed patient, in the event of a decision to reduce or simplify to a sequential treatment (4 days out of 7 or 5 days out of 7) without changing any ARV agents, prior genotyping on HIV DNA is not essential for clinical decision-making, even in the absence of usable or sufficiently informative RNA data.	Strong consensus	Good consensus	STRONG CONSENSUS
5	In the absence of usable or sufficiently informative RNA data in a virologically suppressed patient, in the event of a decision to reduce/simplify to a treatment that reduces the number of ARV, prior genotyping on HIV DNA may be useful for clinical decision-making.	Strong consensus	Strong consensus	STRONG CONSENSUS
Techniques for performing HIV DNA genotyping				
6	In current practice, when the amount of DNA is very low, the performance (sensitivity, representativeness of viral populations) of HIV DNA genotyping is reduced.	Strong consensus	N.A.	STRONG CONSENSUS
7	In current practice, genotyping on HIV DNA can be performed either from whole blood, from the cells isolated from peripheral blood (PBMC) or blood cell pellets.	Strong consensus	N.A.	STRONG CONSENSUS
8	Duplicate DNA genotyping increases performance (sensitivity, representativeness of viral populations) but is not possible in current clinical practice.	Strong consensus	N.A.	STRONG CONSENSUS
9	Sanger and ultra-deep sequencing can be used to perform HIV DNA genotyping.	Strong consensus	N.A.	STRONG CONSENSUS
10	For ultra-high throughput DNA sequencing techniques (ultra-deep sequencing), with the current state of knowledge, the clinically relevant detection threshold is not defined. Nevertheless, it may be interesting to report any viral minority population detected for multidisciplinary discussions.	No Consensus	N.A.	NO CONSENSUS
Consideration of APOBEC mutations				
11	Detection of the <i>M184I</i> mutation in HIV DNA is suggestive of the presence of a defective genome due to the APOBEC enzyme when it is associated with other suggestive mutations (e.g., <i>M41I</i> , <i>M230I</i> on reverse transcriptase) and/or stop codons.	Strong consensus	Strong consensus	STRONG CONSENSUS
12	When resistance mutations attributable to APOBEC are detected, their significance should be interpreted with caution and based on the clinical context and treatment history.	Strong consensus	Strong consensus	STRONG CONSENSUS
13	The presence of resistance mutations attributable to APOBEC should be reported in the HIV DNA genotyping analysis.	Strong consensus	N.A.	STRONG CONSENSUS
Reporting of genotyping results				
14	The clinical interpretation of resistance mutations on HIV DNA genotyping should be discussed at multidisciplinary consultation meetings.	Strong consensus	Strong consensus	STRONG CONSENSUS
15	In a virologically suppressed patient, the detection on HIV DNA genotyping of new resistance mutations (excluding APOBEC & stop codons) previously undetected must be	Strong consensus	Strong consensus	Strong consensus

	considered for the treatment switch decision and subsequent follow-up.			
ARV recycling				
16	In a patient with an undetectable viral load for at least one year, in the presence of a documented M184V substitution over the past five years, the use of a 2 nd generation InSTI + XTC + 1 NRTI combination presents low risk of virological failure over time.	Strong consensus	Strong consensus	STRONG CONSENSUS
17	In a patient with an undetectable viral load for at least one year, in the presence of a documented M184V substitution over the past five years, the use of a 2 nd generation InSTI + XTC combination might present a risk of virological failure over time.	Strong consensus	No consensus	NO CONSENSUS
18	The use / recycling of NNRTIs in the event of documented resistance to ARV agents of this class is associated with a greater risk of virological failure, independently of the duration of undetectable viral load, especially in drug-reduction strategies using ARV agents of this class.	Strong consensus	Strong consensus	STRONG CONSENSUS
Test availability and delays				
19	HIV DNA genotypic tests should be accessible in clinical practice to all clinicians managing PLWH.	Strong consensus	Strong consensus	STRONG CONSENSUS
20	Reports of genotypic HIV DNA test results should be given to clinicians within 30 days.	Strong consensus	Strong consensus	STRONG CONSENSUS

N.A.: Not available; ARV: Antiretroviral; APOBEC: Apolipoproteins B mRNA editing enzyme; InSTI: Integrase Strand Transfer Inhibitor; XTC: Lamivudine/Emtricitabine; PLWH: People living with HIV.

Table S2 – Statements, detailed virologists’ and clinicians’ voting results, and cumulative results for both groups

For each statement, a total number of voters different from 21 for virologists and 40 or 47 for clinicians indicates the use of the ‘I don't know’ option by voters during the second voting round.

STATEMENTS		VOTING SCORES FROM VIROLOGISTS					VOTING SCORES FROM CLINICIANS					CUMULATIVE SCORES
		Scores 1-2-3 (n)	Scores 4-5-6 (n)	Scores 7-8-9 (n)	Median	Results	Scores 1-2-3 (n)	Scores 4-5-6 (n)	Scores 7-8-9 (n)	Median	Results	
Clinical situations for the use of HIV DNA genotyping												
1	In the context of a therapeutic decision requiring genotyping data, when HIV RNA is not amplifiable, when cumulative HIV RNA genotyping is not available and/or in the event of an incomplete or unusable genotypic history, HIV DNA genotyping is recommended.	4.8% (1)	0% (0)	95.2% (20)	9	Strong consensus	2.1% (1)	10.6% (5)	87.2% (41)	8	Strong consensus	STRONG CONSENSUS
2	For the following therapeutic targets - reverse transcriptase, protease, integrase - HIV DNA sequencing has a good positive predictive value (excluding APOBEC mutations) and an imperfect negative predictive value.	4.8% (1)	4.8% (1)	90.5% (19)	9	Strong consensus	2.1% (1)	17.0% (8)	80.9% (38)	8	Strong consensus	STRONG CONSENSUS
3	In the context of a patient with virological success, in the event of a decision to reduce or simplify treatment to some long-acting regimens, prior genotyping on HIV DNA is useful for clinical decision-making in the absence of usable or sufficiently informative RNA data.	9.5% (2)	4.8% (1)	85.7% (18)	9	Strong consensus	6.4% (3)	14.9% (7)	78.7% (37)	8	Strong consensus	STRONG CONSENSUS
4	In a virologically suppressed patient, in the event of a decision to reduce or simplify to a sequential treatment (4 days out of 7 or 5 days out of 7) without changing any ARV agents, prior genotyping on HIV DNA is not essential for the clinical decision, even in the absence of usable or sufficiently informative RNA data.	10% (2)	5% (1)	85.0% (17)	8	Strong consensus	15% (6)	12.5% (5)	72.5% (29)	8	Good consensus	STRONG CONSENSUS
5	In the absence of usable or sufficiently informative RNA data in a virologically suppressed patient, in the event of a desire to reduce/simplify towards a regimen that reduces the number of ARV agents prior genotyping on HIV DNA may be useful for clinical decision-making.	9.5% (2)	4.8% (1)	85.7% (18)	8	Strong consensus	7.5% (3)	17.5% (7)	75.0% (30)	8	Strong consensus	STRONG CONSENSUS
Techniques for performing HIV DNA genotyping												
6	In current practice, when the amount of DNA is very low, the performance (sensitivity, representativeness of viral populations) of HIV DNA genotyping is reduced.	5.3% (1)	5.3% (1)	89.5% (17)	9	Strong consensus	N.A.	N.A.	N.A.	N.A.	N.A.	STRONG CONSENSUS
7	In current practice, genotyping on HIV DNA can be performed either from whole blood, from the cells isolated from peripheral blood (PBMIC) or from blood	14.3% (3)	9.5% (2)	76.2% (16)	8	Strong consensus	N.A.	N.A.	N.A.	N.A.	N.A.	STRONG CONSENSUS

	cell pellets.											
8	Duplicate DNA genotyping increases performance (sensitivity, representativeness of viral populations) but is not possible in current clinical practice.	10.5% (2)	5.3% (1)	84.2% (16)	8	Strong consensus	N.A.	N.A.	N.A.	N.A.	N.A.	STRONG CONSENSUS
9	Sanger and ultra-deep sequencing can be used to perform HIV DNA genotyping.	0% (0)	23.8% (5)	76.2% (16)	8	Strong consensus	N.A.	N.A.	N.A.	N.A.	N.A.	STRONG CONSENSUS
10	For ultra-high throughput DNA sequencing techniques (ultra-deep sequencing), with the current state of knowledge, the clinically relevant detection threshold is not defined. Nevertheless, it may be interesting to report to report any viral minority population detected for multidisciplinary discussions.	10% (2)	25% (5)	65% (13)	7.5	No Consensus	N.A.	N.A.	N.A.	N.A.	N.A.	
Consideration of APOBEC mutations												
11	Detection of the <i>M184I</i> mutation in HIV DNA is suggestive of the presence of a defective genome due to the APOBEC enzyme when it is associated with other suggestive mutations (e.g., <i>M41I</i> , <i>M230I</i> on reverse transcriptase) and/or stop codons.	0% (0)	9.5% (2)	90.5% (19)	8	Strong consensus	3.1% (1)	15.6% (5)	81.3% (26)	8.5	Strong consensus	STRONG CONSENSUS
12	When resistance mutations attributable to APOBEC are detected, their significance should be interpreted with caution and based on the clinical context and treatment history.	4.8% (1)	0% (0)	95.2% (20)	9	Strong consensus	5.4% (2)	5.4% (2)	89.2% (33)	9	Strong consensus	STRONG CONSENSUS
13	The presence of resistance mutations attributable to APOBEC should be reported in the HIV DNA genotyping analysis.	9.5% (2)	4.8% (1)	85.7% (18)	8	Strong consensus	N.A.	N.A.	N.A.	N.A.	N.A.	STRONG CONSENSUS
Reporting of genotyping results												
14	The clinical interpretation of resistance mutations on HIV DNA genotyping should be discussed at multidisciplinary consultation meetings.	4.8% (1)	19% (4)	76.2% (16)	9	Strong consensus	4.8% (1)	19% (4)	76.2% (16)	9	Strong consensus	STRONG CONSENSUS
15	In a virologically suppressed patient, the detection on HIV DNA genotyping of new resistance mutations (excluding APOBEC & stop codons) previously undetected must be considered for a treatment switch decision and subsequent follow-up.	4.8% (1)	14.3% (3)	81% (17)	8	Strong consensus	4.8% (1)	14.3% (3)	81% (17)	8	Strong consensus	STRONG CONSENSUS
ARV recycling												
16	In a patient with an undetectable viral load for at least one year, in the presence of a documented M184V substitution over the past five years, the use of a 2 nd generation INSTI + XTC + 1 NRTI combination presents low risk of virological failure over time.	14.3% (3)	9.5% (2)	76.2% (16)	8	Strong consensus	4.3% (2)	6.4% (3)	89.4% (42)	8	Strong consensus	STRONG CONSENSUS

17	In a patient with an undetectable viral load for at least one year, in the presence of a documented <i>M184V</i> substitution over the past five years, the use of a 2 nd generation InSTI + XTC combination might present a risk of virological failure over time.	19% (4)	4.8% (1)	76.2% (16)	8	Strong consensus	22.5% (9)	25% (10)	52.5% (21)	7	No consensus	NO CONSENSUS
18	The use / recycling of NNRTIs in the event of documented resistance to ARV agents of this class is associated with a greater risk of virological failure, independently of the duration of undetectable viral load, especially in drug-reduction strategies using ARV agents of this class.	4.8% (1)	19% (4)	76.2% (16)	8	Strong consensus	0% (0)	17.9% (7)	82.1% (32)	8	Strong consensus	STRONG CONSENSUS
Test availability and delays												
19	HIV DNA genotypic tests should be accessible in clinical practice to all clinicians managing PLWH.	4.8% (1)	9.5% (2)	85.7% (18)	9	Strong consensus	10.6% (5)	6.4% (3)	83.0% (39)	9	Strong consensus	STRONG CONSENSUS
20	Reports of genotypic HIV DNA test results should be given to clinicians within 30 days.	4.8% (1)	4.8% (1)	90.5% (19)	9	Strong consensus	10.0% (4)	7.5% (3)	82.5% (33)	9	Strong consensus	STRONG CONSENSUS

N.A.: Not available; ARV: Antiretroviral; APOBEC: Apolipoproteins B mRNA editing enzyme; InSTI: Integrase Strand Transfer Inhibitor; XTC: Lamivudine/Emtricitabine;
PLWH: People living with HIV

Table S3 – APOBEC context drug resistance mutations

ARV class	Major resistance mutations
PI	<i>M46I, G73S, D30N</i>
NRTI	<i>D67N, M184I</i>
NNRTI	<i>M230I, E138K, G190E, G190S</i>
InSTI	<i>G163R, G163K, G140R, D232N, E138K, G140S, G118R, R263K</i>

PI: Protease Inhibitor; InSTI: Integrase Strand Transfer Inhibitor