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## Development of a set of SNP markers for population genetics of the sea rose, *Pentapora fascialis*

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1 **Development of a set of SNP markers for population genetics of the sea rose, *Pentapora***  
2 ***fascialis***

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10

11 **Abstract**

12 A set of single nucleotide polymorphisms (SNP) was developed from the transcriptome of 10  
13 individuals of the sea rose *Pentapora fascialis* (Cheilostomata) collected in Banyuls-sur-Mer  
14 Bay (NW Mediterranean sea). 53042 putative SNPs were identified and mapped on a de novo  
15 assembled transcriptome. A selected set of 320 SNPs with highest coverage and uniquely  
16 mapped in the assembled transcriptome was tested using a MassARRAY System on 95  
17 individuals sampled in natural rocks and artificial reefs distributed over a hundred kilometers  
18 in the NW Mediterranean (outside Banyuls-sur-Mer Bay). A total of 177 SNPs were  
19 successfully genotyped and found to be polymorphic. Among these, 154 SNPs were in  
20 Hardy-Weinberg equilibrium over all samples, with significant linkage disequilibrium in only  
21 21 pairs of SNPs. The newly developed loci will be a valuable tool for population genetics  
22 studies of this calcifying bryozoan species whose erect structure makes it an engineering  
23 species and a target for conservation plans.

24 **Keywords**

25 *Pentapora fascialis*, Single nucleotide polymorphism, MassARRAY, bryozoan

26

## 27 **Introduction**

28 The erect cheilostomata bryozoan *Pentapora fascialis* (Pallas, 1766) is a common sessile  
29 mega-benthic species with medium size ranging from 10 to 20 cm which distributes in the  
30 Mediterranean on hard rock substrates, cobble and boulder areas and other living species  
31 (gorgonians) down to 60 m or more (Zabala 1986, Lombardi et al. 2008). *P. fascialis* has a  
32 dome-like shape made of convoluted bright orange bilamina sheets. Considered as a synonym  
33 species of *P. foliacea* (North Atlantic) by some taxonomists (Hayward and Ryland 1999), *P.*  
34 *fascialis* (Mediterranean) was redescribed as a distinct species based on a phylogenetic  
35 analysis of skeletal morphological differences (Lombardi et al. 2010). The present day  
36 geographical distribution of the *P. foliacea*–*P. fascialis* species complex extends  
37 discontinuously from the Hebrides, English Channel, the Bay of Biscay, along the Portuguese  
38 coast to Morocco, and into the Mediterranean as far east as the Adriatic Sea (Gautier, 1962;  
39 Zabala, 1986; Hayward & Ryland, 1999).

40 This calcifying bio-constructor is an habitat-forming species of temperate and cold-water  
41 oceanic shelves which responds to the ocean climate variations (Wood et al. 2012, Lombardi  
42 et al. 2014, Fortunato et al. 2015, Pagès-Escolà et al. 2018). *P. fascialis* is a modular  
43 organism growing at a rate up to 3.5 cm y<sup>-1</sup> in height through asexual budding process and  
44 estimates of growth rate suggest that *P. fascialis* colonies in the Mediterranean can live up to  
45 ten years old or more (Cocito et al. 1998a). Little is known about its reproduction, except the  
46 presence of ovicells, a reliable indicator of the sexual reproductive status. Colonies of *P.*  
47 *fascialis* as small as 2.8 cm have been recorded as having ovicells, suggesting sexual maturity  
48 occurs at two to three year old (Cocito *et al.*, 1998b). By analogy with the other  
49 cheilostomata bryozoan species, *Bugula neritina*, it is assumed to reproduce sexually every  
50 year with a pelagic lecithotrophic larva dispersing during less than one day (Keough &

51 Chernoff, 1987). Despite larval dispersal traits are actually unknown, species presence on  
52 artificial reefs deployed over soft-bottom substrates (Blouet et al. 2022) and high recruitment  
53 rates found on artificial substrates such as plastic nets (Pagès-Escolà et al. 2020) indicate  
54 efficient larval dispersal over a few kms at least. This process is essential in the resilience of a  
55 fragile species vulnerable to physical disturbances such as storms and scuba-diving activity  
56 (Cocito et al. 1998b, Sala et al. 1996).

57 The objective of the present study is to propose a set of Single Nucleotide Polymorphism  
58 markers that will be useful to confirm the morphological species differentiation between *P.*  
59 *fascialis* and *P. foliacea* and study *P. fascialis* population genetics and connectivity.

60

## 61 **Material and Methods**

### 62 *Transcriptome sequencing*

63 The transcriptome of *P. fascialis* was sequenced from the total RNA of ten individuals  
64 collected by SCUBA diving in sites a few kms apart between 10 and 20 meters depth in  
65 Banyuls sur Mer, France. While a short sampling distance from the laboratory ensured the  
66 best preservation conditions for RNA extraction, it may be a limit for finding Single  
67 Nucleotide Polymorphism useful for future connectivity studies. However, with an assumed  
68 limited dispersal potential, population genetics differentiation was expected at the scale of a  
69 few kms. Taking samples from the Mediterranean sea only ensured SNPs determination will  
70 apply specifically to *P. fascialis*. Small fragments were immediately frozen in liquid nitrogen,  
71 and preserved at -80 °C. Total RNA extractions were performed with Maxwell 16  
72 LEVsimplyRNA purification kits. RNA concentrations and quality were analyzed with an  
73 Agilent 2100 Bioanalyzer, and sequencing was done using paired-end in one lane of Illumina  
74 HiSeq 3000. RNA-seq libraries were prepared according to Illumina's protocols using the  
75 Illumina TruSeq Stranded mRNA sample prep kit to analyze mRNA. Briefly, mRNA was

76 selected using poly-T beads. Then, RNA were fragmented to generate double stranded cDNA  
77 and adaptators were ligated to be sequenced. 11 cycles of PCR were applied to amplify  
78 libraries. Library quality was assessed using a Fragment Marker development and screening.  
79 Analyser and libraries were quantified by qPCR using the Kapa Library Quantification Kit.  
80 RNA-seq experiments have been performed on an Illumina HiSeq3000 using a paired-end  
81 read length of  $2 \times 150$  pb with the Illumina HiSeq3000 sequencing kits.

82

### 83 *Marker development and screening*

84 Sequence reads quality was assessed using FastQC v0.10.1 (Andrews 2010). Trimmomatic  
85 v0.32 was used to remove low quality reads with a Phred score below 20, as well as the  
86 Illumina adapters (Bolger et al. 2014). FastQC was performed again to verify the integrity of  
87 the remaining raw Illumina sequence reads. High-quality reads were then used for the de  
88 novo transcriptome assembly, using Trinity with its default k-mer value of 25 (Grabherr et al.  
89 2011). DiscoSnp + + v2.2.10 (Uricaru et al. 2015) was used with k-mer size of 61 and  
90 minimum coverage of 20 per read set to detect SNPs from the 10 different individual  
91 transcriptomes. Among the 53042 SNPs detected, a total of 21481 SNPs had a good coverage  
92 ( $> 100$  reads) in all 10 individual transcriptomes and 44575 SNPs could be mapped on the  
93 transcriptome assembly, either in a unique or in multiple position. The 380 SNPs with higher  
94 coverage among the SNPs mapped at a unique position were selected and tested as candidate  
95 markers for the MassArray genotyping. Using Assay Designer version 4.0.0.2 (Agena  
96 Biosciences), with a mass range of 4000-9000Da, an amplicon length range of 70-130bp, an  
97 extension primer length range of 11-30 bases and a minimum analyte peak separation of 20  
98 Da, four cost-effective multiplexes of 40 markers were developed for a total of 320 SNPs.  
99 The assay was performed on the genomic DNA extracted from 95 samples of *P. fascialis*  
100 collected in the NW Mediterranean (Table 1), using the DNeasy Blood and Tissue kits from

101 QIAGEN. Allele calling was carried out with Typer Viewer v.4.0.24.71 (Agena Biosciences).

102

Location	Site	Latitude (decimal degrees)	Longitude (decimal degrees)	Depth (m)	N
Cap d'Agde - natural rocks	A3	43.252095	3.524767	30	11
	A4	43.244578	3.519568	25	24
Cap d'Agde – artificial reef	AGD26	43.2386	3.4992	20	20
Barcarès – artificial reef	BAR51	42.8241	3.0578	16	10
Leucate – artificial reef	LEU47	42.8955	3.0701	16	10
Canet – artificial reef	CAN53	42.7216	3.0722	28	10
Saint Cyprien – artificial reef	CYP 54	42.6293	3.0720	30	10

103

104 **Table 1.-** Location and number of individuals (N) of *Pentapora fascialis* used for SNPs  
105 validation using MassArray

106

107 Monomorphic SNPs, loci with weak or ambiguous signal (i.e., displaying more than three  
108 clusters of genotypes or unclear cluster delimitation) and loci with too much missing data  
109 were all discarded. Genetics parameters, observed heterozygosity and expected  
110 heterozygosity were calculated using Genepop 4.7.5 (Rousset 2008). Hardy–Weinberg  
111 equilibrium (HWE) for each locus and Linkage Disequilibrium (LD) between each pair of  
112 loci (with Bonferroni correction) was tested using Genepop 4.7.5 (Rousset 2008).

113

## 114 **Results**

115 Across the 320 SNPs tested, 143 SNPs either did not amplify or were monomorphic,  
116 probably resulting from the limited spatial extent of the ten individuals used for searching  
117 putative SNPs (Supplementary Material Table S1). The average call rate of the 177 SNPs  
118 detected in more than half of the 95 samples and polymorphic was 97.4 %. The observed  
119 heterozygosity ranged from 0 to 1, while the expected heterozygosity varied from 0.0161 to

120 1, with 32 SNPs displaying an heterozygosity lower than 0.1 (highlighted in orange in the  
121 Supplementary Material Table S2). Among the 177 SNPs, 23 departed significantly from  
122 HWE proportions over the 95 samples spanning the 7 locations, ( $p$ -value < 0.05, highlighted  
123 in red in Supplementary Material Table S2). LD was rare, present in only 21 pairs of loci out  
124 of the 51040 pairs tested (the 32 loci involved are highlighted in yellow in Supplementary  
125 Material Table S2). These results provide a valuable resource for future population genetic  
126 studies of this endemic bryozoan species in the Mediterranean Sea.

127

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140 Author contributions MP and KG designed the study. MM, AD, and EG, performed the  
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142 reviewed the manuscript.

143

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147 2017 7268 / 2018 0697, PI K. Guizien, AAP 2016).

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149 Data Availability The RNA seq data will be released publicly through National Center for  
150 Biotechnology Information portal in March 2024 (<https://submit.ncbi.nlm.nih.gov>).

151

## 152 **References**

153 Andrews S (2010) FastQC: a quality control tool for high throughput sequence data. [http://](http://www.bioinformatics.babraham.ac.uk/projects/fastqc)  
154 [www.bioinformatics.babraham.ac.uk/projects/fastqc](http://www.bioinformatics.babraham.ac.uk/projects/fastqc)

155 Blouet S, Bramanti L, Guizien K (2022) Artificial reefs geographical location matters more  
156 than shape, age and depth for sessile invertebrate colonization in the Gulf of Lion  
157 (NorthWestern Mediterranean Sea). Peer Community Journal 2(article no e24).  
158 <https://doi.org/10.24072/pcjournal.107>.

159 Bolger AM, Lohse M, Usadel B (2014) Trimmomatic: a flexible trimmer for Illumina  
160 Sequence Data. *Bioinformatics* 30(15):2114–2120. [https://doi.org/10.1093/](https://doi.org/10.1093/bioinformatics/btu170)  
161 [bioinformatics/btu170](https://doi.org/10.1093/bioinformatics/btu170)

162 Cocito S, Sgorbini S, Bianchi C (1998a) Aspects of the biology of the bryozoan *Pentapora*  
163 *fascialis* in the northwestern Mediterranean. *Marine Biology* **131**, 73–82  
164 <https://doi.org/10.1007/s002270050298>

165 Cocito S., Ferdeghini F., & Sgorbini S. (1998b) *Pentapora fascialis* (Pallas) [Cheilostomata:  
166 Ascophora] colonization of one sublittoral rocky site after sea-storm in the northwest  
167 Mediterranean. *Hydrobiologia*, **375/376**, 59-66.

168 Eisenbarth S., Gehling M., Harder A., Steffan B. (2002) Pentaporins A, B and C: disulfides



169 from the marine bryozoan *Pentapora fascialis*, *Tetrahedron*, 58 (42): 8461-8464, ISSN 0040-  
170 4020, [https://doi.org/10.1016/S0040-4020\(02\)01050-5](https://doi.org/10.1016/S0040-4020(02)01050-5).

171 Fortunato H (2015) Bryozoans in climate and ocean acidification research: A reappraisal of  
172 an under-used tool, *Regional Studies in Marine Science* 2: 32-44, ISSN 2352-4855,  
173 <https://doi.org/10.1016/j.rsma.2015.08.010>.

174 Gautier YV (1962) Recherches écologiques sur les Bryozoaires cheilostomes en  
175 Méditerranée occidentale. Thèse de l'Université d'Aix-Marseille.

176 Grabherr M, Haas B, Yassour M, Levin J, Thompson D, Amit I, Adiconis X, Fan L,  
177 Raychowdhury R, Zeng Q, Chen Z, Mauceli E, Hacohen N, Gnirke A, Rhind N, di Palma F,  
178 Birren BW, Nusbaum C, Lindblad-Toh K, Friedman N, Regev A (2011) Full-length  
179 transcriptome assembly from RNA-seq data without a reference genome. *Nat Biotechnol*  
180 2011 May 15(7):644–652. doi:<https://doi.org/10.1038/nbt.1883>

181 Hayward PJ, Ryland JS (1999) Synopses of the British Fauna (new series): Cheilostomatous  
182 Bryozoa Part II: Hippothoidea-Celleporoidea. London: The Linnean Society of London and  
183 Estuarine and Coastal Sciences Association.

184 Keough MJ, Chernoff H (1987) Dispersal and population variation in the bryozoan *Bugula*  
185 *neritina*. *Ecology*, **68**, 199 - 210.

186 Lombardi C., Cocito S., Occhipinti-Ambrogi A., & Porter J. (2008). Distribution and  
187 morphological variation of colonies of the bryozoan *Pentapora fascialis* (Bryozoa:  
188 Cheilostomata) along the western coast of Italy. *Journal of the Marine Biological Association*  
189 *of the United Kingdom*, 88(4), 711-717. doi:10.1017/S0025315408001525

190 Lombardi C, Taylor PD, Cocito S (2010) Systematics of the Miocene–Recent bryozoan genus

191 Pentapora (Cheilostomata). Zoological Journal of the Linnean Society, 2010, 160, 17–39.

192 Lombardi C, Taylor PD, Cocito S (2014) Bryozoan constructions in a changing  
193 Mediterranean Sea. In: Godofredo, S., Dubinsky, (Eds.), The Mediterranean Sea: Its History  
194 and Present Challenges. Springer, Dordrecht, pp. 373–384.

195 Pagès-Escolà M, Hereu B, Garrabou J, Montero-Serra I, Gori A, Gómez-Gras D, Figuerola B,  
196 Linares C (2018) Divergent responses to warming of two common co-occurring  
197 Mediterranean bryozoans. Scientific Reports, 8, 17455. [https://doi.org/10.1038/s41598-018-](https://doi.org/10.1038/s41598-018-36094-9)  
198 36094-9

199 Pagès-Escolà M, Linares C, Gómez-Gras D, Medrano A, Hereu B. (2020) Assessing the  
200 effectiveness of restoration actions for Bryozoans: The case of the Mediterranean Pentapora  
201 fascialis. Aquatic Conserv: Mar Freshw Ecosyst. 30:8–19. <https://doi.org/10.1002/aqc.3236>

202 Rousset, F., 2008. Genepop'007: a complete reimplementation of the Genepop software for  
203 Windows and Linux. Mol. Ecol. Resources 8: 103-106.

204 Sala E, Garrabou J, Zabala M (1996) Effects of diver frequentation on Mediterranean  
205 sublittoral populations of the bryozoan *Pentapora fascialis* . *Marine Biology* **126**, 451–459.  
206 <https://doi.org/10.1007/BF00354627>

207 Uricaru R, Rizk G, Lacroix V, Quillery E, Plantard O, Chikhi R, Lemaitre C, Peterlongo P  
208 (2014) Reference-free detection of isolated SNPs. Nucleic Acids Res. doi:[https://doi.org/10.](https://doi.org/10.1093/nar/gku1187)  
209 1093/nar/gku1187

210 Wood, A.C.L., Probert, P.K., Rowden, A.A., Smith, A.M., 2012. Complex habitat generated  
211 by marine bryozoans: a review of its distribution, structure, diversity, threats and  
212 conservation. Aquat. Conserv. 22, 547–563. <http://dx.doi.org/10.1002/aqc.2236>.

213 Zabala, M., 1986. In: Fauna dels briozous dels Països Catalans. Institut d'Estudis Catalans,  
214 pp. 833.