

Rapid tests should be used with caution for HIV-1 primary infection screening

Vincent Guiraud, Quentin Beaulieu, Antoine Fauchois, Pascale Jean-Charles, Marie-Capucine Costes, Bruno Le Labousse, Pr Agnès Gautheret-Dejean

▶ To cite this version:

Vincent Guiraud, Quentin Beaulieu, Antoine Fauchois, Pascale Jean-Charles, Marie-Capucine Costes, et al.. Rapid tests should be used with caution for HIV-1 primary infection screening. Medical Microbiology and Immunology, In press, 213 (1), pp.10. 10.1007/s00430-024-00792-1. hal-04621131

HAL Id: hal-04621131 https://hal.sorbonne-universite.fr/hal-04621131

Submitted on 23 Jun 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Rapid tests should be used with caution for HIV-1 primary infection screening 1 2 Vincent Guiraud^{a,&}, Quentin Beaulieu^{a,&}, Antoine Fauchois^a, Pascale Jean-Charles^a, Marie-3 Capucine Costes^a, Bruno Le Labousse^a, Agnès Gautheret-Dejean^{a,b*} 4 ^a AP-HP, Hôpitaux Universitaires La Pitié Salpêtrière-Charles Foix, Service de Virologie, F-5 6 75013 Paris, France ^b Université Paris cité, INSERM UMR-S 1139 Physiopathologie et pharmacotoxicologie 7 8 placentaire humaine : microbiote pré & post-natal, F-75006 Paris, France 9 10 &These authors also contributed equally to this work. 11 12 * Corresponding author: agnes.gautheret@aphp.fr 13 Corresponding author: 14 Pr Agnès Gautheret-Dejean 15 Hôpital Universitaire La Pitié Salpêtrière, Service de Virologie 16 83 Bd de l'Hôpital, 75013 Paris, France 17 Tel: +33 1 42177401 / Fax: +33 1 42177411 18 E-mail: agnes.gautheret@aphp.fr 19 20 21 Keywords: rapid test, HIV-1, primary infection, seroconversion, screening, sensitivity 22 23

ABSTRACT:

24

25 Rapid tests allow outpatient, low cost, reliable, screening for chronic HIV infection. However, data regarding their sensitivity on primary infection remain scarce. The objective of this study 26 was to assess sensitivity of nine HIV rapid tests for primary HIV-1 infection screening. 27 Seventy-five serum samples from patients during HIV-1 primary infection were included. 28 Primary infection was diagnosed by a positive 4th generation ELISA and HIV-1 RNA 29 positivity confirmed by Western blot patterns associated with HIV-1 primary infection. Early 30 31 seroconversion was defined as the absence of antibodies on HIV-1 Western blot associated 32 with HIV-1 RNA and p24-antigen positivity. An identical sensitivity (95% CI) of 76.7% (65.2-84.2%) was observed for HIV 1/2 STAT-PAK® Assay (STAT-PAK), INSTITM HIV-33 1/HIV-2 antibody Test (INSTI), SURE CHECK® HIV 1/2 (SURE CHECK) and 34 MULTISURE HIV rapid test (MULTISURE) with visual reading. Sensitivity was 74.7% 35 36 (63.8-83.1%) for MULTISURE (automatic result), 77.0% (66.3-85.1%) for FIRST RESPONSE® Test VIH 1-2.O CARTE (FIRST RESPONSE), 83.8% (73.8-90.5%) for VIKIA 37 HIV1/2[®] (VIKIA), 88.0% (78.7-93.6%) for GenieTM Fast HIV 1/2 (Genie Fast), 88.6% (79.0-38 94.1%) for Hexagon HIV (Hexagon), and 92.8% (83.6-96.3%) for Exacto® TEST HIV Pro 39 40 (Exacto). However, rapid tests performed poorly for the early seroconversion subgroup (n=14), with sensitivities ranging from 7% (1.3-31.5) for STAT-PAK, INSTI, SURE 41 42 CHECK, MULTISURE (automatic reading), to 29% (12-55%) for FIRST RESPONSE, 31% 43 (13-58%) for VIKIA, 43% (21-67%) for Hexagon and 57.1% (32.6-78.6%) for Exacto and Genie Fast. Overall, despite significant discrepancies in sensitivity, HIV rapid tests should be 44 45 used with caution in the context of a suspected primary infection.

46

47

48

Introduction

49

50 In 2021, more than 38 million people where living with HIV worldwilde [1]. With the aim 51 to end the AIDS epidemic as a public health threat, UNAIDS defined new global 95-95-95 52 targets for 2025 which corresponds to 95% of people living with HIV knowing their status, 95% of them receiving antiretroviral treatment, and among them 95% with viral suppression 53 54 [2]. To achieve these objectives, diagnosis is essential. Among barriers to HIV screening are 55 costs, need to present to a medical setting, fear of stigma or delays in test result receipt [3–6]. 56 Rapid tests address most of those points and are reliable for chronic HIV infection diagnosis 57 [7–10]. However, their sensitivity to detect HIV-1 primary infection exhibits discrepant 58 results [10–14]. This point remains a major concern given that patients are around 26 times 59 more contaminant at this stage of the infection, originating a significant part of HIV 60 contaminations [15–19]. However, to date only a small proportion of patients is diagnosed at an early stage [20]. 61 62 The objective of this study was to determine the sensitivity of nine HIV rapid tests for

6364

65

67

68

69

70

71

72

73

Methods

66 Sample collection

HIV-1 primary infection screening.

Seventy-five samples (74 serums and one plasma) from consecutive patients at the stage of HIV-1 primary infection were included at Pitié Salpêtrière Hospital (Paris, France). HIV diagnosis was performed in accordance with French legislation [21], following the routine algorithm. HIV screening performed using the laboratory-based enzyme immunoassay Liaison XL Murex HIV Ag/Ab (DiaSorin, Antony, France) and a confirmatory assay was performed using the New Lav Blot I (Bio-Rad laboratories, Marnes-la-Coquette, France) Western blot. Diagnosis of primary HIV infection was based on Western blot pattern

according to the WHO guidelines [22], associated with a positive HIV-1 viral load, using
Cobas® AmpliPrep/Cobas TaqMan® HIV-1 Test, v2.0 (Roche Diagnostics, Manheim
Germany). Early seroconversion was defined as the absence of antibodies on the HIV-1
Western blot associated with both HIV-1 RNA (assessed on Cobas® AmpliPrep/Cobas
TaqMan® HIV-1 Test, v2.0) and p24 antigen positivity, assessed on VIDAS HIV p24 II

79 (bioMerieux, France). Samples were stored at -20°C before use.

Sample processing

Samples were prospectively analyzed within the same freezing defrosting cycle with all the nine HIV rapid tests: INSTITM HIV-1/HIV-2 Antibody Test (INSTI) (Biolytical, Canada), MULTISURE HIV Rapid Test (MULTISURE) (MP Diagnostics Asia Pacific Pte. CE Ltd), SURE CHECK® HIV1/2 (SURE CHECK) (Chembio Diagnostics, Medford, USA), HIV 1/2 STAT-PAK® Assay (STAT-PAK) (Chembio Diagnostics, Medford, USA), Exacto® TEST HIV PRO (Exacto) (Biosynex, Illkirch-Graffenstaden, France), GenieTM Fast HIV 1/2 (Genie) (Bio-Rad laboratories, Marnes-la Coquette, France), VIKIA® HIV 1/2 (VIKIA) (bioMérieux, Marcy l'Etoile, France), FIRST RESPONSE® Test VIH 1-2.0 CARTE (FIRST RESPONSE) (Premier Medical Corporation, Sarigam, India) and Hexagon HIV (Hexagon) (Human, Wiesbaden, Germany). For MULTISURE, visual and automation readings were performed. For Hexagon, FIRST RESPONSE and VIKIA, 71, 74 and 74 of the 75 samples were tested, respectively, due to an insufficient sample quantity. Assays are described Table 1. Each assay was performed and interpreted according to the manufacturers' recommendations by two independent operators. Samples with invalid results according to the manufacturers'

Statistical analysis

recommendations were controlled once.

Statistical analyses were conducted using R version 4.2.1 [23]. Test comparisons were done using Chi-squared test. Statistical tests were two sided with a significance assigned at a p value < 0.05. Confidence intervals were calculated using Wilson's score bound [24]. Finally, we determined that with an assumed 90% sensitivity, a sample size of 75 samples would be enough to estimate our results with around 5% accuracy [25].

Ethics

This study complies with Good Clinical Practices and ethical principles of the Helsinki declaration. All data were anonymized before analysis. Patients were systematically notified of any supplementary biological analyses on frozen samples, initially collected as part of routine clinical practice.

Results

Sensitivity varied widely for the 75 samples tested (Table 2). A sensitivity (95% CI) of 76.7% (65.2-84.2%) was observed for STAT-PAK, INSTI, SURE CHECK and MULTISURE (visual reading). Sensitivity was 74.7% (63.8-83.1%) for MULTISURE (automatic result), 77.0% (66.3-85.1%) for FIRST RESPONSE, 83.8% (73.8-90.5%) for VIKIA, 88.0% (78.7-93.6%) Genie Fast, 88.6% (79.0-94.1%) for Hexagon, and 92.8% (83.6-96.3%) for Exacto (Table 2). As a consequence, using Chi-squared tests, Exacto and Hexagon performed statistically better than STAT-PAK, INSTI, SURE CHECK and MULTISURE (both readings), while Genie Fast was better than MULTISURE automatic reading only (Table 3). We performed an exploratory analysis on the 14 samples from an early seroconversion stage, defined as the absence of antibody on HIV-1 Western blot associated with HIV-1 RNA and p24 antigen positivity (stage 2 and 3 Fiebig) [26]. All HIV rapid tests performed poorly 123 (Table 2), with sensitivities ranging from 7% (1-31%) for MULTISURE (automatic reading),

INSTI, STAT-PAK and SURE-CHECK to 57% (33-79%) for Exacto and Genie Fast.

Of note, three samples had a negative HIV-antibody signal (S/CO <1) with Liaison XL. The first, with a 0.515 S/CO signal was non-reactive for all rapid tests, the second, with a 0.661 S/CO signal had a trace for Exacto and Multisure. The last, with a 0.973 S/CO value had a trace for Exacto and Genie. These few results might imply that lack of sensitivity during early seroconversion stage could result directly from a lack of HIV antibody, although much more samples would be needed to address this point. As expected, Liaison XL p24 S/CO signal was

Finally, one sample tested invalid with Genie was negative upon retesting, and one invalid

Discussion

highly positive for the three samples.

sample with VIKIA returned positive upon retesting.

This study highlighted significant sensitivity discrepancies between 9 rapid tests to screen for HIV-1 primary infection, ranging from 74.7% for the MULTISURE (automatic reading) to 92.8% for Exacto. This study also highlighted a significant concern regarding the early seroconversion subgroup, since sensitivity was at most 57% only. Although these results applied to a much lower number of samples due to the very stringent criteria applied, they raised concerns regarding rapid tests diagnostic windows. Consequently, the use of rapid tests might be discouraged in a context of a suspected HIV-1 primary infection, and an ELISA test should be favored instead.

To the best of our knowledge, sensitivities of MULTISURE, Exacto, SURE CHECK, FIRST RESPONSE, Genie Fast and STAT-PAK were only studied on chronically HIV-infected people [3,9,10,14,27–31]. A previous study assessed INSTI sensitivity during HIV-1

primary infection, with values within the ranges observed in this study: 69% (54-81%) [32].

VIKIA sensitivity during HIV-1 primary infection was estimated as 75% (35%–97%) and 91% (73-98%) on 7 serum and 23 plasma samples, respectively [33,34], and Hexagon 90% (60-98%) on a panel of 10 serum samples of seroconversion [35].

This study has some limitations. First, the 2 investigators were unblinded. This may have slightly overestimated our results, although sensitivity between automatic and visual results for MULTISURE was roughly the same. Second, as the study was performed on thawed sera, results may show some discrepancies with fresh whole blood that is usually used for outpatients screening. Third, most of the samples were originated from HIV-1 group M subtype B (43%) or CRF02_AG (20%) infected patients, which reflected European epidemiology [36]. As a consequence, these results might be taken with caution depending on local epidemiology [37]. Moreover, our results are based on 75 samples only. However all our samples originated from seroconversion samples, while those from commercial seroconversion panels include a non-negligible number of pre-HIV infection and post-seroconversion samples [32]. Fifth, as HIV-1 primary infection was determined by 4th generation ELISA and HIV-1 Western blot patterns and early seroconversion by both HIV-1 RNA and p24-antigen positivity, we were unable to give precise insights on the time between contamination and HIV screening, and stage I Fiebig (HIV-RNA positivity only) could not be identified.

Conclusions

This study highlighted important discrepancy in sensitivity of rapid tests for HIV-1 primary infection, and a lack of sensitivity for the early seroconversion period. As a consequence, rapid tests should be used with an extreme caution in the context of a suspected primary infection.

173 174 175 **Declaration of Competing Interests** The authors declare that they have no competing financial interests or personal relationships 176 that could have influenced the work reported in this paper. 177 178 179 **Author contributions** 180 Vincent Guiraud acquisition and analyses of data, writing-original draft preparation, 181 submission of the final manuscript, Quentin Beaulieu supervision, samples testing, validation, Antoine Fauchois analyses of data, Pascale Jean-Charles samples testing, Marie-Capucine 182 183 Costes samples testing, Bruno Le Labousse samples testing, Agnès Gautheret-Dejean 184 conceptualization, methodology, validation writing-reviewing and editing. 185 186 **Fundings** 187 This study received no specific funding. 188 189 Acknowledgments 190 The authors thanks ADEBIOPHARM ER28 association for the financial participation which 191 allowed us to present our results in congresses. The authors thank Abbott, bioMérieux, 192 BioSynex, Bio-Rad, Chembio diagnostic system, Servibio/Human, Nephrotek/MP 193 Biomedicals Nephrotek/Premier Medical corporation for providing free kits for this study. 194 Laboratories that gave HIV detection kits had no part on study design, data collection, data 195 analyses, data interpretation, manuscript writing nor submission to publication.

196

197

199

Data Availability Statement

- The data that support the findings are available as supplementary table 1 for HIV rapid tests,
- 201 Liaison XL S/CO values and Western blot results.

202 References

- 203 1. UNAIDS data 2022. In Geneva, Switzerland: World Health Organization; 2022. Available from: 204 https://www.unaids.org/sites/default/files/media asset/data-book-2022 en.pdf
- 205 2. Frescura L, Godfrey-Faussett P, Feizzadeh A. A, El-Sadr W, Syarif O, Ghys PD, et al. Achieving the 206 95 95 95 targets for all: A pathway to ending AIDS. Ambrose Z, editor. PLoS ONE. 2022 Aug 4;17(8):e0272405.
- Kendrick SR, Kroc KA, Withum D, Rydman RJ, Branson BM, Weinstein RA. Outcomes of Offering
 Rapid Point-of-Care HIV Testing in A Sexually Transmitted Disease Clinic: JAIDS Journal of
 Acquired Immune Deficiency Syndromes. 2005 Feb;38(2):142–6.
- 4. Kobrak P, Remien RH, Myers JE, Salcuni P, Edelstein Z, Tsoi B, et al. Motivations and Barriers to Routine HIV Testing Among Men Who Have Sex with Men in New York City. AIDS Behav. 2022 Nov;26(11):3563–75.
- Laprise C, Bolster-Foucault C. Understanding barriers and facilitators to HIV testing in Canada from 2009–2019: A systematic mixed studies review. CCDR. 2021 Mar 4;47(2):105–25.
- the Adolescent Medicine Trials Network for HIV/AIDS Interventions, Gamarel KE, Nelson KM, Stephenson R, Santiago Rivera OJ, Chiaramonte D, et al. Anticipated HIV Stigma and Delays in Regular HIV Testing Behaviors Among Sexually-Active Young Gay, Bisexual, and Other Men Who Have Sex with Men and Transgender Women. AIDS Behav. 2018 Feb;22(2):522–30.
- 7. Fisher DG, Hess KL, Reynolds GL, Alonzo TA, Huckabay LM, Van Otterloo L, et al. Comparisons of New HIV Rapid Test Kit Performance. AIDS Behav. 2019 Feb;23(2):313–7.
- 222 8. O'Connell RJ, Agan BK, Anderson SA, Malia JA, Michael NL. Sensitivity of the Multispot HIV-223 1/HIV-2 Rapid Test Using Samples from Human Immunodeficiency Virus Type 1-Positive 224 Individuals with Various Levels of Exposure to Highly Active Antiretroviral Therapy. J Clin 225 Microbiol. 2006 May;44(5):1831–3.
- 226 9. Tan WS, Chow EPF, Fairley CK, Chen MY, Bradshaw CS, Read TRH. Sensitivity of HIV rapid tests compared with fourth-generation enzyme immunoassays or HIV RNA tests. AIDS. 2016 Jul 31;30(12):1951–60.
- 229 10. Mourez T, Lemée V, Delbos V, Delaugerre C, Alessandri-Gradt E, Etienne M, et al. HIV rapid screening tests and self-tests: Be aware of differences in performance and cautious of vendors. 231 EBioMedicine. 2018 Nov;37:382–91.
- 232 11. Duong YT, Mavengere Y, Patel H, Moore C, Manjengwa J, Sibandze D, et al. Poor Performance of the Determine HIV-1/2 Ag/Ab Combo Fourth-Generation Rapid Test for Detection of Acute

- Infections in a National Household Survey in Swaziland. Caliendo AM, editor. J Clin Microbiol. 2014 Oct;52(10):3743–8.
- 236 12. Faraoni S, Rocchetti A, Gotta F, Ruggiero T, Orofino G, Bonora S, et al. Evaluation of a rapid antigen and antibody combination test in acute HIV infection. Journal of Clinical Virology. 2013 May;57(1):84–7.
- 239 13. Patel P, Bennett B, Sullivan T, Parker MM, Heffelfinger JD, Sullivan PS. Rapid HIV screening: 240 Missed opportunities for HIV diagnosis and prevention. Journal of Clinical Virology. 2012 May;54(1):42–7.
- 242 14. Lewis JM, Macpherson P, Adams ER, Ochodo E, Sands A, Taegtmeyer M. Field accuracy of fourth-generation rapid diagnostic tests for acute HIV-1: a systematic review. AIDS. 2015 Nov 28;29(18):2465–71.
- 245 15. Pilcher CD, Tien HC, Eron, Jr. JJ, Vernazza PL, Leu S, Stewart PW, et al. Brief but Efficient: Acute HIV Infection and the Sexual Transmission of HIV. J INFECT DIS. 2004 May 15;189(10):1785–92.
- 247 16. Yerly S, Vora S, Rizzardi P, Chave JP, Vernazza PL, Flepp M, et al. Acute HIV infection: impact on the spread of HIV and transmission of drug resistance: AIDS. 2001 Nov;15(17):2287–92.
- 249 17. Verhofstede C, Mortier V, Dauwe K, Callens S, Deblonde J, Dessilly G, et al. Exploring HIV-1 250 Transmission Dynamics by Combining Phylogenetic Analysis and Infection Timing. Viruses. 2019 251 Nov 26;11(12):1096.
- 252 18. Kroon EDMB, Phanuphak N, Shattock AJ, Fletcher JLK, Pinyakorn S, Chomchey N, et al. Acute 253 HIV infection detection and immediate treatment estimated to reduce transmission by 89% 254 among men who have sex with men in Bangkok. Journal of the International AIDS Society. 2017;20(1):21708.
- 19. Hollingsworth TD, Anderson RM, Fraser C. HIV-1 Transmission, by Stage of Infection. J INFECT
 DIS. 2008 Sep;198(5):687–93.
- 258 20. Santé Publique France. Bulletin de santé publique VIH-IST. Décembre 2022. 28/11/2022 259 [Internet]. 2022 Dec; Available from: https://www.santepubliquefrance.fr/maladies-et-traumatismes/infections-sexuellement-transmissibles/vih-sida/documents/bulletin-national/bulletin-de-sante-publique-vih-ist.-decembre-2022
- 262 21. Arrêté du 28 mai 2010 fixant les conditions de réalisation du diagnostic biologique de l'infection 263 à virus de l'immunodéficience humaine (VIH 1 et 2) et les conditions de réalisation du test 264 rapide d'orientation diagnostique dans les situations d'urgence [Internet]. 2010 [cited 2023 Oct 265 20]. Available from: https://www.legifrance.gouv.fr/loda/id/JORFTEXT000022320859
- 22. AIDS: proposed WHO criteria for interpreting western blot assays for HIV-1, HIV-2, and HTLV-267 I/HTLV-II. Bull World Health Organ. 1991;69(1):127–9, 131–3.
- 268 23. R Core Team. R: A Language and Environment for Statistical Computing.
- 24. Brown LD, Cai TT, DasGupta A. Interval Estimation for a Binomial Proportion. Statist Sci [Internet]. 2001 May 1 [cited 2023 Jun 14];16(2). Available from: https://projecteuclid.org/journals/statistical-science/volume-16/issue-2/Interval-Estimation-
- 272 for-a-Binomial-Proportion/10.1214/ss/1009213286.full

- 273 25. Motulsky H, Citta M, Citta-Vanthemsche M. Biostatistique: une approche intuitive. 3e éd. Louvain-la-Neuve: De Boeck supérieur; 2019.
- 26. Fiebig EW, Wright DJ, Rawal BD, Garrett PE, Schumacher RT, Peddada L, et al. Dynamics of HIV viremia and antibody seroconversion in plasma donors: implications for diagnosis and staging of primary HIV infection. AIDS. 2003 Sep;17(13):1871–9.
- 27. Boadu R, Darko G, Nortey P, Akweongo P, Sarfo B. Assessing the sensitivity and specificity of First Response HIV-1-2 test kit with whole blood and serum samples: a cross-sectional study. AIDS Res Ther. 2016 Dec;13(1):9.
- 28. Kim HN, Hur M, Kim H, Moon HW, Yun YM, Oh EJ, et al. Evaluation of the MULTISURE HIV Rapid
 Test in a Korean population with low human immunodeficiency virus prevalence. Clinical
 Chemistry and Laboratory Medicine (CCLM). 2019 Jul 26;57(8):e189–91.
- 29. Jaspard M, Le Moal G, Saberan-Roncato M, Plainchamp D, Langlois A, Camps P, et al. Finger-Stick Whole Blood HIV-1/-2 Home-Use Tests Are More Sensitive than Oral Fluid-Based In-Home HIV Tests. Abrams WR, editor. PLoS ONE. 2014 Jun 27;9(6):e101148.
- 287 30. Manak MM, Njoku OS, Shutt A, Malia J, Jagodzinski LL, Milazzo M, et al. Evaluation of Performance of Two Rapid Tests for Detection of HIV-1 and -2 in High- and Low-Prevalence Populations in Nigeria. Tang YW, editor. J Clin Microbiol. 2015 Nov;53(11):3501–6.
- 290 31. Dagnra AY, Dossim S, Salou M, Nyasenu T, Ali-Edje K, Ouro-Médeli A, et al. Evaluation of 9 rapid diagnostic tests for screening HIV infection, in Lomé, Togo. Médecine et Maladies Infectieuses. 292 2014 Dec;44(11–12):525–9.
- 293 32. Adams S, Luo W, Wesolowski L, Cohen SE, Peters PJ, Owen SM, et al. Performance evaluation of the point-of-care INSTITM HIV-1/2 antibody test in early and established HIV infections. Journal of Clinical Virology. 2017 Jun;91:90–4.
- 296 Delaugerre C, Antoni G, Mahjoub N, Pialoux G, Cua E, Pasquet A, et al. Assessment of HIV 297 Screening Tests for Pre-Exposure Prophylaxis Programs. The Journal of Infectious Diseases 298 2017 Jun 27 [cited 2022 Dec 29]; Available from: 299 https://academic.oup.com/jid/article-lookup/doi/10.1093/infdis/jix297
- 300 34. Gomes C, Azevedo-Pereira JM. The performance of the VIKIA® HIV1/2 rapid test—Evaluation of the reliability and sensitivity. Journal of Virological Methods. 2011 May;173(2):353–6.
- 35. Ferreira OC, Ferreira C, Riedel M, Widolin MRV, Barbosa-Júnior A. Evaluation of rapid tests for anti-HIV detection in Brazil. AIDS. 2005 Oct;19(Suppl 4):S70–5.
- 36. Bbosa N, Kaleebu P, Ssemwanga D. HIV subtype diversity worldwide. Current Opinion in HIV and AIDS. 2019 May;14(3):153–60.
- 306 37. Biyang YVS, Parkouda S, Bivigou-Mboumba B, Iroungou BA, Ondeme AM, Bisseye C. Evaluation of HIV-1 rapid diagnostic tests in the context of viral genetic diversity in Libreville (Gabon). Pan Afr Med J. 2022;42:194.

309

Table 1. Detailed characteristics of HIV Rapid tests.

Name of the Kit	Manufacturer /	Ab/Ag used	Immunoassay type /	Technology /	Matrix / volume used (μL)	Self-testing /	
	distributor		Ig class detected	time of reading		Professional	
						purpose	
Exacto® TEST HIV PRO	BioSynex (FR)	None / Gp41 ^c and gp36 ^c	Sandwich d	IC / 10-20 min	S, P, WB1 / 5	Self testing	
Genie TM Fast HIV 1/2	BioRad (US)	None / Gp120a, gp41a, gp36a	Sandwich d	IC / 10-30 min	S, P, WB1 / 80	Professional	
						purpose	
HIV 1/2 STAT-PAK® Assay	Chembio (US)	None / Gp41 ^a and gp36 ^a	Sandwich d	IC / 15 min	S, P, WB1/5	Professional	
						purpose	
NSTI TM HIV-1/HIV-2 Antibody	Biolytical (CA)/	None / Gp41 ^a and gp36 ^a	Indirect immunoassay /	IF / immediate	S, P, WBI / 50	Self-testing	
Tests	Nephrotek (FR)		IgG and IgM				
SURE CHECK® HIV 1/2	Chembio (USA)	None / Gp120a, gp41a, gp36a	Sandwich d	IC / 15-20 min	S, P / 2.5, WBl / 1 drop	Self-testing	
VIKIA® HIV 1/2	bioMérieux® (FR)	None / Gp41 ^b and gp36 ^b	Sandwich d	IC / 30 min	S, P, WB1 / 75	Professional	
						purpose	
FIRST RESPONSE® Test VIH 1-	Premier Medical (India)	None / Gp41 ^a , p24 ^a and	Sandwich d	IC / 15 min	S, P / 10	Professional	
2.O CARTE		gp36 ^a			WB1 / 20	purpose	
Hexagon HIV	Human (De)/	None / Gp41a, p24a and	Sandwich d	IC / 5-20 min	S, P / 10	Professional	
	Servibio (FR)	gp36 ^a			WB1 / 20	purpose	
MULTISURE HIV Rapid Test	MP Biomedicals	None / Gp120a, gp41a, p24a	Indirect immunoassay /	IC / 20-25 min	S, P / 25	Professional	
	(Singapour)/	and gp36 ^a	IgG		WB1 / 20	purpose	
	Nephrotek (FR)						

^a Recombinant antigens.

Ag, antigen; Ab, antibody; IC, immunochromatography; IF, immunofiltration; S, serum; P, plasma; WBl, whole blood; US, United States of America; FR, France; CA, Canada; De, Germany.

^b Synthetic peptides.

^c Not specified

^d All immunoglobulin class are recognized, in particular IgG and IgM

Table 2. Sensitivity rate for HIV-1 primary infection screening.

Assay	Overall No	Overall	No Positive samples/	Sensitivity (95% CI), early seroconversion		
	Positive samples/	Sensitivity	early seroconversion			
	sample tested	(95% CI)	subgroup samples	subgroup		
			tested			
Exacto® TEST HIV PRO	69/75	92.8 (83.6-96.3)	8/14	57 (33-78)		
Genie TM Fast HIV 1/2	66/75	88.0 (78.7-93.6)	8/14	57 (33-78)		
HIV 1/2 STAT-PAK®	57/75	76.7 (65.2-84.2)	1/14	7 (1-31)		
Assay						
INSTI TM HIV-1/HIV-2	57/75	76.7 (65.2-84.2)	1/14	7 (1-31)		
Antibody Tests						
SURE CHECK® HIV 1/2	57/75	76.7 (65.2-84.2)	1/14	7 (1-31)		
VIKIA® HIV 1/2	62/74	83.8 (73.8-90.5)	4/14	31 (13-58)		
FIRST RESPONSE® Test	57/74	77.0 (66.3-85.1)	4/14	29 (12-55)		
VIH 1-2.O CARTE						
Hexagon HIV	62/70	88.6 (79.0-94.1)	6/14	43 (21-67)		
MULTISURE HIV Rapid	57/75	76.7 (65.2-84.2)	2/14	14 (4-40)		
Test ^a						
MULTISURE HIV Rapid	56/75	74.7 (63.8-83.1)	1/14	7 (1-31)		
$Test^b$						

Results are expressed as percent (95% CI). Early seroconversion subgroup is defined as the absence of HIV-1 antibodies on the Western blot associated with both viral and p24 antigen positivity.

^a Visual reading.

^b Automatic reading.

Table 3. P values for pairwise Chi-squared tests.

	Genie TM Fast HIV 1/2	HIV 1/2 STAT-PAK® Assay	INSTI TM HIV-1/HIV-2 Antibody Tests	SURE CHECK® HIV 1/2	VIKIA® HIV 1/2	FIRST RESPONSE® Test VIH 1-2.0 CARTE	Hexagon HIV	MULTISURE HIV Rapid Test ^a	MULTISURE HIV Rapid Test ^b
Exacto® TEST HIV PRO	0.41	0.0075	0.0075	0.0075	0.12	0.011	0.48	0.0075	0.0044
Genie TM Fast HIV 1/2		0.056	0.056	0.056	0.46	0.078	0.91	0.056	0.036
HIV 1/2 STAT-PAK® Assay			1	1	0.24	0.88	0.049	1	0.85
INSTI TM HIV-1/HIV-2 Antibody				1	0.24	0.88	0.049	1	0.85
Tests									
SURE CHECK® HIV 1/2					0.24	0.88	0.049	1	0.85
VIKIA® HIV 1/2						0.3	0.41	0.24	0.17
FIRST RESPONSE® Test VIH							0.068	0.88	0.74
1-2.O CARTE									
Hexagon HIV								0.049	0.032
MULTISURE HIV Rapid Test ^a									0.85
MULTISURE HIV Rapid Test ^b									

Significant values are in bold.

^a Visual reading.

^b Automatic reading.